## Copyright

by

Liping Feng

The Dissertation Committee for Liping Feng Certifies that this is the approved version of the following dissertation:

# The Chemistry of Aza-enediynes, Aza-enyne Allenes and Related AzaBergman and Aza-Myers-Saito Rearrangements 

Committee:

Sean M. Kerwin, Supervisor

Christian Whitman

Hung-wen (Ben) Liu

Walter Fast

David W. Hoffman

The Chemistry of Aza-enediynes, Aza-enyne Allenes, and Related AzaBergman and Aza-Myers-Saito Rearrangements
by

Liping Feng, B.S.; M.S.

Dissertation<br>Presented to the Faculty of the Graduate School of The University of Texas at Austin<br>in Partial Fulfillment<br>of the Requirements<br>for the Degree of

## Doctor of Philosophy

The University of Texas at Austin
December, 2005

## Dedication

To my husband, Shiju Wang, for his endless encouragement and patience

## Acknowledgements

I would like to thank my supervising professor Dr. Kerwin for his continuous support, guidance and understanding to me during these years. The members of my dissertation committee: Drs. Liu, Whitman, Fast and Hoffman have generously given their time and expertise to better my work. I thank them for their support and contribution. I thank Dr. Shoulders and Steve Sorey for generously sharing their knowledge with me. I must acknowledge as well many friends, colleagues, teachers and librarians who assisted and supported my research over many years. Especially, I need to thank my friends Asha Nadipuram and Bodin Tuesuwan for their friendship. I would not be where I am without endless support from my family, especially my parents Guanghou Feng and Zongfang Chang, my brothers Shiping and Jianping, and my sister Xiaoping. Finally, I would like to acknowledge my husband and my best friend, Shiju for his constant encouragement.

# The Chemistry of Aza-enediynes, Aza-enyne Allenes and Related AzaBergman and Aza-Myers-Saito Rearrangements 

Publication No. $\qquad$

Liping Feng, Ph. D
The University of Texas at Austin, 2005

Supervisor: Sean M. Kerwin

Naturally occurring enediynes are a class of potent antibiotic antitumor agents that cleave DNA by generating reactive diradicals via the Bergman cyclization or MyerSaito type cyclization. However, the lack of tumor specificity of these compounds has limited their application in cancer chemotherapy. We have undertaken an approach to achieve cancer cell specific targeting by enediynes involving the re-design of the enediyne core. Classes of $C, N$-dialkynyl imines (3-ene-3-aza-1,5-diynes, aza-enediynes) and 3-ene-4-aza-1,6-diynes (skipped aza-enediynes) have been designed and synthesized. These aza-enediynes undergo a facile aza-Bergman cyclization to generate 2,5didehydropyridine diradicals that undergo a rapid retro-aza-Bergman rearrangement to ( $Z$ )- $\beta$-alkynyl acrylonitriles. Certain aza-enediynes can undergo reaction under acidic conditions to afford carbene intermediates which can be trapped efficiently. On the other hand, aza-enediynes that undergo aza-Bergman rearrangement slowly can convert to enediynes through a dimerization mechanism. The skipped aza-enediynes can isomerize to the corresponding $C$-alkynyl- $N$-allenyl imines (aza-enyne allenes) under basic
conditions. The resulting aza-enyne allenes undergo a facile aza-Myers-Saito cyclization to generate previously un-reported $\alpha, 5$-didehydro-3-picoline diradicals that can be efficiently trapped. Both 2,5-didehydropyridine diradicals and $\alpha, 5$-didehydro-3-picoline diradicals have the potential to cleave DNA. DNA cleavage abilities of these azaenediynes and skipped aza-enediynes are under investigation.

## Table of Contents

Chapter 1 Enediynes .....  1
1.1 Naturally occurring enediynes .....  1
1.1.1 Nine-membered ring naturally occurring enediynes. .....  1
1.1.2 Ten-membered ring naturally occurring enediynes .....  3
1.2 Proposed mechanisms of the action of naturally occurring enediynes .....  4
1.3 Limitations of naturally occurring enediynes in cancer chemotherapy and strategies to overcome these limitations .....  6
1.4 Bergman cyclization .....  7
1.4.1 The factors affecting Bergman cyclization ..... 7
1.4.2 Reactivity of 1,4-didehydrobenzene diradical intermediates (1,4-ddb) ..... 12
1.5 Myers-Saito cyclization and Schmittel cyclization. ..... 13
1.6 Aza-enediyne and aza-Bergman cyclization. ..... 17
1.7 Heteroatom-substituted enyne allenes ..... 20
1.7.1 Enyne-ketenes and Moore cyclization ..... 21
1.7.2 Enyne ketenimines and enyne carbodiimides ..... 21
1.7.3 Enyne-isocyanate ..... 22
1.8 Skipped aza-enediyne (3-ene-4-aza-1,6-diyne): a potential route to aza- enyne allene ..... 23
1.9 Dissertation outline ..... 25
Reference ..... 26
Chapter 2 An extremely facile aza-Bergman rearrangement of sterically unencumbered acyclic 3-aza-3-ene-1,5-diynes. ..... 31
2.1 Introduction ..... 31
2.2 Experiment design ..... 31
2.3 Synthesis of a series of acyclic aza-enediynes ..... 32
2.4 Aza-Bergman rearrangement ..... 36
2.4.1 Aza-Bergman kinetics ..... 36
2.4.2 Substituent effects ..... 43
2.4.3 Activation energy for the aza-Bergman cyclization ..... 47
2.4.4 Stereochemical aspects of aza-Bergman cyclization ..... 48
2.4.5 Solvent effects on the aza-Bergman cyclization ..... 51
2.5 Conclusions ..... 52
Reference ..... 54
Chapter 3 Attempts to trap the 2,5-didehydropyridine diradical intermediates. ..... 56
3.1 Introduction ..... 56
3.2 Attempts to methylate aza-enediyne 40a ..... 57
3.3 Isolation of a cyclopropane-containing product 44 from the rearrangement of aza-enediyne 40a under acid catalysis ..... 59
3.4 Conclusions ..... 64
Reference ..... 65
Chapter $4 \alpha, 5$-Didehydro-3-picoline diradicals from skipped aza-enediynes: computational and trapping studies of an aza-Myers-Saito cyclization ..... 66
4.1 Introduction ..... 66
4.2 A facile isomerization of the skipped aza-enediynes to aza-enyne allenes based on computational studies at B3LYP/6-31G* level ..... 68
4.3 A facile aza-Myers-Saito/aza-Schmittel cyclization of aza-enyne allenes based on computional strudies at (U)B3LYP/6-31* level ..... 71
4.4 Synthesis of acyclic skipped aza-enediyne 53e,f. ..... 73
4.5 Isomerization of skipped aza-enediyne $53 \mathrm{e}, \mathrm{f}$ to aza-enyne allene 4 d , e under basic conditions and trapping of $\alpha, 5$-didehydro-3-picoline diradicals 74
4.6 Conclusions ..... 78
Reference ..... 79
Chapter 5 Aza-enediynes undergo both aza-Bergman rearrangement to nitriles and dimerization to enediynes simultaneously ..... 81
5.1 Introduction ..... 81
5.2 Synthesis of $\left[4-{ }^{13} \mathrm{C}\right]-26 \mathrm{~d}$ ..... 82
5.3 Kinetic and mechanistic studies of the transformation of aza-enediynes 26d,e,g to enediynes 60d,e ..... 83
5.4 Conclusions ..... 90
5.5 Summary and future directions ..... 90
Reference ..... 96
Chapter 6 Experimental section ..... 98
6.1 General ..... 98
6.2 Syntheses ..... 99
6.3 Spectroscopic (UV-Vis) kinetics (40a-c) ..... 140
6.4 Spectroscopic ( ${ }^{1} \mathrm{H}$ NMR and ${ }^{13} \mathrm{C}$ NMR) kinetics (26d) ..... 141
Reference ..... 143
Appendices. ..... 144
Appendix A Energies calculated at the (U)B3LYP/6-31* level of theory . 14
Appendix B Spectroscopic kinetics for aza-enediyne 26d ..... 146
Appendix C Selected ${ }^{1} \mathrm{H}$ NMRs for compounds prepared in Experimental section ..... 150
Bibliography ..... 191
Vita ..... 199

## Chapter 1 Enediynes

### 1.1 NATURALLY OCCURRING ENEDIYNES

Naturally occurring enediynes belong to a class of very potent antitumor antibiotics. ${ }^{1,2}$ Except shishijimicins and namenamicins, which are produced by marine didemnum, ${ }^{3}$ the rest of naturally occurring enediynes discovered so far are produced by the soil bacteria. ${ }^{2}$ The structures of naturally occurring enediynes are highly conserved; they all have a conserved unsaturated core structure with two acetylenic groups conjugated to a double bond or incipient double bond, the so called enediyne core structure. These naturally occurring enediynes can be classified into two subfamilies based on the ring size of the enediyne core structure: those possessing a nine-membered ring chromophore core and those possessing a ten-membered ring chromophore core.

### 1.1.1 Nine-membered ring naturally occurring enediynes

All of nine-membered ring enediynes with an exception of N1999A2 are not stable and cannot exist without the related apo-protein which tightly, but non-covalently binds and stabilizes the enediyne chromophore. ${ }^{4}$ To date, the nine-membered ring enediyne family consists nine compounds: neocarzinostatin (NCS), kedarcidin, C-1027, maduropeptin, N1999A2, actinoxanthin, largomycin, auromomycin, and sporamycin. ${ }^{5}$ These all contain a common bicycle[7.3.0]dodecdadiynene chromophore. ${ }^{5}$ Among these nine-membered ring enediynes, there are only five whose structures have been fully established (Scheme 1-1). They are NCS, ${ }^{6}$ kedarcidin, ${ }^{7}$ C-1027, ${ }^{8 a}$ maduropeptin, ${ }^{9 a}$ N1999A2. ${ }^{10}$ These enediynes, except $\mathrm{C}-1027^{8 \mathrm{~b}}$ and maduropeptin, ${ }^{9 b}$ require a diradical initiation step which normally is thiol activation to trigger the electronic rearrangement
and diradical formation. ${ }^{2}$ Shen ${ }^{11}$ and co-workers cloned and characterized the complete polyketide synthase (PKS) gene cluster responsible for C-1027 biosynthesis from $S$. globisporus. These findings allowed Shen and co-workers to prepare a new enediyne which is analogous to $\mathrm{C}-1027$ by manipulating genes governing C-1027 biosynthesis. ${ }^{11 \mathrm{a}}$


Scheme 1-1 The structures of nine-membered ring naturally occurring enediynes

### 1.1.2 Ten-membered ring naturally occurring enediynes

Similar to nine-membered ring enediynes, ten-membered ring enediynes also contain the enediyne moiety and require a diradical initiation step (usually thiol activation). ${ }^{5}$ In contrast to nine-membered enediynes, ten-membered enediynes do not require sequestration by an apo-protein. ${ }^{2}$ To date, this subfamily includes calicheamicin, ${ }^{12}$ esperamicin, ${ }^{13}$ namenamicin, ${ }^{3}$ shishijimicin, ${ }^{3}$ and dynemicin ${ }^{14}$ (Scheme 1-2). Thorson and Farnet and co-workers ${ }^{15}$ discovered the calicheamicin gene cluster from Micromomospora ethinospora ssp. Calichensis consisting a polyketide synthase (PKS), which is a homolog of the PKS gene cluster for C-1027 biosynthesis. These findings together with the findings from Shen ${ }^{11}$ and co-workers suggest that PKS is critical for enediyne formation and all enediynes may share a common biosynthetic origin.






Scheme 1-2 The structures of ten-membered ring naturally occurring enediynes

### 1.2 PROPOSED MECHANISMS OF THE ACTION OF NATURALLY OCCURRING ENEDIYNES

In 1972, Robert Bergman ${ }^{16}$ and co-workers reported a simple enediyne $\mathbf{1}$ undergoes a cyclization (so-called Bergman cyclization) to produce 1,4didehydrobenzene (1,4-ddb) diradical 2 which then abstracts two hydrogen atoms from solvent to form benzene 3 (Scheme 1-3).


Scheme 1-3 Bergman cyclization

Later, Myers ${ }^{17}$ and Saito ${ }^{18}$ and their co-workers independently reported that enyne allene 4a can undergo a cyclization to produce $\alpha, 3$-didehydrotoluene diradical 5a•• (Scheme 1-4), which can also abstract hydrogen atoms from hydrogen donor reagents such as 1,4 -cyclohexadiene ( $1,4-\mathrm{CHD}$ ).


Scheme 1-4 Myers-Saito cyclization

Studies have shown that naturally occurring enediynes (Scheme 1-1 and Scheme 1-2) non-specifically bind to DNA minor groove, followed by tracking along the groove for a target site for specific binding. ${ }^{2}$ It is not clear whether enediynes bind to DNA before or after their thiol activation step. Once bound to DNA and activated, enediynes then either undergo reductively-induced Bergman cyclization (Scheme 1-5, path A) or a Myers-Saito type cyclization of the enediyne via enyne-cumulene (Scheme 1-5, path B),
depending on their structures, ${ }^{2}$ to produce reactive diradical intermediates, which then abstract a hydrogen atom from the DNA sugar phosphate backbone, forming a aryl radical. ${ }^{19}$ This newly formed aryl radical is more reactive than the previous diradical and rapidly abstracts another hydrogen atom from DNA. ${ }^{19,20}$ The molecular oxygen can react with the resulting DNA radicals leading to site-specific DNA breaks and ultimately cell death. ${ }^{19,21,22}$


B


Scheme 1-5 Proposed mechanisms of the action of naturally occurring enediynes (taken from Ref. 2 b ); A: Bergman cyclization type; B: Myers-Saito cyclization type (in the case of neocarzinostatin).

### 1.3 LIMITATIONS OF NATURALLY OCCURRING ENEDIYNES IN CANCER CHEMOTHERAPY AND STRATEGIES TO OVERCOME THESE LIMITATIONS

Although these naturally occurring enediynes are among the most potent anticancer antibiotics ever discovered, they cannot distinguish normal cells and cancer cells. The lack of tumor specificity causes the problem of general toxicity and subsequent systemic side effects. ${ }^{23}$ Many approaches have been employed to achieve improved
cancer-cell specific targeting by enediynes. One approach involves the creation of monoclonal antibody (mAB)-enediyne conjugates. The most successful example is antiCD33 mAB-calicheamicin, also known as gemtuzumab ozogamicin (GO), which is currently approved by the FDA under the trade name of Mylotarg to treat acute myeloid leukemia (AML). ${ }^{24 a, b} \mathrm{GO}$ is formed through a covalent linkage between a humanized anti-CD33 mAB and a semisynthetic derivative of calicheamicin. ${ }^{24 a, b}$ The anti-CD33 mAB directs the mAB-calichemicin to the CD33 cell surface glycoprotein antigen expressed by blast cells in $90 \%$ of patients with AML. ${ }^{24 c}$ Several mAB-C-1027 conjugates have been prepared and currently are being evaluated for clinical application as anticancer drugs. ${ }^{25}$ An alternative approach relies on a linkage of an enediyne-specific activation enzyme to tumor cells via a tumor specific mAB. Upon administration, the enediyne will be activated only at the tumor site by the tumor associated mAB-enzyme. ${ }^{26}$ The more general approach employs re-design of the enediyne core to achieve more selective tumor cell targeting in combination with minimum toxicity towards normal cells. ${ }^{27}$

### 1.4 BERGMAN CYCLIZATION

### 1.4.1 The factors affecting Bergman cyclization

To design new enediynes which target tumor cells with minimum toxicity towards normal cells, it is necessary to understand the factors which influence the Bergman cyclization. The Bergman cyclization is an endothermic process. The experimentally determined activation energy for cyclization of the parent enediyne ( $Z$ )-hex-3-ene-1,5diyne $\mathbf{1}$ is $28.5 \mathrm{kcal} / \mathrm{mol}$. ${ }^{28}$ Enediyne 1 cyclizes only at rather high temperature, for example, at $200{ }^{\circ} \mathrm{C}$ with a half-life 30 s . Therefore, overcoming this activation barrier is
important for re-designing enediynes. All naturally occurring enediynes mentioned in Section 1.1 are embedded into either a nine-membered or ten-membered ring system. The distance between the terminal acetylenic carbon atoms (so called c-d distance ${ }^{29 \mathrm{a}}$ ) among these enediynes decreases dramatically due to the ring strain compared to the parent $(Z)$ enediyne 1. After the thiol activation step, these enediynes cyclize spontaneously at physiological temperature. Scheme 1-6 shows the comparison of c-d distances and halflives between parent enediyne ( $Z$ )-hex-3-ene-1,5-diyne 1 and a simple 10 -membered cyclic enediyne 7. ${ }^{16,30}$ Nicolaou ${ }^{29}$ and co-workers suggested that the c-d distance should be smaller than 331 pm for enediynes to cyclize at room temperature. In contrast to $\mathrm{c}-\mathrm{d}$ distance, Magnus ${ }^{31}$ and Snyder ${ }^{32}$ and their co-workers suggested that the difference in ring strain between ground and transition state indeed accounts for the change in the energetics of the Bergman cyclization.


Scheme 1-6 Comparison of c-d distances and half-lives between acyclic enediyne $\mathbf{1}$ and cyclic enediyne 7

Enediyne-metal chelation can also alter the geometry of enediynes and lower the Bergman cyclization barriers (in some cases, electronic effects cannot be ruled out). Buchwald $^{33}$ and co-workers prepared the acyclic enediyne 8, which undergoes a facile Bergman cyclization upon chelation with $\mathrm{PdCl}_{2}$ or $\mathrm{PtCl}_{2}$ (Scheme 1-7). Enediyne 8 cyclizes at $243{ }^{\circ} \mathrm{C}$; in contrast, enediynes $\mathbf{9 a}$ and $\mathbf{9 b}$ formed upon chelation of $\mathbf{8}$ with Pd
and Pt , respectively, cyclize at relatively low temperatures: $61{ }^{\circ} \mathrm{C}$ and $81{ }^{\circ} \mathrm{C}$, respectively.


Scheme 1-7 Enediyne-metal chelation facilities Bergman cyclization

Basak ${ }^{34}$ and co-workers reported a novel diazoenediyne 11 and its macrocyclic analogue 12 (Figure 1-1) and demonstrated that Cu (II) complexation of these enediynes could promote the Bergman cyclization at reduced temperature. Zaleski ${ }^{35}$ and co-workers showed that enediyne ligands can be activated by non-hazardous $\mathrm{Mg}^{2+}$ salts.


11


Figure 1-1 Cyclization temperatures of enediynes $\mathbf{1 1}$ and $\mathbf{1 2}$ reduced upon chelation with Cu (II)

Geometric effects are not the only factor to influence the Bergman cyclization. Electronic effects, including the effects of substituents in the vinylic and terminal alkyne
position, and the influence of benzo-fusion, also, have important impacts on the Bergman cyclization of enediynes as discussed below. ${ }^{36,37,38,39,40}$

## Substitution of the terminal alkyne position

Schreiner ${ }^{36}$ and co-workers theoretically predict that strong $\sigma$-acceptors and/or $\pi$ donors such as $-\mathrm{F},-\mathrm{OH},-\mathrm{OH}_{2}^{+}$groups lower the Bergman cyclization barrier, while $\pi$ withdrawing groups such as $-\mathrm{BH}_{2},-\mathrm{AlH}_{2}$ substituents raise the barrier of the Bergman cyclization due to the interaction with $\sigma$-antibonding and $\pi$-bonding orbitals in the transition state.

Schmittel and $\mathrm{Kiau}^{37}$ confirmed this prediction by proving that replacing of the electron-donating methoxy group at alkyne position of enediyne $\mathbf{1 3}$ by electronwithdrawing nitro group (Scheme 1-8) leads to a decrease in the activation enthalpy of the Bergman cyclization.


Scheme 1-8 Nitro group facilities the Bergman cyclization of enediyne 13

## Substitution of the vinyl position

Enediyne 15a cyclizes spontaneously at room temperature (Scheme 1-9); enediyne 15b, having 4-methoxyphenyl substituent in the vinyl position, does not cyclize at temperature below $80^{\circ} \mathrm{C}$. Maier ${ }^{38}$ and Greiner suggested that the 4-methoxyphenyl substituent stabilizes the ground state more than the reaction intermediate, disfavoring the Bergman cyclization. In addition, the electron-donating aryl group may increase the repulsion in the transition state, also leading to retardation of the Bergman cyclization. Jones and Warner ${ }^{39}$ prepared a series of cyclic enediynes 16a-e with chlorine in the vinyl position (Scheme 1-9). The nine-membered enediyne 16a is extremely labile, but the chlorinated analogue $\mathbf{1 6 b}$ has a half-life of 8 h at $0^{\circ} \mathrm{C}$. The same retardation trend by addition of chlorine in the vinyl position is also found for the ten-membered enediynes 16c-e. Significantly, the addition of the second chlorine in $\mathbf{1 6 e}$ retards the Bergman cyclization rate further. ${ }^{39}$


16a $R_{1}=R_{2}=H, n=1$
16b $R_{1}=C l, R_{2}=H, n=1$
16c $R_{1}=R_{2}=H, n=2$
16d $\mathrm{R}_{1}=\mathrm{Cl}, \mathrm{R}_{2}=\mathrm{H}, \mathrm{n}=2$
$16 e R_{1}=R_{2}=C l, n=2$
Scheme 1-9

## Benzo-fusion

Benzo-fusion in the ene position of the enediynes increases the cyclization barrier compared with non-fused system. ${ }^{34 c, 40}$ Benzo-fused enediyne 7 is stable over several weeks at room temperature, in contrast, enediyne $\mathbf{1 6 c}$ has a half-life time of 18 h at $37^{\circ} \mathrm{C}$ (Scheme 1-10). Similar trend was found in enediyne 17 and its benzo-fused analogue 18. ${ }^{34 \mathrm{c}}$


16c


17


7


7 Stable at r.t.
$17 \quad 72 \mathrm{~h}$ at $23^{\circ} \mathrm{C}$


18

Scheme 1-10 Comparison of reactivity of benzo-fused and non-benzo-fused enediynes

### 1.4.2 Reactivity of $\mathbf{1 , 4}$-didehydrobenzene diradical intermediates (1,4-ddb)

In the presence of hydrogen atom source, the $1,4-\mathrm{ddb}$ diradical generated from the Bergman cyclization can either abstract two hydrogen atoms or undergo retro-Bergman cyclization (Fig 1-2). In general, the overall Bergman cyclization is first order reaction with the formation of $1,4-\mathrm{ddb}$ as the rate-limiting step. In the case of the benzannulated enediynes, hydrogen atom abstraction is the rate-determining step. ${ }^{30}$

Chen ${ }^{41 \mathrm{a}}$ and co-workers have emphasized that the reactivity of $1,4-\mathrm{ddb}$ intermediate is largely determined by the small separation between its singlet - triplet (S-
T) gap ( $3.8 \mathrm{kcal} / \mathrm{mol}$ for the enediyne 1 determined experimently ${ }^{42 \mathrm{a}}$ ). The stronger the interaction between radical centers, the larger the S-T gap is and the less reactive the diradical is towards hydrogen atoms abstraction. ${ }^{41 \mathrm{a}}$


Figure 1-2 One-dimensional reaction coordinate for the parent Bergman cyclization. Energy differences in $\mathrm{kcal} / \mathrm{mol}$ derived from experimental data (ref. 42b) and from CASSCF level (numbers in brackets, ref. 41b).

### 1.5 MYERS-SAITO CYCLIZATION AND SCHMITTEL CYCLIZATION

Myers ${ }^{17}$ and Saito ${ }^{18}$ and their co-workers reported the cyclization of $(Z)$-1,2,4-heptatrien-6-ynes (enyne-allenes, such as compound 4a) (Scheme 1-11). Enyne allene 4a undergoes $\mathrm{C}^{2}-\mathrm{C}^{7}$ cyclization to produce $\alpha, 3$-didehydrotoluene diradical which abstracts the hydrogen atoms as shown in Scheme 1-11. This pathway is similar to that found in naturally occurring enediyne neocarzinostatin, except that the $\alpha, 3$-didehydrotoluene
diradical is a $\sigma, \pi$-diradical instead of one having $\sigma, \sigma$-character in neocarzinostatin. The $\sigma, \pi$-diradical is less reactive than the $\sigma, \sigma$-diradical. ${ }^{17}$ Unlike Bergman cyclization, the Myers-Saito cyclization is exothermic by $15 \mathrm{kcal} / \mathrm{mol}$ and was found to have a half-life of 20.5 h at $39{ }^{\circ} \mathrm{C}$ for the parent enyne allene $\mathbf{4 a}$, corresponding to an activation barrier of $22 \mathrm{kcal} / \mathrm{mol} .{ }^{17 \mathrm{a}}$ Pyrolysis of compound $\mathbf{4 a}$ in methane led to the products corresponding to the both polar reaction (methyl benzyl ether, Scheme 1-11, 23) and free diradical trapping reaction ${ }^{17}$ (2-phenylethanol, Scheme 1-11, 22). Myers ${ }^{17}$ and co-workers proposed that methyl benzyl ether 23 was formed via zwitterionic intermediate 5at instead of diradical $\mathbf{5 a \cdot \bullet}$ and suggested that intermediates $\mathbf{5 a} \pm$ and $\mathbf{5 a \cdot \bullet}$ share a common rate-limiting cyclization step and may be a pair of rapidly equilibrating species. Later, Carpenter ${ }^{43}$ and Cramer ${ }^{44}$ and their co-workers pointed out that diradical 5a•• and zwitterions $\mathbf{5 a} \pm$ cannot co-exist as a pair of equilibrating species. Diradical $\mathbf{5 a \bullet \bullet}$ has A" symmetry and zwitterions $\mathbf{5 a} \pm$ has A' symmetry; mixing of these electronic configurations is therefore symmetry forbidden. Computational studies ${ }^{43}$ showed that ground state closed shell structure for $\alpha, 3$-didehydrotoluene was in fact not planar, but a puckered cyclic allene 24a, which may account for the formation of product 23. Recently, Carpenter ${ }^{45}$ and co-workers ruled out three mechanisms mentioned above, and proposed that enyne-allene $\mathbf{4 a}$ must directly form the zwitterion $\mathbf{5 a} \pm$ and exhibit a post-ratedetermining bifurcation to explain the formation of both $\mathbf{2 3}$ and trapping products $\mathbf{6}$, 19a,b. Apparently, more research needs to be done to clarify the mechanism of the Myers-Saito cyclization.

20



Scheme 1-11 Myers-Saito cyclization $\left(C^{2}-C^{7}\right)$ of enyne-allene $4 \mathbf{a}$

The initial reports on the thermolysis of enyne allenes stimulated a number of studies of the thermal cyclization of related enyne allenes to aromatic diradicals. Among these studies, Schmittel ${ }^{46}$ and co-workers and others ${ }^{47,48}$ discovered an alternate cyclization of enyne allenes - Schmittel cyclization (Scheme 1-12). Schmittel cyclization joins positions $\mathrm{C}^{2}$ and $\mathrm{C}^{6}$ to generate a fulvene-like diradical, which also has $\sigma, \pi$ character.


Scheme 1-12 Schmittel cyclization $\left(C^{2}-C^{6}\right)$ of enyne-allene $\mathbf{2 5}$

Computational studies demonstrated that Schmittel cyclization has much higher activation barriers than Myers-Saito cyclization does ( $25 \mathrm{kcal} / \mathrm{mol}$ for Myers-Saito cyclization, $35 \mathrm{kcal} / \mathrm{mol}$ for Schmittel cyclization). ${ }^{49}$ Under normal circumstances, enyne allenes favor Myers-Saito cyclization. Schmittel ${ }^{50}$ and co-workers discovered that enyne allenes may favor Schmittel cyclization when radical stabilizating substituents such as aryl or bulky alkyl groups are introduced at $\mathrm{C}^{7}$ position (Scheme 1-13). Other factors such as steric effect ${ }^{51 \mathrm{a}}$ and ring strain effect ${ }^{51 b}$ also play important roles on the relative barriers of the Myers-Saito cyclization and Schmittel cyclization.


Scheme 1-13 A switch of Myers-Saito and Schmittel cyclization by changing substituent at $\mathrm{C}^{7}$ position.

### 1.6 AZA-ENEDIYNE AND AZA-BERGMAN CYCLIZATION

Kerwin and David ${ }^{52}$ reported the first synthesis of a new class of enediynes, the 3-aza-enediynes, or C,N-dialkynyl imines 26e,f and their Bergman cyclization in 1997 (Scheme 1-14). The replacement of a $s p^{2}$ carbon with a $s p^{2}$ nitrogen would have a profound impact on facility of the Bergman cyclization due to the following reasons: the presence of nitrogen in the aza-enediyne would disturb $\pi$-delocalization compared with the enediyne, which destabilizes aza-enediyne; in addition, nitrogen also decreases repulsion of the in plane $\pi$-orbital in transition state. ${ }^{52}$ Aza-enediynes 26e,f undergo a cyclization analogous to the Bergman cyclization (aza-Bergman cyclization) to produce 2,5-didehydropyridine (2,5-ddp) diradical intermediates ${ }^{52}$ 27e,f (Scheme 1-14).


Scheme 1-14 Thermolysis of aza-enediynes 26e,f

Although these aza-enediynes undergo an aza-Bergman cyclization to produce 2,5-didehydropyridine diradical intermediates, all attempts to trap these intermediates only result in the formation of the retro-aza-Bergman rearrangement products $(Z)-\beta$ alkynyl acrylonitriles 28e, $\mathbf{f}^{52}$ (Scheme 1-14). The only experimental support for the existence of the 2,5-didehydropyridine diradical intermediate had been reported by Chen ${ }^{41 \mathrm{~b}}$ and co-workers, who reported the detection of miniscule amounts of pyridine products by GC/MS in the thermolysis of the aza-enediyne 26e under acidic conditions or upon prolonged standing in freezer.

The S-T gap of 2,5-ddp is calculated to be $8.2 \mathrm{kcal} / \mathrm{mol}$ at CASMPS level compared with $2.1 \mathrm{kcal} / \mathrm{mol}$ for $1,4-\mathrm{ddb}$ at the same level. ${ }^{41 \mathrm{~b}, 53}$ Chen ${ }^{41 \mathrm{~b}}$ and co-workers and Cramer ${ }^{53}$ proposed that the larger S-T gap of $2,5-\mathrm{ddp}$ diradical compared to $1,4-\mathrm{ddb}$ makes $2,5-\mathrm{ddp}$ less diradical in character and less reactive towards hydrogen atom abstraction reaction. In addition, 2,5-ddp diradical easily undergoes retro-aza-Bergman rearrangement to nitrile. The activation barrier for this retro rearrangement is calculated to be only $7 \mathrm{kcal} / \mathrm{mol}$, and strongly exothermic with the formation of more stable nitrile ${ }^{41 \mathrm{~b}}$ (Fig 1-3).


Figure 1-3 One dimensional reaction coordinate for the parent aza-Bergman cyclization. Energy differences in $\mathrm{kcal} / \mathrm{mol}$ at the CASSCF (ref. 41b) and BPW91 levels (numbers in brackets, ref. 53).

The difficulties encountered in trapping of 2,5-ddp diradicals are in agreement with the theoretical predictions. Protonation of nitrogen atom is predicted to reduce the ST gap of 2,5-ddp. ${ }^{4 \mathrm{~b}}$ In addition, protonation of the nitrogen atom is also predicted to increase the barrier for retro-aza-Bergman cyclization ${ }^{41 \mathrm{~b}}$ (Fig 1-4). As mentioned in section 1.3, the application of naturally occurring enediynes in chemotherapy has been limited due to their lack of cancer cell specificity and toxicity. It has been previously reported certain drugs such as amiloride, nigericin, and hydralazine preferentially lower the intracellular pH in tumor cells, with no such effect in normal cells. ${ }^{54}$ These drugs can drop the intracellular pH in tumor cells from 7.2 to between 6.2 and $6.6 .{ }^{54} \mathrm{~A}$ switch of the reactivity of $2,5-\mathrm{ddp}$ diradical upon protonation of nitrogen atom may provide the basis
for the design of novel aza-enediynes which undergo aza-Bergman cyclization and abstract hydrogen atoms from DNA only in tumor cells.


Figure 1-4 One dimensional reaction coordinate for the protonated parent aza-Bergman cyclization. Energy differences in $\mathrm{kcal} / \mathrm{mol}$ at the CASSCF (ref. 41b) and BPW91 levels (numbers in brackets, ref. 53).

### 1.7 HETEROATOM-SUBSTITUTED ENYNE ALLENES

All of the heteroatom-substituted enyne allene analogues examined to date involve heteroatom substitution on the allene moiety, including enyne ketenes, enyne ketenimines, enyne carbodiimides, and enyne-isocyanate (Fig. 1-5).


```
enyne-ketenes: X = CH, Y = O
enyne-ketenimines: X = N, Y = CR'R"
enyne-carbodiimides: X = N, Y = NR'
enyne-isocyanate: X = N, Y = O
```

Figure 1-5 Heteroatom-substituted enyne allene analogues

### 1.7.1 Enyne-ketenes and Moore cyclization

Enyne-ketenes having an oxygen atom in the allene termini undergo a cyclization analogous to Myers-Saito cyclization (also known as Moore cyclization after H. W. Moore ${ }^{55}$ who first reported the cyclization in 1985) via diradical intermediates (Scheme 1-15). Enyne-ketenes having a phenyl substituent or other radical stabilizing groups at the alkynyl terminius undergo a cyclization analogous to the Schmittel cyclization to form the fulvene-like diradicals. ${ }^{55}$


Scheme 1-15 Enyne-ketenes

### 1.7.2 Enyne ketenimines and enyne carbodiimides

Wang ${ }^{56}$ and co-workers reported that benzannulated enyne-ketenimines (Scheme $1-16, X=C R ' R '$ ') and benzannulated enyne carbodiimides (Scheme $1-16, X=N R$ ') also
undergo the $C^{2}-C^{7}$ and/or $C^{2}-C^{6}$ cyclization via diradical formation depending on the alkynyl substituents.


Scheme 1-16 Enyne ketenimines and enyne carbodiimides

### 1.7.3 Enyne-isocyanate

Enyne-isocyanates prepared by Wang ${ }^{57}$ and co-workers undergo a cyclization analogous to the Myers-Saito cyclization via diradical formation pathway (Scheme 1-17). In contrast to other hetero-enyne allene systems, such as enyne-allene, enyne-ketene, enyne-ketenimine and enyne-carbodiimide, the cycloaromatization reactions of enyneisocyanates appear to require higher temperature and are more in favor of the $\mathrm{C}^{2}-\mathrm{C}^{7}$ cyclization pathway. ${ }^{57}$


Scheme 1-17 Enyne-isocyanates

### 1.8 SKIPPED AZA-ENEDIYNE (3-ENE-4-AZA-1,6-DIYNE): A POTENTIAL ROUTE TO AZAENYNE ALLENE

$K^{K} \operatorname{Kerwin}^{58}$ and co-workers first designed the 3-ene-4-aza-1,6-diyne (skipped azaenediyne) with a methylene carbon interrupting the enediyne conjugation system (Scheme $1-18, \mathbf{A}$ ). These skipped aza-enediynes may isomerize to the corresponding $C$ -alkynyl- $N$-allenyl imines (aza-enyne allenes) under basic conditions ${ }^{58}$ (Scheme 1-18, B). The potential aza-Myers-Saito cyclization or aza-Schmittel cyclization of the resulting aza-enyne allene (Scheme 1-18, path a) may afford reactive diradical intermediates such as diradical $\mathbf{C}$ (Scheme 1-18). These diradical intermediates may cleave DNA by abstracting hydrogen atoms from DNA sugar phosphate backbone. In addition, if the nitrogen atom of these skipped aza-enediyne systems is incorporated into an electrondeficient heterocyclic ring, the alkyne moeity can serve as an electrophile which may cleave DNA as DNA, a nucleophile, attacks this electrophile forming DNA-skipped enediyne adducts D (Scheme 1-18).


Scheme 1-18 Potential reactions of skipped aza-enediyne

The heterocyclic skipped aza-enediynes such as AZB002 ${ }^{58,60}$ and AZB037 ${ }^{59}$ (Fig. 1-6) have shown the promising pH and temperature dependent DNA cleavage abilities and have the potential to function as effective antitumor agents. ${ }^{58,59,60}$ Compound AZB002 was treated with strong base in order to generate the corresponding aza-enyne allene which may undergo an aza-Myers-Saito cyclization to reactive diradical intermediates with the potential to cleave DNA. Although no trapping products of these diradical intermediates were observed, ${ }^{13} \mathrm{C}$ NMR of the crude reaction demonstrated the formation of very unstable aza-enyne allene with resonances at $\delta 198,111$, and 106 ppm corresponding to resonances of a typical allene system. ${ }^{58}$ The mechanism of DNA cleavage affected by AZB002 and AZB037 remains to be elucidated. It is possible that this cleavage is from the combination of DNA-adduct and diradical derived from an aza-Myers-Saito cyclization. ${ }^{58,59,60}$


AZB002


AZB037

Figure 1-6 Heterocyclic aza-skipped enediynes with DNA cleavage abilities

### 1.9 DISSERTATION OUTLINE

Here, I present our studies on $C, N$-dialkynyl imines (aza-enediynes) and 3-ene-4-aza-1,6-diynes (skipped aza-enediynes). Briefly, these aza-enediynes undergo a facile aza-Bergman cyclization to generate 2,5-didehydropyridine diradicals that undergo a rapid retro-aza-Bergman rearrangement to ( $Z$ )- $\beta$-alkynyl acrylonitriles (Chapter 2). Certain aza-enediynes can undergo reaction under acidic conditions to afford carbene intermediates which can be trapped efficiently (Chapter 3). On the other hand, azaenediynes that undergo aza-Bergman rearrangement slowly can undergo a remarkable transformation to produce enediynes through a dimerization mechanism (Chapter 5). Skipped aza-enediynes can isomerize to the corresponding $C$-alkynyl- $N$-allenyl imines (aza-enyne allenes) under basic conditions. The resulting aza-enyne allenes undergo a facile aza-Myers-Saito cyclization to generate previously un-reported $\alpha, 5$-didehydro-3picoline diradicals that can be efficiently trapped (Chapter 4).

## REFERENCE

1. (a) Brukner, I. Curr. Opin. Oncol. Endocr. Met. Invest. Drug 2000, 2, 344. (b) Schor, N. F. Current Therapeutics 1997, 229. (c) Thorson, J. S. Curr. Pharm. Design 2000, 6, 1841. (d). Shen, B.; Liu, W.; Nonaka, K. Curr. Med. Chem. 2003, 10, 2317.
2. (a) Xi, Z.; Goldberg, I. H. in Comprehensive Natural Products Chemistry; Elsevier: New York, 1999, vol 7, pp 533-592. (b) Cosgrove, J. P.; Dedon, P. C. in Small molecule DNA and RNA binders 2003, vol 2, pp 609-642.
3. Oku, N.; Matsunaga, S.; Fusetani, N. J. Am. Chem. Soc. 2003, 125, 2044.
4. (a) Hirama, M.; Akiyama, K.; Tanaka, T.; Noda, T.; Iida, K.; Sato, I.; Hanaishi, R.; Fukuda-Ishisaka, S.; Ishiguro, M.; Otani, T.; Leet, J. E. J. Am. Chem. Soc. 2000, 122, 720. (b) Thorson, J. S.; Shen, B.; Whitwan, R. E.; Liu, W.; Li, Y.; Ahlert, J. Bioorg. Chem. 1999, 27, 172.
5. Galm, U.; Hager, M. H.; Van Lanen, S. G.; Ju, J.; Throson, J. S.; Shen, B. Chem. Rev. 2005, 105, 739.
6. Koide, Y.; Ishii, F.; Hasuda, K.; Koyama, Y.; Edo, K.; Katamine, S.; Kitame, F.; Ishida, N. J. Antibiot. 1980, 33, 342.
7. Leet, J. E.; Schroeder, D. R.; Hofstead, S. J.; Golik, J.; Colson, K. L.; Huang, S.; Klohr, S. E.; Doyle, T. W.; Matson, J. A. J. Am. Chem. Soc. 1992, 114, 7946.
8. (a) Otani, T.; Minami, Y.; Sakawa, K.; Yoshida, K. J. Antibiot. 1991, 44, 564. (b) Xu, Y. J.; Zhen, Y. S.; Goldberg, I. H. Biochemistry 1994, 33, 5947.
9. (a) Schroeder, D. R.; Colson, K. L.; Klohr, S. E.; Zein, N.; Langley, D. R.; Lee, M. S.; Marson, J. A.; Doyle, T. W. J. Am. Chem. Soc. 1994, 116, 9351. (b) Zein, N.; Schroeder, D. R. Adv. DNA Seq.-Spec. Agents 1998, 3, 201.
10. Ando, T.; Ishii, M.; Kajiura, T.; Kameyama, T.; Miwa, K.; Sugiura, Y. Tetrahedron Lett. 1998, 39, 6495.
11. (a) Liu W.; Christenson, S. D.; Standage, S.; Shen, B. Science, 2002, 297, 1170. (b) Murrell, J. M.; Liu, W.; Shen, B. J. Nat, Prod. 2004, 67, 206.
12. Lee, M. D.; Dunne, T. S.; Siegel, M. M.; Chang, C. C.; Morton, G. O.; Borders, D. B. J. Am. Chem. Soc. 1987, 109, 3464.
13. Konishi, M.; Ohkuma, H.; Saitoh, K.; Kawaguchi, H.; Golik, J.; Dubay, G.; Groenewold, G.; Krishnan, B.; Doyle, T. W. J. Antibiot. 1985, 38, 1605.
14. Konishi, M.; Ohkuma, H.; Matsumoto, K.; Tsuno, T.; Kamei, H.; Miyaki, T.; Oki, T.; Kawaguchi, H.; Van Duyne, G. D.; Clardy, J. J. Antibiot. 1989, 42, 1449.
15. Ahlert, J.; Shepard, E.; Lomovskaya, N.; Zazopoulos, E.; Staffa, A.; Bachmann, B. O.; Huang, K.; Fonstein, L.; Czisny, A.; Whitwan, R. E.; Farnet, C. M.; Thorson, J. S. Science 2002, 297, 1173.
16. Jones, R. R.; Bergman, R. G. J. Am. Chem. Soc. 1972, 94, 660.
17. (a) Myers, A. G.; Kuo, E. Y.; Finney, N. S. J. Am. Chem. Soc. 1989, 111, 8057. (b) Myers, A. G.; Dragovich, P. S.; Kuo, E. Y. J. Am. Chem. Soc. 1992, 114, 9369.
18. (a) Nagata, R.; Yamanaka, H.; Okazaki, E.; Saito, I. Tetrahedron Lett. 1989, 30, 4995. (b) Nagata, R.; Yamanaka, H.; Murahashi, E.; Saito, I. Tetrahedron Lett. 1990, 31, 2907.
19. (a) Dedon, P. C.; Goldberg, I. H. J. Biol. Chem. 1990, 265, 14713. (b) Dedon, P. C.; Goldberg, I. H. Chem. Res. Toxicol. 1992, 5, 311. (c) Xu, Y. J.; Xi, Z.; Zhen, Y. S.; Goldberg, I. H. Biochemistry 1995, 34, 12451. (d) Rule, J. D.; Wilson, S. R.; Moore, J. S. J. Am. Chem. Soc. 2003, 125, 12992.
20. De Voss, J. J.; Townsend, C. A.; Ding, W. D.; Morton, G. O.; Ellestad, G. A.; Zein, N.; Tabor, A. B.; Schreiber, S. L. J. Am. Chem. Soc. 1990, 112, 9669.
21. Totsuka, R.; Aizawa, Y; Uesugi, M.; Okune, Y.; Matsumoto, T.; Sugiura, Y. Biochem. Biophys. Res. Commun 1995, 208, 168.
22. (a) Lopez-Larraza, D. M.; Moore, K.; Dedon, P. C. Chem. Res. Toxicol. 2001, 14, 528. (b) Wu, J. R.; Xu, J. H.; Dedon, P. C. Biochemistry 1999, 38, 15641.
23. Thorson, J. S.; Sievers, E. L.; Ahlert, J.; Shepard, E.; Whitwan, R. E.; Onwueme, K. C.; Ruppen, M. Curr. Pharm. Des. 2000, 6, 1841.
24. (a) Sievers, E. L.; Appelbaum, F. R.; Spielberger, R. T.; Forman, S. J.; Flowers, D.; Smith, F. O.; Shannon-Dorcy, K.; Berger, M. S.; Bernstein, I. D. Blood 1999, 93, 3678. (b) Giles, F. J.; Estey, E.; O’Brien, S. Cancer 2003, 98, 2095. (c) Dedon, P. C. Biol. Ther. Leukemia 2001, 2, 2.
25. Brukner, I. Curr. Opin. Oncol. Endocr. Met. Invest. Drugs 2000, 2, 344 and references therein.
26. Shabat, D.; Rader, C.; List, B.; Lerner, R. A.; Barbas, C. F.; Proc. Natl. Acad. Sci. USA 1999, 96, 6925.
27. Jones, G. B.; Fouad, F. S. Curr. Pharm. Des. 2002, 8, 2415.
28. Roth, W. R.; Hopf, H.; Horn, C. Chem. Ber. 1994, 127, 1765.
29. (a) Nicolaou, K. C.; Zuccarello, G.; Ogawa, Y.; Schweiger, E. T.; Kumazawa, T. J. Am. Chem. Soc. 1988, 110, 4866. (b) Nicolaou, K. C.; Zuccarello, G.; Riemer, C.; Estevez, V. A.; Dai, W-M. J. Am. Chem. Soc. 1992, 114, 7360. (c) Nicolaou, K. C.; Smith, A. L.; Yue, E. E. Proc. Natl. Acad. Sci. USA 1993, 90, 5881.
30. Semmelhack, M. F.; Neu, T.; Foubelo, F. J. Org. Chem. 1994, 59, 5038.
31. (a) Magnus, P.; Kewis, R. T.; Huffman, J. C. J. Am. Chem. Soc. 1988, 110, 1626. (b) Carter, P. A.; Magnus, P. J. Am. Chem. Soc. 1988, 110, 6921. (c) Magnus, P.; Fortt, S.; Pitterna, T.; Snyder, J. P. J. Am. Chem. Soc. 1990, 112, 4986.
32. (a) Snyder, J. P. J. Am. Chem. Soc. 2000, 122, 7208. (b) Snyder, J. P. J. Am. Chem. Soc. 1989, 111, 7630.
33. Warner, B. P.; Millar, S. P.; Broene, R. D.; Buchwald, S. L. Science 1995, 269, 814.
34. (a) Basak, A.; Shain, J. C. Tetrahedron Lett. 1998, 30, 3029. (b) Basak, A.; Bdour, H. M.; Shain, J. C.; Mandal, S.; Rudra, K. R., Nag, S. Bioorg. Med. Chem. Lett. 2000, 10, 1321. c) Basak, A.; Shain, J. C.; Khamrai, U. K.; Rudra, K. R. J. Chem. Soc., Perkin Trans. 1 2000, 1955.
35. Rawat, D. S.; Zaleski, J. M. J. Am. Chem. Soc. 2001, 123, 9675.
36. Prall, M.; Wittkopp, A.; Fokin, A. A.; Schreiner, P. R. J. Computational Chem. 2001, 22, 1605.
37. Schmittel, M.; Kiau, S. Chem. Lett. 1995, 953.
38. Maier, M. E.; Greiner, B. Liebigs Ann. Chem. 1992, 855.
39. (a) Jones, G. B.; Warner, P. M. J. Am. Chem. Soc. 2001, 123, 2134. (b) Jones, G. B.; Plourde, G. W. Org. Lett. 2000, 2, 1757.
40. Halter, R. J.; Fimmen, R. L.; McMahon, R. J.; Peebles, S. A.; Kuczkowski, R. L.; Stanton, J. F. J. Am. Chem. Soc. 2001, 123, 12353.
41. (a) Logan, C. F.; Chen, P. J. Am. Chem. Soc. 1996, 118, 4896. (b) Hoffer, J.; Schottelius, M. J.; Feichhtinger, D.; Chen, P. J. Am. Chem. Soc. 1998, 120, 376.
42. (a) Wenthod, P. G.; Hu, J.; Squires, R. R.; Lineberger, W. C. J. Am. Chem. Soc. 1998, 120, 5279. (b) Wenthold, P. G.; Squires, R. R. J. Am. Chem. Soc. 1994, 116, 6961.
43. Hughes, T. S.; Carpenter, B. K. J Chem. Soc., Perkin Trans. 2 1999, 2291.
44. Cramer, C. J.; Squires, R. R. Org. Lett. 1999, 1, 215.
45. Cremeens, M. E.; Hughes, T. S.; Carpenter, B. K. J. Am. Chem. Soc. 2005, 127, 6652.
46. (a) Schmittel, M.; Strittmatter, M.; Kiau, S. Tetrahedron Lett. 1995, 36, 4975. (b) Schmittel, M.; Strittmatter, M.; Vollman, K.; Kiau, S. Tetrahedron Lett. 1996, 37, 999. (c) Schmittel, M.; Strittmatter, M.; Kiau, S. Angew. Chem., Int. Ed. Engl. 1996, 25, 1843. (d) Schmittel, M.; Maywald, M.; Strittmatter, M. Synlett 1997, 165.
47. Frutos, O. D.; Echavarren, A. M. Tetrahedron Lett. 1997, 38, 7941.
48. Garcia, J. G.; Ramos, B.; Pratt, L.; Rodriguez, A. Tetrahedron Lett. 1995, 36, 7391.
49. Cramer, C. J.; Kormos, B. L.; Seierstad, M.; Sherer, E. C.; Winget, P. Org. Lett. 2001, 3, 1881.
50. Engels, B.; Lennartz, C.; Hanrath, M.; Schmittel, M.; Strittmatter, M. Angew. Chem. Int. Ed. 1998, 37, 1960.
51. (a) Schmittel, M.; Kiau, S.; Siebert, T.; Strittmatter, M. Tetrahedron Lett. 1996, 37, 7691. (b) Schmittel, M.; Steffen, J.-P.; Auer, D.; Maywald, M. Tetrahedron lett. 1997, 38, 6177.
52. David, W. M.; Kerwin, S. M. J. Am. Chem. Soc. 1997, 119, 1464.
53. Cramer, C. J. J. Am. Chem. Soc. 1998, 120, 6261.
54. (a) Raghunand, N.; Gillies, R. J. Drug Resis. Updates 2000, 3, 39 and references therein. (b) Song, C. W.; Kim, G. E.; Lyons, J. C.; Makepeace, C. M.; Griffin, R. J.; Rao, G. H.; Cragoe, E. J. Jr. Int. J. Radiat. Oncol. Biol. Phys. 1994, 30, 1161. (c)Newell, K.; Wood, P.; Stratford, I.; Tannock, I. Br. J. Cancer 1992, 66, 311. (d) Priyadarsini, K. I.; Dennis, M. F.; Naylor, M. A.; Stratford, M. R.; Wardman, P. J. Am. Chem. Soc. 1996, 118, 5648 and references therein.
55. (a) Karlsson, J. O.; Nguyen, N. V.; Foland, L. D.; Moore, H. W. J. Am. Chem. Soc. 1985, 107, 3392. (b) Moore, H. W.; Chow, K.; Nguyen, N. V.; J. Org. Chem. 1987, 52, 2530. (c) Foland, L. D.; Karlsson, J. O.; Perri, S. T.; Schwade, R.; Xu, S. L.; Patil, S.; Moore, H. W. J. Am. Chem. Soc. 1989, 111, 975.
56. (a) Shi, C.; Wang, K. K. J. Org. Chem. 1998, 63, 3517. (b) Shi, C.; Zhang, Q.; Wang, K. K. J. Org. Chem. 1999, 64, 925. (c) Li, H.; Peterson, J. L.; Wang, K. K. J. Org. Chem. 2003, 68, 5512.
57. Li, H; Yang, H.; Petersen, J. L.; Wang, K. K. J. Org. Chem. 2004, 69, 4500.
58. David, W. M.; Kumar, D.; Kerwin, S. M. Bioorg. Med. Chem. Lett. 2000, 10, 2509.
59. Tuesuwan, B; Kerwin, S. M. unpublished data.
60. Tuntiwechapikul. W; David, W. M.; Kumar, D.; Salazar, M.; Kerwin, S. M. Biochemistry 2002, 41, 5283.

## Chapter 2 An extremely facile aza-Bergman rearrangement of sterically unencumbered acyclic 3-aza-3-ene-1,5-diynes

### 2.1 Introduction

We have investigated a new class of enediynes - $C, N$-dialkynyl imines (also known as aza-enediynes, Scheme 1-11, 26e,f). ${ }^{1}$ The replacement of a $s p^{2}$ carbon with a $s p^{2}$ nitrogen is predicted to have a profound impact on facility of the Bergman cyclization. These aza-enediynes undergo aza-Bergman cyclization to produce 2,5didehydropyridine diradicals (Scheme 1-11, 27e,f) which then undergo retro-azaBergman rearrangement to $(Z)-\beta$-alkynyl acrylonitriles (Scheme 1-11, 28e,f). ${ }^{1}$ Since the initial report of the aza-enediynes and the corresponding aza-Bergman cyclization reaction, these compounds have been the subject of a number of theoretical studies. ${ }^{2,12,13,15,17}$ However, the factors that affect the kinetics of the aza-Bergman cyclization of aza-enediynes have not been elucidated. To date, there are only two reports of kinetic studies of this process, ${ }^{1,2}$ and both of these were carried out on similar azaenediynes. In this chapter, I present our kinetic studies on a series of acyclic 3-azaenediynes. ${ }^{5}$

### 2.2 EXPERIMENT DESIGN

The goals of this part of my dissertation research are to design and synthesize a series of aza-enediynes with different R substitution groups as shown in Figure 2-1 and to study the thermal rearrangements of these aza-enediynes. These aza-enediynes will be synthesized according to our previously reported procedure ${ }^{1}$ with some modification if necessary. The substituent effects on the kinetics of aza-Bergman rearrangement will be
studied. UV/Vis spectroscopy and/or ${ }^{1} \mathrm{H}$ NMR will be used to follow the reaction kinetics.


Figure 2-1 Designed aza-enediynes with different R substitution groups

### 2.3 SYNTHESIS OF A SERIES OF ACYCLIC AZA-ENEDIYNES

Scheme 2-1 shows our previous procedure ${ }^{1}$ to synthesize aza-enediynes 26e,f. (E)-oxime 30 was obtained from the addition of lithium phenylacetylide to the tertbutyldimethylsilyl nitronate ester of nitroethane 29. The oxime $\mathbf{3 0}$ was then activated to its sulfonate ester 31. Addition of the cuprate reagent ${ }^{3}$ derived from phenylacetylene or 6-phenyl-5-ene-hexyne to the sulfonate ester $\mathbf{3 1}$ afforded the $C, N$-dialkynyl imines 26e,f in modest yields (Scheme 2-1).


Scheme 2-1 Previously reported procedure for the synthesis of aza-enediynes 26e,f

Employing the same route to access other aza-enediynes was met with difficulty. ${ }^{1 b}$ Addition of lithium phenylacetylide to the silyl nitronate derived from nitromethyl benzene led to intractable mixtures from which the desired oxime could not be isolated. Condensation of 1,3-diphenylpropynone with hydroxylamine only afforded 3,5-diphenylisoxazole. ${ }^{1 \mathrm{~b}}$ To avoid the problem of isoxazole formation, we chose to prepare propynone oximes bearing a large substituent on the terminal acetylenic carbon. Thus, a series of aza-enediynes with bulky triisopropylsilyl group at $\mathrm{C}^{6}$ terminal acetylenic carbon were synthesized according to the route shown in Scheme 2-2. The first step is the cuprous iodide-catalyzed addition ${ }^{4}$ of triisopropylsilyl acetylene to benzoyl chloride, $p$-anisoyl chloride, or $o$-anisoyl chloride to afford the corresponding propynones 32a-c in good yield. The conversion of these acetylenic ketones to the corresponding oximes 33a-c proceeded in good yield, although the reactions required 10 days for completion. The oximes 33a-c were isolated as predominately single isomers of undetermined stereochemistry, as judged from ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR spectra. The oximes were then activated to their methylsulfonate esters 34a-c by the treatment with methylsulfonyl chloride. The corresponding mesylates 34a-c were also isolated as single isomers of undetermined stereochemistry. Addition of the cuprate reagent ${ }^{3}$ derived from phenylacetylene to the mesylates 34a-c afforded the $C, N$-dialkynyl imines 26a-c in modest yields ( $12 \%-31 \%$ ) along with variable amounts of reduction products 1-aryl-3-(triisopropylsilyl)-prop-2-ynylidenamines 35a-c. ${ }^{6}$ The mechanism for the formation of reduction products $\mathbf{3 5 a} \mathbf{a}$ c is not clear. It may involve water induced cleavage of the imidocopper intermediate complex. ${ }^{7,8}$ The cuprate coupling reaction is very sensitive to moisture. All glassware used for this reaction has been dried in a $160{ }^{\circ} \mathrm{C}$ oven for a period of time before use, and then transferred directly into a desiccator to cool. The azaenediynes 26a-c can be purified by flash chromatography on silica gel and isolated as
relatively stable, yellow oils that can be stored at $0^{\circ} \mathrm{C}$ for several months without any signs of decomposition. The aza-enediynes 26a-c were isolated as predominantly one isomer, as judged by ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR.



$$
\begin{aligned}
& \text { 34a } \mathrm{Ar}=\mathrm{Ph}(78 \%) \\
& \text { 34b } \mathrm{Ar}=4-(\mathrm{MeO}) \mathrm{Ph}(50 \%) \\
& \text { 34c } \mathrm{Ar}=2-(\mathrm{MeO}) \mathrm{Ph}(44 \%)
\end{aligned}
$$

$$
\text { 26a } \mathrm{Ar}=\mathrm{Ph}(26 \%)
$$

26b Ar = 4-(MeO)Ph (12\%)
26c Ar = 2-(MeO)Ph (31\%)

Scheme 2-2 Synthesis of a series of acyclic aza-enediynes 26a-b

The yields for cuprate addition were relatively poor (12-31\%). An alternative synthetic route was developed as shown in Scheme 2-3. Instead of the methylsulfonate esters 34, the propynones reacted with mesitylenesulfonyl hydroxylamine 37 to afford the mesitylenesulfonate esters 38 directly. ${ }^{9}$ There are several advantages to this approach. First of all, mesitylenesulfonyl group is a better leaving group than methylsulfonyl group; it may facilitate the cuprate addition reaction and therefore increasing the yield for this reaction. Secondly, the oxime formation step which took 10 days for completion was avoided. In addition, some propynones such as 1,3-diphenylpropynone reacted with hydroxylamine only to afford 3,5-diphenylisoxazole. ${ }^{\text {1b }}$ By switching to
mesitylenesulfonate ester, propynones were converted to mysitylenesulfonate ester directly. Mesitylenesulfonyl hydroxylamine $\mathbf{3 7}$ was synthesized according to the previously reported procedure. ${ }^{9 a}$ Briefly, condensation of 2-mesitylenesulfonyl chloride with $t$-butyl $N$-hydroxycarbamate afforded $t$-butyl $N$-mesitylenesulfonyl oxycarbamate 36 in near quantitative yield, which then underwent hydrolysis by the treatment with trifluoroacetic acid (TFA) to yield mesitylenesulfonyl hydroxylamine 37 in good yield. Compound 37 was not stable; it decomposed even when stored in a $4{ }^{\circ} \mathrm{C}$ fridge. In addition, compound $\mathbf{3 7}$ is extremely explosive; ${ }^{9 b}$ therefore, it is used immediately to react with propynones.

Mesitylate 38a,d were obtained from the condensation of the propynones 32a,d with mesitylenesulfonyl hydroxylamine $\mathbf{3 7}$ in $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ in good yield. The resulting mesitylates 38a,d were treated with cuprate reagent derived from the phenylactylene or 4-methylphenylactylene to afford the corresponding aza-enediynes 26a,d,g (Scheme 2-3) in good yields ( $>50 \%$ ). The yields for the cuprate addition were improved from 12-31\% to $55 \%$. Aza-enediynes 26a,d,g were isolated as one isomer of undetermined stereochemistry, as judged by ${ }^{1} \mathrm{H}$ NMR and ${ }^{13} \mathrm{C}$ NMR. Unlike the cuprate addition to mesylates 34a, no reduction by-product 1-phenyl-3-(triisopropylsilyl)-prop-2ynylidenamine 35a was isolated from cuprate reaction employing 38a. Further studies revealed that cuprate addition reactions carried out on more than 300 mg of mesitylates gave lower yields (less than 50\%).


Scheme 2-3 Optimized synthetic route for aza-enediynes 26a,d,g

### 2.4 AZA-BERGMAN REARRANGEMENT

### 2.4.1 Aza-Bergman kinetics

David and Kerwin ${ }^{1}$ reported that aza-enediynes 26e,f undergo aza-Bergman-retro-aza-Bergman rearrangement to the corresponding nitriles under relatively mild condition. The thermolysis of aza-enediyne 26e follows first order kinetics and has a half-life of 47 min at $110{ }^{\circ} \mathrm{C}$ in $\mathrm{CH}_{3} \mathrm{CN}$. In contrast, the half-life for the Bergman cyclization of the corresponding (Z)-1,6-diphenylhex-3-ene-1,5-diyne is reported to be 71 min at $280{ }^{\circ} \mathrm{C} .{ }^{10}$

Thermolysis of 26a was performed by heating compound 26a in chlorobenzene in the presence of 20 equiv of 1,4 -cyclohexadiene ( $1,4-\mathrm{CHD}$ ) as proton donor in a sealed tube at $150{ }^{\circ} \mathrm{C}$. The disappearance of starting material was monitored by TLC, which demonstrated that the reaction required 3 days to reach completion. We were unable to identify the product derived from trapping of the intermediate 2,5-didehydropyridine
diradical from the reaction mixture; however, the retro-Bergman cyclization product, 2,5-diphenyl-3-triisopropylsilyl-pent-2-en-4-ynenitrile 39 (Scheme 2-4), was isolated in 35\% yield.


Scheme 2-4 Thermolysis of aza-enediyne 26a

Initial attempts to deprotect aza-enediyne 26a by treatment with TBAF in THF afforded only the rearranged nitrile 28a as the sole product in $89 \%$ yield (Scheme 2-5).


Scheme 2-5 Deprotection of aza-enediynes 26a-c.

Despite the inability to isolate and purify the intermediate deprotected azaenediyne 40a, rapid workup of the deprotection reactions at low temperature followed by ${ }^{1} \mathrm{H}$ NMR of the unpurified reaction product in $\mathrm{CDCl}_{3}$ demonstrated the presence of the deprotected aza-enediyne 40a (Figure 2-2). The terminal alkyne resonance for 40a at 3.9
ppm diminished with time and was replaced by a resonance for the alkene proton at 6.8 ppm as 40a underwent conversion to the nitrile 28a over a period of minutes (Figure 2-2). The relative concentrations of aza-enediyne 40a was calculated by normalizing the integrals of the alkyne resonance for $\mathbf{4 0 a}$ at 3.9 ppm obtained at different time intervals against the integral of the alkyne resonance for $\mathbf{4 0 a}$ at 3.9 ppm obtained at reaction time equals zero minute. Based on the first-order kinetic equation (eq 2-2), the first-order kinetics for the conversion of 40a to 28a was obtained by plotting the relative concentration of $\mathbf{2 8 a}$ vs reaction time (Figure 2-4, A). The conversion of 40a to 28a determined by ${ }^{1} \mathrm{H}$ NMR follows first-order kinetics for both the disappearance of 40 a and the appearance of 28a. At $20^{\circ} \mathrm{C}$, the ${ }^{1} \mathrm{H}$ NMR-determined half-life for the conversion of 40a to 28a is $79 \pm 4 \mathrm{~min}$ as an average of two runs (Figure 2-4, A).
$-\mathrm{d}[$ reactant $] / \mathrm{dt}=\mathrm{d}[$ product $] / \mathrm{dt}=\mathrm{k}[$ reactant $] \quad(\mathrm{eq} \mathrm{2-1})$
[reactant], [product]: concentrations of reactant and product, respectively;
k : first-order kinetic rate constant; t : reaction time.

Integrating eq 2-1 to yield eq 2-2:
$\ln [$ reactant $]=-k t+\ln [\text { reactant }]_{\circ} \quad($ eq $2-2)$
[reactant $]_{0}$ : the initial reactant concentration


Figure 2-2 The conversion of aza-enediyne 40a (alkyne resonance at 3.9 pm ) to nitrile 28a (alkene resonance at 6.8 ppm ) followed by ${ }^{1} \mathrm{H}$ NMR ( 500 MHz ) in $\mathrm{CDCl}_{3}$ at $20{ }^{\circ} \mathrm{C}$. Spectra were acquired every 1010 sec , starting with the bottom spectrum.

The conversion of the aza-enediyne 40a to nitrile 28a could also be followed by UV spectroscopy. Nitrile 28a has a UV absorbance peak at 345 nm and the aza-enediyne 40a has an absorbance peak at 390 nm . Dilute $(\mu \mathrm{M})$ solutions of the product mixture from the deprotection of $\mathbf{2 6 a}$ in $\mathrm{CHCl}_{3}$ were quickly prepared and placed in temperaturecontrolled cuvettes to monitor the UV absorbance change with time. Figure 2-3 shows the conversion of aza-enediyne 40a to nitrile 28a followed by UV-Vis spectroscopy in $\mathrm{CHCl}_{3}$ at $45{ }^{\circ} \mathrm{C}$; the spectra were taken every 1.8 min for 20 min . The arrows indicate the decrease in absorbance due to the aza-enediyne 40a at 390 nm and the increase in absorbance due to the nitrile $28 a$ at 345 nm with time. The absorbance spectra demonstrate isobestic behavior and the expected first-order kinetics.


Figure 2-3 The conversion of aza-enediyne 40a to nitrile 28a followed by UV-absorption spectroscopy in $\mathrm{CHCl}_{3}$ at $45^{\circ} \mathrm{C}$ (The arrows indicate the decrease in absorbance due to the aza-enediyne 40a at 390 nm and the increase in absorbance due to the nitrile 29a at 345 nm with time).

At $20{ }^{\circ} \mathrm{C}$, the half-life for the conversion of 40 a to 28a determined spectrophotometrically is $71 \pm 1 \mathrm{~min}$ as an average of two runs (Figure 2-4, B), which is in agreement with that determined by ${ }^{1} \mathrm{H}$ NMR ( $79 \pm 4 \mathrm{~min}$, Figure 2-4, A). Our results indicate that both ${ }^{1} \mathrm{H}$ NMR and UV spectroscopy are good tools to follow the conversion of aza-enediyne 40a to nitrile 28a.
${ }^{1} \mathrm{H}$ NMR data for 40 a at $20^{\circ} \mathrm{C}$


UV-Vis data for 40 a at $20^{\circ} \mathrm{C}$


Figure 2-4 The first-order kinetics of the conversion of 40a to 28a followed by ${ }^{1} \mathrm{H}$ NMR in $\mathrm{CDCl}_{3}(\mathbf{A})$ and by UV-absorption spectroscopy in $\mathrm{CHCl}_{3}(\mathbf{B})$ at $20^{\circ} \mathrm{C}$.

### 2.4.2 Substituent effects

The substituent effects that influence the kinetics of the aza-Bergman cyclization of aza-enediynes have not been elucidated. Aza-enediyne 26a is relatively stable and undergoes aza-Bergman cyclization at rather forcing conditions ( $150{ }^{\circ} \mathrm{C}$ for 3 d ); however, removal of the bulky TIPS group from 26a to give 40a results in a facile azaBergman cyclization spontaneously even at $-10^{\circ} \mathrm{C}$. Our kinetic data shows that 40 a is a good model for kinetic study since the conversion of aza-enediyne 40a to nitrile 28a can be easily followed by ${ }^{1} \mathrm{H}$ NMR and UV spectroscopy. Therefore, $p$-(methoxy)phenylsubstituted aza-enediyne 40b and o-(methoxy)phenyl-substituted aza-enediyne 40c were prepared to study the substituent effects on the kinetics of the aza-Bergman cyclization.

A combination of ${ }^{1} \mathrm{H}$ NMR and UV-Vis were used to follow the conversion of 40b to 28b at $20{ }^{\circ} \mathrm{C}$. The time-dependent ${ }^{1} \mathrm{H}$ NMR spectra of crude $\mathbf{4 0 b}$ in $\mathrm{CDCl}_{3}$ demonstrates the same features noted above for 40a, namely, the first-order disappearance of a peak at 3.9 ppm corresponding to the terminal alkyne proton in $\mathbf{4 0 b}$ and the appearance of a new peak at 6.9 ppm , corresponding to the alkene proton in $\mathbf{2 8 b}$. In addition, in the case of $\mathbf{4 0 b}$, the methoxy protons' resonance for the aza-enediyne (3.86 ppm ) is well resolved from the methoxy protons' resonance for the nitrile 28b (3.82 ppm), and these signals also undergo a first-order decay and increase, respectively, with a kinetic yield of $>99 \%$. The time-dependent UV absorbance spectra of dilute solutions of 40b in $\mathrm{CHCl}_{3}$ are characterized by a decrease in a peak at 415 nm , corresponding to the aza-enediyne, and an increase in a peak at 360 nm , corresponding to the nitrile $\mathbf{2 8 b}$.

The conversion of the $o$-(methoxy)phenyl-substituted aza-enediyne 40c to nitrile 28c can also be followed by ${ }^{1} \mathrm{H}$ NMR spectroscopy in $\mathrm{CDCl}_{3}$. The overlap of the ${ }^{1} \mathrm{H}$ NMR signals due to the terminal alkyne proton and methoxy group of 40c and the methoxy group of 28c preclude the use of these signals to accurately follow the course of
the reaction. Instead, a distinctive aromatic ${ }^{1} \mathrm{H}$ NMR resonance for $\mathbf{4 0 c}$ at 7.9 ppm can be used to determine a first-order half-life of 35 min at $20^{\circ} \mathrm{C}$ (one run). The UV absorbance changes accompanying the conversion of $40 \mathbf{c}$ to $\mathbf{2 8 c}$ in $\mathrm{CHCl}_{3}$ are similar to those described above for the para-isomer 40b, and give a first-order half-life of $36 \pm 1 \mathrm{~min}$ at $20^{\circ} \mathrm{C}$ (for two runs). Table 2-1 summarizes the half-lives for the aza-Bergman cyclization of aza-enediynes 40a-c and 26e.

Table 2-1 Half-lives ( $\mathrm{t}_{1 / 2}$ ) for the aza-Bergman cyclization of aza-enediynes 26e and 40a,b,c


| Aza-enediyne | Temperature $\left({ }^{\circ} \mathrm{C}\right)$ | $\mathrm{t}_{1 / 2}(\mathrm{~min})$ |
| :---: | :---: | :---: |
| $\mathbf{4 0 a}$ | 0 | $667^{\mathrm{a}}$ |
| 40a | 10 | $276 \pm 13^{\mathrm{b}}$ |
| 40a | 20 | $75 \pm 6^{\mathrm{c}}$ |
| 40a | 25 | $40.6 \pm 0.1^{\mathrm{d}}$ |
| 40a | 30 | $21.5 \pm 0.1^{\mathrm{d}}$ |
| 40a | 37 | $10.4 \pm 0.1^{\mathrm{d}}$ |
| 40a | 45 | $4.3 \pm 0.1^{\mathrm{d}}$ |
| 40b | 20 | $77 \pm 3^{\mathrm{e}}$ |
| 40c | 20 | $35.6 \pm 0.5^{\mathrm{e}}$ |
| 26e | 110 | $47^{\mathrm{f}}$ |

[^0]As shown in Table 2-1, at $20^{\circ} \mathrm{C}$, the first-order half-life for the conversion of $\mathbf{4 0 b}$ to $\mathbf{2 8 b}$ is $77 \pm 3 \mathrm{~min}$; the first-order half-life for the conversion of $\mathbf{4 0 a}$ to $\mathbf{2 8 a}$ is $75 \pm 6$ min. Thus, there is no apparent electronic substituent effect on the rate of the azaBergman cyclization when comparing 40a and 40b. This result is in contrast to a number of reports of substituent effects in the Bergman cyclization. Maier and Greiner ${ }^{11}$ have reported a pronounced retardation of the Bergman cyclization of the bicyclo[7.3.1]enediyne 15b (Scheme 1-9) when the double bond is substituted with an electron-donating group ( $p-\mathrm{C}_{6} \mathrm{H}_{4} \mathrm{OMe}$ ). Jones and Plourde have reported the retardation of the Bergman cyclization of a series of cyclic enediynes 16a-e (Scheme 1-9) when the double bond is substituted with chlorine. On the other hand, calculations of a variety of aza-enediynes in which the imine double bond is replaced with an amide, amidine, or amidinium group predict that the barrier to aza-Bergman cyclization is relatively insensitive to these changes. ${ }^{12,13}$ The lack of an observable difference in the rate of azaBergman reaction of $\mathbf{4 0 a}$ and $\mathbf{4 0 b}$ is not surprising in light of these theoretical studies.

The first-order half-life for the conversion of $\mathbf{4 0 c}$ to $\mathbf{2 8 c}$ is $35.6 \pm 0.5 \mathrm{~min}$ at 20 ${ }^{\circ} \mathrm{C}$. Thus, the aza-Bergman reaction of 40c at $20{ }^{\circ} \mathrm{C}$ occurs approximately two-times faster than that of either 40a or $\mathbf{4 0 b}$ (Table 2-1). The origin of this modest substituent effect is not known; however, it is likely that the steric interaction between the $o$-methoxy group and the adjacent alkyne moiety in 40c plays a role.

The relative insensitivity of the rate of aza-Bergman cyclization to the nature of the imine double bond substituent indicates that this region of the aza-enediynes provides a great deal of flexibility in the design of potential warheads for targeting DNA and other receptors. This is significant because the imine double bond of aza-enediynes is a site of hydrolytic instability. Chen ${ }^{2}$ and co-workers note that thermolysis reactions of 26e under acidic conditions afford primarily products of imine hydrolysis. This hydrolytic
instability could be remedied by replacing the imine double bond with the amidine group as proposed by Kraka and Cremer, ${ }^{12,13}$ or by incorporating the imine double bond into a heterocyclic ring. ${ }^{14}$

In contrast to the modest substituent effects observed when comparing the azaBergman cyclization rate of $\mathbf{4 0 a} \mathbf{a} \mathbf{c}$, the nature of the alkyne substituents appears to play an important role in the rate of aza-Bergman cyclization. The aza-enediyne 26a, which bears a bulky triisopropylsilyl substituent on the C-alkynyl group, undergoes azaBergman cyclization only under rather forcing conditions $\left(150{ }^{\circ} \mathrm{C}, 3 \mathrm{~d}\right.$ for completion). Removal of the triisopropylsilyl group from 26a to give 40a results in a facile azaBergman cyclization. The phenyl-substituted aza-enediyne 26e undergoes aza-Bergman cyclization at a rate that is intermediate between that for 26a and 40a (Table 2-1). Schreiner ${ }^{15}$ and co-workers have performed calculations on substituent effects on the Bergman cyclization of ( $Z$ )-1,5-hexadiyne-3-enes. According to these calculations, 1,6-diphenyl-3-hexene-1,5-diyne, a diphenyl-substituted enediyne, has a Bergman cyclization transition state that is $10 \mathrm{kcal} / \mathrm{mol}$ higher than that for 6 -phenyl-3-hexene-1,5-diyne, mono-phenyl-substituted enediyne. The authors explained that this effect is due to a combination of additional stabilization of the starting enediyne and destabilization of the transition state due to steric repulsion between the phenyl groups in the diphenylsubstituted case. In the parent enediyne case, the effect of changing the size of the terminal substituents on the facility of Bergman cyclization cannot be employed as a triggering mechanism at physiological temperatures due to the large barrier to Bergman cyclization of the unsubstitued enediyne; however, in the case of the aza-enediynes studied here, the terminal alkyne substituent effect can be viewed as a potential triggering device due to the facility with which sterically unencumbered aza-enediynes such as 40ac undergo aza-Bergman cyclization.

### 2.4.3 Activation energy for the aza-Bergman cyclization

As shown in the Table 2-1, the half-lives of the conversion of 40a to 28a were measured at different temperatures $\left(0,10,20,25,30,37,45^{\circ} \mathrm{C}\right)$. At $10^{\circ} \mathrm{C}$, the ${ }^{1} \mathrm{H}$ NMRdetermined half-life is 4.6 h , and at $0^{\circ} \mathrm{C}$ the half-life is 11 h (Table 1 ). The half-life for the aza-Bergman reaction of 40 a at $37^{\circ} \mathrm{C}$ is only 10.4 min , as determined spectrophotometrically (Table 2-1). A plot of the logarithmic rate of disappearance of 40a versus temperature yields an Arrhenius plot for the aza-Bergman reaction of 40a to nitrile 28a, shown in Figure 2-5. Arrhenius activation parameters were extracted from the linear fit in Figure 2-5 giving $E_{\mathrm{a}}=19.9 \pm 0.3 \mathrm{kcal} / \mathrm{mol}$ and $\log A=11.1 \pm 0.2$. Chen ${ }^{2}$ and co-workers determined the activation energy for 1,6-bis(4-tert-butylphenyl)-3-aza-4-methylhex-3-ene-1,5-diyne to be $E_{\mathrm{a}}=23.1 \pm 1.5 \mathrm{kcal} / \mathrm{mol}$. The lower activation energy for the aza-Bergman reaction of 40a relative to the 1,6-diaryl aza-enediyne studied by Chen and co-workers is most likely due to the substituent effects discussed above in the comparison of the facility of cyclization of $\mathbf{4 0 a}$ versus $\mathbf{2 6 e}$. The activation energy for the aza-Bergman cyclization of 40a is lower than the barrier for aza-Bergman cyclization of the parent aza-enediyne system calculated at the CASMP2 level of theory (27.7 $\mathrm{kcal} / \mathrm{mol}),{ }^{2}$ but close to the value determined by B3LYP calculations ( $21.8 \mathrm{kcal} / \mathrm{mol}$ ), ${ }^{12,13}$ and that calculated by Cramer using a composite energy term ( $19.1 \mathrm{kcal} / \mathrm{mol}$ ). ${ }^{16,17}$


Figure 2-5 Arrhenius plot of the aza-Bergman cyclization of aza-enediyne 40a. Fifteen points at seven temperatures $\left(0,10,20,25,30,37,45^{\circ} \mathrm{C}\right)$ yield $E_{\mathrm{a}}=19.9 \pm 0.3 \mathrm{kcal} / \mathrm{mol}$ and $\log A=11.1 \pm 0.2\left(R^{2}=0.993\right)$.

### 2.4.4 Stereochemical aspects of aza-Bergman cyclization

The aza-Bergman cyclization of aza-enediynes can only occur through the $(Z)$ imine isomer. Previous studies of the aza-Bergman cyclization have employed azaenediynes 26e,f that were mixtures of stereochemical isomers about the imine double bond. ${ }^{1,2}$ As a result, the observed kinetics for the conversion of these aza-enediynes to products of aza-Bergman cyclization may have been affected by the rate of $(E)$ - to $(Z)$ imine isomerization. Aza-enediyne 26a-c and the deprotected aza-enediynes 40a-c appear to be single isomers ( $>9: 1$ ) by ${ }^{1} \mathrm{H}$ NMR. Density functional theory calculations ${ }^{18}$ at B3LYP/6-31G ${ }^{* *}$ level ${ }^{19}$ predict that the $(Z)$-isomer of 40a is more stable than the $(E)$ isomer by $4.4 \mathrm{kcal} / \mathrm{mol}$. Assuming that these aza-enediynes undergo thermodynamic equilibration during their preparation and isolation, the predominant isomers observed for 40a-c are the more thermodynamically stable $(Z)$-isomer. The nitriles produced by the
aza-Bergman-retro-aza-Bergman rearrangement can only be $(Z)$-stereoisomer. The spontaneous rearrangement of 40a-c to the nitriles 28a-c occurs stereoselectively to afford the nitrile products as single isomers as judged by ${ }^{1} \mathrm{H}$ NMR and ${ }^{13} \mathrm{C}$ NMR. Assignment of the $(Z)$-stereochemistry of these nitriles is based upon their isomerization to a mixture of $(E)$ - and ( $Z$ )-isomers (Scheme 2-6).


Scheme 2-6 Isomerization of nitrile 28 upon irradiation at 371 nm

The ${ }^{1} \mathrm{H}$ NMR spectrum of $\mathbf{2 8 a}$ in $\mathrm{CD}_{3} \mathrm{CN}$ contains a singlet resonance at 7.1 ppm , corresponding to the alkene hydrogen (Figure 2-6, A). When this solution is subjected to irradiation at 371 nm , using the excitation beam of a spectroflourimeter, ${ }^{20}$ the ${ }^{1} \mathrm{H}$ NMR spectrum shows a new single peak at 6.8 ppm , which is due to the alkene proton of the isomer of 28a (Figure 2-6, B, after 1.5 h irradiation). Because the resonance of the alkene proton of $(E)$-28a is predicted to lie 0.34 ppm upfield of that for the $(Z)$-isomer by the ChemDraw Ultra program, we assign the 7.1 -ppm peak to the $(Z)$-isomer and the $6.8-\mathrm{ppm}$ peak to the $(E)$-isomer. After 4.5 h of irradiation, $\mathbf{3 4 \%}(E)$-28a was formed based on the ratio of the peaks at 6.8 and 7.1 ppm (Figure 2-6). For conversion of aza-enediyne $\mathbf{4 0 b}$ to nitrile 28b, the ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right)$ of the product mixture prior to purification showed one singlet peak at 6.8 ppm , due to the $(Z)$-isomer of $\mathbf{2 8 b}$. Photochemical isomerization of
$\mathbf{2 8 b}$ results in the formation of an isomer with a new ${ }^{1} \mathrm{H}$ NMR resonance at 6.6 ppm , which is associated with the $(E)$-nitrile (20\%).


Figure 2-6 Isomerization of $(Z)$-nitrile 28a to its $(E)$ isomer upon irradiation at 371 nm . ${ }^{1} \mathrm{H}$ NMR of the nitrile 28a in $\mathrm{CD}_{3} \mathrm{CN}$ before irradiation (A); ${ }^{1} \mathrm{H}$ NMR of the nitrile 28a in $\mathrm{CD}_{3} \mathrm{CN}$ after irradiation at 371 nm for $1.5 \mathrm{~h}(\mathbf{B})$.

### 2.4.5 Solvent effects on the aza-Bergman cyclization

We have performed kinetic studies of 40a in various solvents to determine solvent effects on the aza-Bergman cyclization. The conversion of 40 a to 28 a at $45^{\circ} \mathrm{C}$ in various solvents was followed by UV-vis spectroscopy. Kinetic data in all solvents were firstorder and analysis of the reaction mixtures by TLC and mass spectroscopy confirmed the presence of 28a as the sole reaction product. The aza-Bergman cyclization rates were found to be solvent dependent, with slightly faster rates in nonpolar solvents (Table 2-2). The effect is not large; the half-lives varied from 3.6 min in hexanes to 8.2 min in acetonitrile. There are examples of enediynes that undergo solvent-dependent Bergman cycloaromatization rates, ${ }^{21,22}$ but these cases are complicated by the solvent effects on the rate of hydrogen atom abstraction by the diradical intermediate, which impacts the observed rate of disappearance of enediyne. In the case of the conversion of 40a to 28a, it is clear that the rate determining step is the initial aza-Bergman cyclization. We believe that the solvent-dependent rate of aza-Bergman reaction of 40a shown in Table 2-2 is due to preferential solvation of 40 a by more polar solvents, resulting in ground-state stabilization and concomitant decreased rate. Although the solvent dependence of the rate of aza-Bergman cyclization of $\mathbf{4 0 a}$ is small, the increased facility of aza-Bergman cyclization in nonpolar solvents implies that aza-enediynes such as 40a would be somewhat more stable to aza-Bergman cyclization in aqueous solution as compared to when bound to a target receptor, where the microenvironment is less polar.

Table 2-2 First-order half-lives ( $\mathrm{t}_{1 / 2}$ ) for the aza-Bergman cyclization of aza-enediyne 40a in various solvents at $45^{\circ} \mathrm{C}$

| Solvent $^{\mathrm{a}}$ | Half-life $^{\mathrm{b}}$ <br> $\left(\mathrm{t}_{1 / 2}, \mathrm{~min}\right)$ | Dielectric <br> constant (D) |
| :--- | :--- | :--- |
| Hexanes | $3.61 \pm 0.03$ | 1.89 |
| Chlorobenzene | $4.2 \pm 0.1$ | 2.71 |
| Chloroform | $4.3 \pm 0.1$ | 4.81 |
| THF | $6.4 \pm 0.2$ | 7.60 |
| Isopropyl alcohol | $7.91 \pm 0.04$ | 18.30 |
| Acetonitrile | $8.14 \pm 0.03$ | 37.50 |

${ }^{\text {a }}$ All solvents were HPLC or ACS grade and used directly without further purification. ${ }^{\mathrm{b}}$ Average value $\pm$ standard deviation for two spectrophotometric runs.

### 2.5 CONCLUSIONS

In this chapter, I report the synthesis and kinetic studies of the aza-Bergman cyclization of a series of 6-triisopropylsilyl and 6-unsubstituted 4-aryl-3-aza-3-ene-1,4diynes. There are two types of substituent effects in this system: The aza-Bergman reaction rate is not greatly affected by changing the 4 -aryl substituent on the azaenediyne imine double bond, but is very sensitive to the nature of the 6 -substitutent at the alkyne terminus. The aza-Bergman cyclization of sterically unencumbered, 6unsubstituted aza-enediynes occurs spontaneously at temperatures as low as $-10^{\circ} \mathrm{C}$. We hope that these observations provide the basis for the design of a novel aza-Bergman triggering device involving the conversion of a sterically blocked aza-enediyne to a sterically unencumbered one mediated by a cancer cell-specific enzymatic activity. One issue that remains to be addressed is the reactivity of the 2,5-didehydropyridine intermediates involved in the aza-Bergman cyclization of these aza-enediynes. In this study, there is no evidence of any products corresponding to trapping of these 2,5didehydropyridine diradical intermediates. These results are in accord with many
theoretical predictions of an extremely low barrier to retro-aza-Bergman cyclization of these intermediates to afford the observed ( $Z$ ) - $\beta$-alkynyl acrylonitrile products. ${ }^{1,12,15,23}$

## REFERENCE

1. (a) David, W. M.; Kerwin, S. M. J. Am. Chem. Soc. 1997, 119, 1464. (b) David, W. M. Ph. D Dissertation 2000, The University of Texas at Austin.
2. Hoffer, J.; Schottelius, M. J.; Feichhtinger, D.; Chen, P. J. Am. Chem. Soc. 1998, 120, 376.
3. Wurthwein, E.-U.; Weigmann, R. Angew. Chem. 1987, 99, 918.
4. Wang, J.-X.; Wei, B.; Hu, Y.; Liu, Z.; Fu, Y. Synth. Commun. 2001, 31, 3527.
5. Feng, L.; Kumar, D.; Kerwin, S. M. J. Org. Chem. 2003, 68, 2234.
6. Hagopian, R. A.; Therien, M. J.; Murdoch, J. R. J. Am. Chem. Soc. 1984, 106, 5753.
7. Tsutsui, H.; Hayashi, Y.; Narasaka, K. Chem. Lett. 1997, 317.
8. Tsutsui, H.; Ichikawa, T.; Narasaka, K. Bull. Chem. Soc. Jan. 1999, 72, 1869.
9. (a) Krause, J. G. Synthesis, 1972, 3, 140. (b) Ning, R. Y. Chem. Engineer News 1973, 51, 36.
10. Schmittel, M.; Kiau, S. Chem. Lett. 1995, 953.
11. Maier, M. E.; Greiner, B. Liebigs Ann. Chem. 1992, 855.
12. Kraka, E.; Cramer, D. J. Am. Chem. Soc. 2000, 122, 8245.
13. Kraka, E.; Cramer, D. J. Comput. Chem. 2001, 22, 216.
14. Nadipuram, A. K.; David, W. M.; Kumar, D.; Kerwin, S. M. Org. Lett. 2002, 4, 4543.
15. Prall, M.; Wittkopp, A.; Fokin, A. A.; Schreiner, P. R. J. Computational Chem. 2001, 22, 1605.
16. Cramer, C. J. J. Am. Chem. Soc. 1998, 120, 6261.
17. Debbert, S. L.; Cremer, D. J. Int. J. Mass Spectrom. 2000, 201, 1.
18. All density functional calculations were performed using GAUSSIAN98 software package: Gaussian 98, Revision A.11.4, Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Zakrzewski, V. G.; Montgomery, Jr., J. A.; Stratmann, R. E.; Burant, J. C.; Dapprich, S.; Millam, J. M.; Daniels, A. D.;

Kudin, K. N.; Strain, M. C.; Farkas, O.; Tomasi, J.; Barone, V.; Cossi, M.; Cammi, R.; Mennucci, B.; Pomelli, C.; Adamo, C.; Clifford, S.; Ochterski, J.; Petersson, G. A.; Ayala, P. Y.; Cui, Q.; Morokuma, K.; Rega, N.; Salvador, P.; Dannenberg, J. J.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Cioslowski, J.; Ortiz, J. V.; Baboul, A. G.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Gomperts, R.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Challacombe, M.; Gill, P. M. W.; Johnson, B.; Chen, W.; Wong, M. W.; Andres, J. L.; Gonzalez, C.; Head-Gordon, M.; Replogle, E. S.; Pople, J. A. Gaussian, Inc., Pittsburgh PA, 2002.
19. Becke, A. D. J. Chem. Phys. 1993, 98, 5648.
20. Yu, H.-T.; Hurley, L. H.; Kerwin, S. M. J. Am. Chem. Soc. 1996, 118, 7040.
21. Yoshida, K.; Minami, Y.; Otani, T.; Tada, Y.; Hirama, M. Tetrahedron Lett. 1994, 35, 5253.
22. Kim, C.; Russell, K. C. Tetrahedron Lett. 1999, 40, 3835.
23. Nam, H. H.; Leroi, G. E.; Harrison, J. F. J. Phys. Chem. 1991, 95, 6514.

## Chapter 3 Attempts to trap the 2,5-didehydropyridine diradical intermediates

### 3.1 INTRODUCTION

We have shown that aza-enediynes undergo a rapid aza-Bergman cyclization through a 2,5 -didehydropyridine diradical intermediate to afford the isomeric $(Z)-\beta$ alkynyl acrylonitriles. ${ }^{1,2 a}$ Unlike the Bergman cyclization of enediynes, the aza-Bergman cyclization of aza-enediynes does not afford any products corresponding to trapping of the intermediate diradicals. ${ }^{1,2 a}$ A number of computational studies of the aza-Bergman cyclization and the resulting 2,5-didehydropyridine intermediate have been carried out. ${ }^{3,4,5}$ These theoretical studies predicted that the parent 2,5-didehydropyridine diradical can undergo a rapid retro-aza-Bergman reaction to the thermodynamically stable nitrile (Figure 1-3). In addition to the small activation barrier for the nitrile formation, the 2,5ddp diradical has a large S-T gap due to the interaction between the nitrogen and the adjacent radical center. The large S-T gap and small activation barrier for the nitrile formation make the $2,5-\mathrm{ddp}$ diradical intermediate an unreactive, very short-lived species. In contrast, the protonated 2,5-ddp diradical (Figure 1-4) is predicted to be more resistance to retro-aza-Bergman rearrangement and have a smaller S-T gap, indicating that this intermediate would more readily undergo trapping reactions. ${ }^{3,4}$ These predictions have led to the proposal that these aza-enediynes might serve as pH -dependent DNA cleavage agents. Significantly, the only experimental support for the existence of the elusive 2,5-didehydropyridine intermediate has come from studies of the aza-Bergman cyclization reaction. Chen ${ }^{3}$ and co-workers reported the detection of miniscule amounts of pyridine product by GC/MS in the thermolysis of the aza-enediyne 26e under acidic conditions. We propose that alkylation of the nitrogen atom would have the same impact on the reactivity of $2,5-\mathrm{ddp}$ intermediates. In this chapter, I report our attempts to 56
protonate and methylate the nitrogen atom of aza-enediynes and our studies on the effect of added acid on the aza-Bergman cyclization and the fates of the reactive diradical intermediates from aza-Bergman cyclization under acidic conditions (Scheme 3-1). Under acidic conditions, no pyridine products were detected; rather, a cyclopropane derivative of 1,4-cyclohexadiene derived from a 5-oxazolylcarbene was isolated. ${ }^{2 b}$


Scheme 3-1 Protonation and methylation of aza-enediynes

### 3.2 ATTEMPTS TO METHYLATE AZA-ENEDIYNE 40A

In chapter 2, we $^{2 \mathrm{a}}$ report that aza-enediyne 40a undergoes a facile aza-Bergman cyclization to 2,5 -ddp diradical intermediate spontaneously even at $-10{ }^{\circ} \mathrm{C}$. We propose that methylated aza-enediyne 40a undergoes aza-Bergman cyclization to diradical intermediate which may abstract hydrogen atoms to form 1-methyl-2,5diphenylpyridinium triflate salt 43 (Scheme 3-2).


Scheme 3-2 Proposed formation of pyridinium salt $\mathbf{4 3}$ via diradical pathway

Pyridinium triflate salt $\mathbf{4 3}$ would be the trapping product of methylated 2,5-ddp diradical if it undergoes hydrogen atom abstraction reaction. Therefore, compound 43 was prepared independently as the authentic sample (Scheme 3-3). Palladium (0) catalyzed coupling of 2,5-dibromopyridine with freshly prepared phenyl zinc chloride afforded 2,5-diphenylpyridine 42 in $30 \%$ yield along with some amount of 2-phenyl-5-bromo-pyridine and biphenyl. ${ }^{6,7}$ Methyl triflate is a powerful $N$-methylating reagent. ${ }^{8}$ Diphenylpyridine $\mathbf{4 2}$ was treated with methyl triflate at $-40^{\circ} \mathrm{C}$ to give pyridinium triflate salt 43 in $75 \%$ yield.


Scheme 3-3 Synthesis of the authentic pyridinium triflate salt 43

Aza-enediyne 26a underwent desilylation reaction to afford 40a under the treatment with TBAF in THF at $-78^{\circ} \mathrm{C}$ followed by rapid aqueous work-up. The organic extracts were dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$, and passed through a plug of silica gel to remove the
tetrabutylammonium species. After evaporation of the solvent, the residue was dissolved in $\mathrm{CH}_{2} \mathrm{Cl}_{2}$. To this solution was added 8.0 equiv of methyl triflate and 20 equiv of $1,4-$ cyclohexdiene as hydrogen atom donor. The reaction mixture was warmed up to $0^{\circ} \mathrm{C}$ and kept at this temperature for 24 h . TLC of the reaction mixture and the authentic pyridinium salt $\mathbf{4 3}$ demonstrated that there was no product corresponding to the trapping of methylated 2,5 -ddp diradical intermediate. ${ }^{1} \mathrm{H}$ NMR of the reaction mixture indicated that majority was unidentified polymer. We conducted this reaction again, this time 1.2 equiv of methyl triflate was used instead of 8.0 equiv. The MS and TLC analysis of the crude reaction mixture did not show any peak corresponding to the methylated azaenediyne 40a or the trapping of methylated 2,5-ddp intermediate; instead, nitrile 28a was isolated in $35 \%$ along with other unidentified polar species, presumably polymer.

### 3.3 ISOLATION OF A CYCLOPROPANE-CONTAINING PRODUCT 44 FROM THE REARRANGEMENT OF AZA-ENEDIYNE 40A UNDER ACID CATALYSIS

Chen ${ }^{3}$ and co-workers reported the detection of miniscule amounts of pyridine product by GC/MS in the thermolysis of the aza-enediyne $\mathbf{2 6 e}$ under acidic conditions. They also found that imine double bond hydrolytic product ketone (4-phenylbut-3-yn-2one) and aza-Bergman-retro-aza-Bergman rearrangement product nitrile (phenylacetonitrile) were formed in high yield. ${ }^{3}$ Imine nitrogen is very acidic and it needs a strong acid to protonate it. We conducted the trapping experiment on aza-enediyne 26a and 40a, respectively, under the treatment with trifluoromethane sulfonic acid (TfOH). 1.0 equiv of TfOH and 20 equiv of 1,4-CHD were added to the solution of aza-enediyne 26a in $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ at $-78{ }^{\circ} \mathrm{C}$. The reaction mixture was kept at $-60^{\circ} \mathrm{C}$ for 40 min and afforded complex mixtures from which 1-phenyl-3-triisopropylsilanyl-propynone 32a was isolated in $47 \%$ along with other unidentified polar products (Scheme 3-4). No pyridine products
corresponding to trapping of $2,5-\mathrm{ddp}$ diradical were detected. It is not surprising since the imine double bond is the site of hydrolytic instability.


Scheme 3-4 Hydrolysis of imine double bond of aza-enediyne 26a under acidic condition

Desilylation of aza-enediyne 26a was carried out as described before. After aqueous work-up, the organic extracts were cooled in a dry-ice bath, and dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$. Upon removal of the solvent, the residue was dissolved in $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ and treated with catalytic TfOH ( 0.1 equiv) in the presence of 20 equiv of $1,4 \mathrm{CHD}$ at $-10^{\circ} \mathrm{C}$ for 4 days. The reaction was monitored by TLC, which demonstrated the disappearance of azaenediyne 40a and appearance of two new products, one corresponds to retro-azaBergman rearrangement product nitrile 28a and the other is a more polar unknown product 44 (Scheme 3-5).


Scheme 3-5 Formation of the cyclopropyloxazole 44 from the aza-enediyne 40a under acidic conditions

After column chromatography, compound 44 was re-crystallized in hexanes and afforded colorless lathes and needles. The unexpected structure of compound 44 has been determined by NMR and single-crystal X-ray diffraction analysis as shown in Figure 3-1.


Figure 3-1 X-Ray structure of 44 showing the atom labeling scheme. Displacement ellipsoids are scaled to the $30 \%$ probability level. The hydrogen atoms are drawn to an arbitrary size.

In order to understand the mechanism of this cyclopropyloxazole 44 formation, a set of control experiments were designed and conducted. When the reaction was carried out in the presence of stoichiometric TfOH , the aza-enediyne 40a was consumed within 30 min at $-78^{\circ} \mathrm{C}$, and the compound 44 was isolated in $30 \%$ yield along with a $15 \%$ yield of the nitrile 28a. When the reaction was conducted in the absence of TfOH , only nitrile 28a was isolated in $89 \%$ yield. When the nitrile 28a was treated with TfOH , only nitrile itself was recovered from the reaction; nitrile 28a can not be converted to cyclopropyloxazole 44 under these acidic conditions. Based on the results obtained from these reactions, we proposed that the cyclopropyloxazole 44 is the trapping product of an intermediate carbene 46 (Scheme 3-6).


Scheme 3-6 Proposed mechanism for the formation of compound 44

Although the origin of the carbene 46 is uncertain, it is possible that water molecule adds to the ynamine triple bond of the aza-enediyne 40a under the acid catalysis to afford an intermediate N -acyl- C -alkynyl imine 45. Feustel and Himbert ${ }^{9}$ observed the addition of water to ynamine triple bond to generate $N$-acyl compound under acid catalysis. The resulting imine $\mathbf{4 5}$ can be in rapid equilibrium with the thermodynamically less stable carbene species 46 which then trap 1,4-CHD by addition to its double bond to afford cyclopropyloxazole 44 . There are several experimental results supporting the carbene formation hypothesis. It is well known that 2-furylcarbenes 47 undergo ringopening rearrangement to aldehyde acetylene 48 (Scheme 3-7). ${ }^{10,11,12}$ Moreover, Schecter ${ }^{13}$ and co-workers reported that certain 5-oxazolylcarbenes 50, generated from the thermolysis or photolysis of the corresponding diazo compounds, undergo $\mathrm{C}-\mathrm{H}$ insertion and cyclopropanation reactions as well as ring-opening to $N$-acyl- C -alkynyl imines 51 (Scheme 3-7). In addition, it has been reported that several heteroatomsubstituted dieneyne system undergoes thermal cyclization to a carbene intermediate. ${ }^{14,15}$ Therefore, we propose that the formation of the carbene 46 from the $N$-acyl- C -alkynyl imine 45 followed by addition of carbene 46 to the double bond of 1,4-CHD leads to consumption of the imine $\mathbf{4 5}$ and the formation of cyclopropyloxazole 44.




Scheme 3-7

Carefully examining our trapping experiment conditions, it is not hard to find out that there may be adventitious water present in the crude desilylated aza-enediyne 40a. The role of water in the formation of compound 44 was established by carrying out the following reactions. The isolation of crude aza-enediyne 40 a was conducted under strictly anhydrous conditions; organic extracts were distilled off through a short path distillation apparatus in the presence of $4 \AA$ molecular sieves. The aza-enediyne 40a thus obtained was subjected to TfOH treatment as previously described. Under these conditions, no cyclopropane 44 was observed; instead, only nitrile 28a was isolated in nearly quantitative yield. In a separate experiment, 20 equiv of $\mathrm{H}_{2}{ }^{18} \mathrm{O}$ along with 0.1 equiv of TfOH and 20 equiv of 1,4-CHD was added into the solution of anhydrous 40a in $\mathrm{CH}_{2} \mathrm{Cl}_{2}$. Under these conditions, cyclopropyloxazole 44 was isolated in $21 \%$ yield along with $30 \%$ of nitrile 28a. The MS analysis of the compound 44 demonstrated the incorporation of ${ }^{18} \mathrm{O}$ atom into its oxazole ring. These experiments clearly establish that the oxygen atom in oxazole ring is from adventitious water presented in the crude desilylation reaction.

### 3.4 CONCLUSIONS

The isolation of the cyclopropyloxazole 44 from the aza-enediyne 40a under acidic conditions is not in agreement with the results of Chen ${ }^{3}$ and co-workers, who obtained the trapping of $2,5-\mathrm{ddp}$ intermediate product and the products corresponding to the hydrolysis of imine double bond under acidic conditions. In contrast, aza-enediyne 26a bearing a bulky TIPS group hydrolyzes to afford the hydrolysis product propynone 32a under acidic conditions. No pyridine product corresponding to trapping of 2,5-ddp diradical was detected by either ${ }^{1}$ H NMR or TLC. Aza-enediyne 40a obtained from the removal of TIPS from 26a was treated under acidic conditions; no pyridine products corresponding to trapping of $2,5-\mathrm{ddp}$ intermediate was detected by either ${ }^{1} \mathrm{H}$ NMR or TLC; rather, a cyclopropane derivative of 1,4-cyclohexadiene 44 derived from a 5oxazolylcarbene 46 was isolated. We understand that our analysis would not be able to detect the extremely low yields ( $\sim 0.05 \%$ ) of pyridine product reported by Chen. ${ }^{3}$ Here we demonstrated that certain aza-enediynes can undergo reaction under acidic conditions to afford carbene intermediates that can be efficiently trapped.

## REFERENCE

1. David, W. M.; Kerwin, S. M. J. Am. Chem. Soc. 1997, 119, 1464.
2. (a) Feng, L.; Kumar, D.; Kerwin, S. M. J. Org. Chem. 2003, 68, 2234. (b) Feng, L.; Kerwin, S. M. Tetrahedron Lett. 2003, 44, 3463.
3. Hoffer, J.; Schottelius, M. J.; Feichhtinger, D.; Chen, P. J. Am. Chem. Soc. 1998, 120, 376.
4. Cramer, C. J. J. Am. Chem. Soc. 1998, 120, 6261.
5. Kraka, E.; Cramer, D. J. Am. Chem. Soc. 2000, 122, 8245.
6. Tilley, J. W.; Zawoiski, S. J. Org. Chem. 1988, 53, 386.
7. Negishi, E.-I.; King, A. O.; Okukado, N. J. Org. Chem. 1977, 42, 1821.
8. (a) Bates, R. B.; Taylor, S. R. J. Org. Chem. 1993, 58, 4469. (b) Rahm, R; Maas, G. Chem. Ber. 1994, 127, 1295. (c) Schlegel, J. Mass, G. Synthesis 1999, 1, 100.
9. Feustel, M.; Himbert, G. Tetrahedron Lett. 1983, 24, 2165.
10. Khasanova, T.; Sheridan, R. S. J. Am. Chem. Soc. 1998, 120, 233.
11. Birney, D. M. J. Am. Chem. Soc. 2000, 122, 10917.
12. Herges, R. Angew. Chem. Int. Ed. Engl. 1994, 33, 255.
13. Hayashi, S.-I.; Nair, M.; Houser, D. J.; Shechter, H. Tetrahedron Lett. 1979, 20, 2961.
14. Kimball, D. B.; Herges, R.; Haley, M. M. J. Am. Chem. Soc. 2002, 124, 1572.
15. Frey, L. F.; Tillyer, R. D.; Ouellet, S. G.; Reamer, R. A.; Grabowski, E. J. J.; Reider, P. J. J. Am. Chem. Soc. 2000, 122, 1215.

## Chapter 4 a,5-Didehydro-3-picoline diradicals from skipped azaenediynes: computational and trapping studies of an aza-Myers-Saito cyclization

### 4.1 Introduction

The naturally occurring antitumor antibiotics neocarzinostatin (Scheme 1-2) undergoes the Myers-Saito type cyclization to produce a reactive diradical (Scheme 1-5, pathway B)..$^{1,2}$ This highly reactive diradical intermediate abstracts hydrogen atoms from the sugar phosphate backbone of DNA, leading to oxidative DNA strand scission and ultimately cell death. ${ }^{3,4,5}$ When the enyne allene functionality is not incorporated into a ring, two different cyclization pathways, Myers-Saito cyclization and Schmittel cyclization, have been demonstrated as shown in Scheme 4-1 $(\mathrm{X}=\mathrm{CH})$.


Scheme 4-1 Cyclization paths for enyne allenes and aza-enyne allenes

The Myers-Saito pathway which generates an $\alpha, 3$-didehydrotoluene diradical is similar to that found in neocarzinostatin. The Schmittel pathway generates a fulvene-like
diradical. Both $\alpha, 3$-didehydrotoluene diradical and fulvene-like diradical are $\sigma, \pi$ diradicals unlike the $\sigma, \sigma$-diradicals generated from neocarzinostatin. ${ }^{6,7}$ The Myers-Saito and Schmittel cyclizations have drawn lot of attention due to the involvement of these cyclizations in the DNA cleavage mechanism of natural and designed compounds and the potential synthetic usefulness of these cyclizations, particularly when coupled to free radical cyclization cascades.

A number of heteroatom-substituted enyne allenes have been reported. ${ }^{8-11}$ As mentioned in section 1.7, all of the heteroatom-substituted enyne allene analogues examined to date involve heteroatom substitution on the allene moiety, including enyne ketenes, ${ }^{8}$ enyne ketenimines, ${ }^{9}$ enyne carbodiimides, ${ }^{10}$ and enyne-isocyanate ${ }^{11}$ (Figure 15). These heteroatom-substituted enyne allene analogues undergo cyclizations analogous to the Myers-Saito and/or Schmittel cyclization of enyne allenes to generate diradical or zwitterion intermediates. We propose that the replacement of a $s p^{2}$ carbon with a $s p^{2}$ nitrogen to afford such previously unreported aza-enyne allenes, or $C$-alkynyl- N -allenyl imines (Scheme 4-1, 4b) will have a profound impact on the facility of these rearrangements. These aza-enyne allenes may have the potential to undergo cyclizations analogous to the Myers-Saito and Schmittel cyclization to produce the previously unreported diradical or zwitterions intermediates.

The skipped aza-enediynes (Scheme 4-2, 53) in which a single methylene group interrupts the conjugation of the aza-enediyne system have the potential to isomerize to the corresponding aza-enyne allenes (Scheme 4-2, 4) under basic conditions. ${ }^{18}$ Here I report our computational and experimental studies of the thermal rearrangements of azaenyne allenes derived from isomerization of skipped aza-enediynes (Scheme 4-2). ${ }^{21}$


53b $R_{1}=R_{2}=R_{3}=H$
53c $R_{1}=R_{3}=H, R_{2}=M e$
53d $\mathrm{R}_{1}=\mathrm{Ph}, \mathrm{R}_{2}=\mathrm{R}_{3}=\mathrm{H}$
53e $R_{1}=R_{2}=H, R_{3}=P h$

4b $\mathrm{R}_{1}=\mathrm{R}_{2}=\mathrm{R}_{3}=\mathrm{H}$
4c $R_{1}=R_{3}=H, R_{2}=M e$
4d $\mathrm{R}_{1}=\mathrm{Ph}, \mathrm{R}_{2}=\mathrm{R}_{3}=\mathrm{H}$
4e $\mathrm{R}_{1}=\mathrm{R}_{2}=\mathrm{H}, \mathrm{R}_{3}=\mathrm{Ph}$

Scheme 4-2 Proposed isomerization of the skipped aza-enediynes 53b-e to aza-enyne allenes 4b-e

### 4.2 A FACILE ISOMERIZATION OF THE SKIPPED AZA-ENEDIYNES TO AZA-ENYNE ALLENES BASED ON COMPUTATIONAL STUDIES AT B3LYP/6-31G* LEVEL

We conducted the computational calculations at B3LYP/6-31G* level ${ }^{12}$ on the isomerization of the skipped aza-enediynes 53b-d to aza-enyne allenes $\mathbf{4 b} \mathbf{- d}$ (Scheme 42). Each skipped aza-enediynes 53b-d and aza-enyne allenes $\mathbf{4 b} \mathbf{- d}$ can have two isomers ( $Z$ and $E$ isomer) about the imine double bond. Each isomer of $\mathbf{4 b}-\mathbf{d}$ can have two conformers (s-trans and $s$-cis) about $N-C$ single bond. The $s$-cis conformers of ( $Z$ ) $\mathbf{- 4 b}, \mathbf{c}$ or (E)-4d (Table 4-1) have the right geometry for the Myers-Saito/Schmittel cyclization.

Table 4-1 summarizes the calculated electronic energies at B3LYP/6-31G* level for the skipped aza-enediynes 53b-d and the corresponding aza-enyne allenes 4b-d. In each series, the relative energies are calculated relative to the more stable s-trans conformers of ( $Z$ )-53b,c or $s$-trans comformer of $(E)$ - $\mathbf{5 3 d}$. Our computational data show that ( $Z$ )-aza-enediynes $\mathbf{5 3 b}, \mathbf{c}$ are only less stable than the corresponding $(E)$-aza-enediyne $\mathbf{5 3 b}, \mathbf{c}$ by just 1.9 and $0.2 \mathrm{kcal} / \mathrm{mol}$, respectively. Thus, the $(Z)$-isomers of $\mathbf{5 3 b}, \mathbf{c}$ required for aza-Myers-Saito/Schmittel cyclization should be thermodynamically accessible. Our computational studies also demonstrate that the aza-enyne allenes $\mathbf{4 b}, \mathbf{c}$ are more stable than the corresponding skipped aza-enediynes $\mathbf{5 3 b}, \mathbf{c}$ by 12.0 and $7.3 \mathrm{kcal} / \mathrm{mol}$,
respectively (Table 4-1), supporting the proposed isomerization. There is only a small predicted energy difference between the less stable ( $Z$ )-aza-enyne allenes $\mathbf{4 b}, \mathbf{c}$ and the corresponding $(E)$-isomers $(0.1 \mathrm{kcal} / \mathrm{mol})$. The $s$-cis conformers of $(Z) \mathbf{- 4 b}, \mathbf{c}$ which are needed for the cyclization are only $3.5 \mathrm{kcal} / \mathrm{mol}$ less stable than the corresponding $s$-trans conformers of $(Z) \mathbf{- 4 b} \mathbf{\mathbf { c }}$. Therefore, the $s$-cis conformers of $(Z)$-aza-enyne allenes $\mathbf{4 b}, \mathbf{c}$ required for the Myers-Saito/Schmittel cyclization should be thermodynamically accessible. In the case of ketimine $\mathbf{4 d}$, the reactive $(E)$-imine isomer (corresponding to $(Z)-\mathbf{4 b}, \mathbf{c})$ should be predominate.

Table 4-1 B3LYP/6-31G* Energies for the skipped aza-enediynes 53b-d and corresponding aza-enyne allenes 4b-d

The geometries required for the aza-Myers-Saito/Schmittel cyclization:


(E)-4d, s-cis
(Z)-4b, s-cis

(Z)-4c, s-cis


| Compound | Electronic <br> energy $^{\mathrm{a}}$ <br> $($ Hartree $)$ | ZPE $^{\mathrm{b}}$ <br> $(\mathrm{kcal} / \mathrm{mol})$ | Relative <br> energy <br> $(\mathrm{kcal} / \mathrm{mol})$ |
| :--- | :--- | :---: | :---: |
| (E)-Skipped aza-enediyne 53b | -286.22186 | 54.7 | $10.1^{\mathrm{c}}$ |
| (Z)-Skipped aza-enediyne 53b | -286.21899 | 54.9 | $12.0^{\mathrm{c}}$ |
| (E)-Aza-enyne allene 4b (s-trans) | -286.23766 | 54.5 | $-0.1^{\mathrm{c}}$ |
| (E)-Aza-enyne allene 4b (s-cis) | -286.23454 | 54.7 | $-2.0^{\mathrm{c}}$ |
| (Z)-Aza-enyne allene 4b (s-trans) | -286.23795 | 54.8 | $0^{\mathrm{c}}$ |
| (Z)-Aza-enyne allene 4b (s-cis) | -286.23233 | 54.7 | $3.5^{\mathrm{c}}$ |
| (E)-Skipped aza-enediyne 53c | -325.54553 | 72.8 | $7.1^{\mathrm{d}}$ |
| (Z)-Skipped aza-enediyne 53c | -325.54547 | 73.0 | $7.3^{\mathrm{d}}$ |
| (E)-Aza-enyne allene 4c (s-trans) | -325.55668 | 72.5 | $-0.1^{\mathrm{d}}$ |
| (E)-Aza-enyne allene 4c (s-cis) | -325.55353 | 72.6 | $2.0^{\mathrm{d}}$ |
| (Z)-Aza-enyne allene 4c (s-trans) | -325.55682 | 72.8 | $0^{\mathrm{d}}$ |
| (Z)-Aza-enyne allene 4c (s-cis) | -325.55124 | 72.7 | $3.5^{\mathrm{d}}$ |
| (E)-Aza-enyne allene 4d (s-trans) | -517.30170 |  | $0^{\mathrm{e}}$ |
| (E)-Aza-enyne allene 4d (s-cis) | -517.29516 |  | $4.1^{\mathrm{e}}$ |
| (Z)-Aza-enyne allene 4d (s-trans) | -517.29068 |  | $6.9^{\mathrm{e}}$ |
| (Z)-Aza-enyne allene 4d (s-cis) | -517.28537 |  | $10.2^{\mathrm{e}}$ |

${ }^{\text {a }}$ Absolute energies in Hartree; ${ }^{\mathrm{b}}$ Zero-point energy correction in kcal/mol; ${ }^{\mathrm{c}}$ ZPE-corrected energy relative to ( $Z$ )-aza-enyne allene $\mathbf{4 b}$ ( $s$-trans) in $\mathrm{kcal} / \mathrm{mol}$; ${ }^{\mathrm{d}}$ ZPE-corrected energy relative to $(E)$-aza-enyne allene $4 \mathbf{c}$ ( $s$-trans) in $\mathrm{kcal} / \mathrm{mol}$; ${ }^{\mathrm{e}}$ Energy relative to $(E)$-azaenyne allene 4 d (s-trans) in $\mathrm{kcal} / \mathrm{mol}$.

### 4.3 A FACILE AZA-MYERS-SAITO/AZA-SCHMITTEL CYCLIZATION OF AZA-ENYNE ALLENES BASED ON COMPUTIONAL STRUDIES AT (U)B3LYP/6-31* LEVEL

The Myers-Saito/Schmittel cyclization and the related diradical intermediates have been the subjects of a number of computational studies. Studies have shown that the diradicals derived from either Myers-Saito or Schmittel cyclization have a small singlettriplet energy gap; therefore, they have high degrees of diradical character. ${ }^{13,14}$ As a consequence, single-determinant such as Hartree-Fock, SCF approaches are not accurate for this situation. Schmittel ${ }^{15}$ and co-workers carried out comparative calculations on enyne-allene cyclization at semiempirical AM1/CI level of theory. However, semiempirical theory such as AM1 does not handle singlet-triplet splitting well. ${ }^{16}$ Density functional theory (DFT) methods are quite suitable to describe the Myers-Saito and Schmittel cyclization barriers and the corresponding diradicals and are nearly as accurate as much time-consuming high-level methods such as coupled cluster $[\operatorname{CCSD}(\mathrm{T})]$ and Brueckner doubles $[\mathrm{BCCD}(\mathrm{T})] .{ }^{17}$ Schreiner and Prall ${ }^{17}$ suggested that hybrid DFT such B3LYP approach towards singlet $\alpha, 3$-didehydrotoluene diradical species suffers from artifactual symmetry breaking; Lipton and Wenthold ${ }^{19}$ discovered that symmetry breaking for the diradical under B3LYP level can be avoided by using the GUESS = ALTER command in the input. The wave function obtained from this procedure is a mixture of singlet and triplet states (spin operator is larger than one). Since singlet-triplet gap of $\alpha, 3$-didehydrotoluene is relatively small, the energy difference between this mixtured wave function and pure open-shell singlet wave function is rather small. ${ }^{19}$ In the present study, the geometries and energies for the diradicals and related species and transition states of the aza-Myers-Saito/Schmittel cyclizations are computated at hybrid density functional theory (U)B3LYP level with a basis set 6-31G*. In the case of singlet diradicals, the geometries and energies are computed by using an unrestricted B3LYP
wave function and the command GUESS = ALTER in the input. In comparison, we also compute the geometries and energies for the diradicals and related species and transition states of the Myers-Saito/Schmittel cyclizations at the same level.

Table 4-2 summarizes the relative energies of transition states, zwitterions and diradicals relative to their respective ( $Z$ )-reactants $\mathbf{4 a , b}$. The results demonstrate that azaMyers cyclization is more exothermic than Myers cyclization by about $-4 \mathrm{kcal} / \mathrm{mol}$ and it has a slightly lower barrier than that of Myers cyclization. There is a closed-shell singlet planar zwitterion $5 \pm$ which is much higher in energy than that of the corresponding openshell singlet diradical $5 \cdot \bullet$, in both the Myers-Saito and aza-Myers-Saito cases. A nonplanar closed-shell singlet cyclic-allene 24b that has lower energy than aza-zwitterion $\mathbf{5 \pm}$ was found for aza-Myers cyclization, as has been reported for Myers-Saito case. ${ }^{14,20}$ The calculation results predict a facile aza-Myers cyclization of aza-enyne allene $\mathbf{4 b}$ via either diradical pathway or ionic pathway. (U)B3LYP calculations also indicates that both transition states for aza-Myers-Saito (TSb1) and aza-Schmittel (TSb2) do not have diradical character, which is also the case for the Myers-Saito ${ }^{17,20,22}$ and Schmittel cyclization. ${ }^{17,22}$ This indicates that a crossing to the open-shell surface from the closed shell surface occurs after these transition states. Although aza-enyne allenes favor aza-Myers-Saito cyclization over aza-Schmittel cyclization, as shown in Table 4-2, the difference of the barriers between these cyclizations is less than $5 \mathrm{kcal} / \mathrm{mol}$, which indicates there might be a competition between aza-Myers cyclization and aza-Schmittel cyclization.

Table 4-2 Calculated energies ${ }^{\text {a }}$ for the Myers, Schmittel, aza-Myers and aza-Schmittel cyclizations of $\mathbf{4 a}, \mathbf{b}$ at (U)B3LYP/6-31G* level

|  | Myers Cyclization |  |  |  |  | Schmittel Cyclization |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | TS1 ${ }^{\text {+ }}$ | s5** ${ }^{\text {b }}$ | t5** ${ }^{\text {c }}$ | $5 \pm{ }^{\text {d }}$ | 24 | TS2 ${ }^{\text { }}$ | S52•* | t52•* |
| 4a | $26.3{ }^{\text {e }}$ | -11.6 | -10.2 | 20.9 | 0.1 | $34.5{ }^{\text {e }}$ | 13.1 | 10.5 |
| Exp | $23.3 \pm 0.5^{\text {f }}$ | $-13.3 \pm 4^{\text {g }}$ |  |  |  |  |  |  |
|  | Aza-Myers Cyclization |  |  |  |  | Aza-Schmittel Cyclization |  |  |
| 4b | $23.2{ }^{\text {e }}$ | -15.4 | -14.0 | 16.5 | $-2.6{ }^{\text {e }}$ | $27.7^{\text {e }}$ | 7.3 | 5.2 |

${ }^{\text {a }}$ calculated energies with zero-point correction relative to respective ( $Z$ )-trans $\mathbf{4}$ isomer in $\mathrm{kcal} / \mathrm{mol}$; ${ }^{\mathrm{b}}$ the singlet diradical; ${ }^{\mathrm{c}}$ the triplet diradical; ${ }^{\mathrm{d}}$ closed-shell singlet with $\mathrm{C}_{\mathrm{s}}$ restriction enforced; ${ }^{\mathrm{f}}$ data from ref $6 \mathrm{a} ;{ }^{\mathrm{g}}$ data from ref 14.

### 4.4 SYNTHESIS OF ACYCLIC SKIPPED AZA-ENEDIYNE 53E,F

The skipped aza-enediynes $\mathbf{5 3 e}$,f were synthesized from the condensation of 1-phenyl-3-triisopropylsilyl-propynone 32a or phenyl propynal 32g with excess amount of propargyl amine in the presence of titanium (IV) ethoxide in modest yield (Scheme 4-3). The skipped aza-enediyne $\mathbf{5 3 f}$ can be purified by flash chromatography on silica gel and isolated as relatively stable, red-yellow oil. The skipped aza-enediyne 53e can be purified by re-crystallization from hexanes as relatively stable colorless cubic crystal. Compound 53f was obtained as predominant single isomer of undetermined stereochemistry, based on ${ }^{1} \mathrm{H}$ NMR and ${ }^{13} \mathrm{C}$ NMR. Compound $\mathbf{5 3} \mathbf{e}$ was isolated as two isomers (1:3) about imine double bond as judged by ${ }^{1} \mathrm{H}$ NMR and ${ }^{13} \mathrm{C}$ NMR.


Scheme 4-3 Synthesis of acyclic skipped aza-enediyne 53e,f

### 4.5 ISOMERIZATION OF SKIPPED AZA-ENEDIYNE 53E,F TO AZA-ENYNE ALLENE 4D,E UNDER BASIC CONDITIONS AND TRAPPING OF A,5-DIDEHYDRO-3-PICOLINE DIRADICALS

Attempts to desilylate compound $\mathbf{5 3 f}$ by treatment with TBAF at $-78{ }^{\circ} \mathrm{C}$ only afforded isomerization product $\mathbf{4 d}$ as the sole product. We did not detect any desilylated skipped aza-enediyne (Scheme 4-4).

$\uparrow \begin{aligned} & \mathrm{MeOH} \\ & 0^{\circ} \mathrm{C}, 14 \mathrm{~h}\end{aligned}$
53f

$53 f R_{1}=P h, R_{2}=$ TIPS
4d
53e $\mathrm{R}_{1}=\mathrm{H}, \mathrm{R}_{2}=\mathrm{Ph}$

$10^{\circ} \mathrm{C}, 12 \mathrm{~h}$


59a,b (27\%)

$$
\begin{array}{ll}
5 \mathrm{~d} \cdot \cdot R_{1}=\mathrm{Ph}, \mathrm{R}_{2}=\mathrm{H} & 5 \mathrm{~d} \pm \mathrm{R}_{1}=\mathrm{Ph}, \mathrm{R}_{2}=\mathrm{H} \\
5 \mathrm{e} \cdot \cdot \mathrm{R}_{1}=\mathrm{H}, \mathrm{R}_{2}=\mathrm{Ph} & 5 \mathrm{e} \pm \mathrm{R}_{1}=\mathrm{H}, \mathrm{R}_{2}=\mathrm{Ph}
\end{array}
$$



Scheme 4-4 Isomerization of 53e,f under basic conditions and trapping of $\alpha, 5$-didehydropicoline diradicals

Although the instability of $\mathbf{4 d}$ prevented its chromatographic purification, its structure was confirmed by NMR. MS analysis of the reaction mixture after aqueous workup further confirms the loss of the triisopropylsilyl group. The ${ }^{1} \mathrm{H}$ NMR of $\mathbf{4 d}$ contains distinctive resonance for the allene moiety; it has a methylene doublet at 5.36 ppm and a methine triple at 7.60 ppm . Decoupled ${ }^{1} \mathrm{H}$ NMR demonstrates that methylene doublet at 5.36 ppm couples with the methine triplet at 7.60 ppm with a coupling constant
6.0 Hz. We proposed that the downfield shift of the methine triplet $(7.60 \mathrm{ppm})$ is due to the favorable ( Z )-s-trans conformation of $\mathbf{4 d}$ (Figure 4-1). In this conformation, the allene methine is proximal to and in the deshielding region of the alkyne triple bond, and the resulting deshielding shifts methine downfield. The ${ }^{13} \mathrm{C}$ NMR of reaction mixture contains expected resonance for the allene $s p$ carbon at 212 ppm .


Figure 4-1 (Z)-s-trans conformation of aza-enyne allene 4d

Solutions of aza-enyne allene $\mathbf{4 d}$ in $\mathrm{CDCl}_{3}$ are unstable, even when refrigerated, and afford complex mixtures from which aldehyde 54 could be isolated (Scheme 4-4). Solutions of $\mathbf{4 d}$ in $1,4-\mathrm{CHD}$ were allowed to stand at $4^{\circ} \mathrm{C}$. TLC demonstrated that all aza-enyne allene $4 d$ was consumed after 8 hours at $4^{\circ} \mathrm{C}$. The reaction afforded a complex mixture of products that included 6-phenyl-3-picoline $\mathbf{5 5}$ isolated in $20 \%$ yield, and a mixture of cyclohexadiene adducts 56a,b in $17 \%$ yield ( $\sim 1: 1$ ratio as judged by ${ }^{1} \mathrm{H}$ NMR, Scheme 4-4). We were unable to separate 56a and 56b by TLC under different solvent systems. Therefore, compounds 56a,b were subjected to hydrogenation to afford the reduced product 57 , which was fully characterized.

The formation of $\mathbf{5 4}, \mathbf{5 5}$, and $\mathbf{5 6 a}, \mathbf{b}$ are all commensurate with the intermediacy of the previously unreported $\alpha, 5$-didehydro-3-picoline diradical $5{ }^{-} \cdot \bullet$. Rapid cyclization of 4d to diradical 5d•• (compare Myers cyclization of $\mathbf{4 a}$ with $t_{1 / 2}$ of 20.5 h at $39{ }^{\circ} \mathrm{C}$ ),
followed by hydrogen atom abstraction from 1,4-CHD affords a radical pair that can recombine to give $\mathbf{5 6 a}, \mathbf{b}$ or undergo subsequent hydrogen atom abstraction to afford $\mathbf{5 5}$. Aldehyde 54 presumably arises from interception of adventitious oxygen by diradical $\mathbf{5 d} \cdot \bullet$. When the solution of $\mathbf{4 d}$ in $d_{8}$-THF was stored at $4{ }^{\circ} \mathrm{C}$ for 10 hour, the pyridine derivative 55 was isolated in $2 \%$ yield. ${ }^{1} \mathrm{H}$ NMR and MS analysis demonstrated that $\sim$ $33 \%$ of $\mathbf{5 5}$ have two deuteriums, which further support for the intermediacy of $\mathbf{5 d} \cdot \boldsymbol{\bullet}$. THF apparently is a poor hydrogen atom donor towards diradical $\mathbf{5 d} \cdot \bullet$. No trapping product which corresponds to the trapping of the aza-Schmittel cyclization diradical 52d•• was isolated from any of the conditions mentioned above.

The solution of the skipped aza-enediyne 53e in THF was maintained at $50^{\circ} \mathrm{C}$ for 1.5 h in the presence of 5 equiv of $\mathrm{Et}_{3} \mathrm{~N}$. The NMR analysis of crude reaction mixture demonstrated that $10 \%$ of $\mathbf{5 3 e}$ was converted to aza-enyne allene $\mathbf{4 e}$. All attempts to purify aza-enyne allene $\mathbf{4 e}$ failed due to its instability. In the presence of large excess of $1,4-\mathrm{CHD}$, the solution of $\mathbf{5 3 e}$ in THF in the presence of 5 equiv of $\mathrm{Et}_{3} \mathrm{~N}$ afforded a complex mixture of products including the cyclohexadiene adducts 59a,b in 27\% ( $\sim 1: 1$ ratio, as judged by ${ }^{1} \mathrm{H}$ NMR) (Scheme 4-4). We were unable to isolate the trapping product containing two hydrogen atoms derived from 1,4-CHD from the reaction mixture. Similar to aza-enyne allene $\mathbf{4 d}$, aza-enyne allene $\mathbf{4 e}$ undergoes a cyclization analogous to Myers-Saito cyclization to produce diradical $\mathbf{5 e \bullet \bullet}$, which abstracts hydrogen atom from 1,4-CHD to afford a radical pair that can recombine to give adducts 59a,b.

When solutions of $\mathbf{4 d}$ in methanol were stored at $0{ }^{\circ} \mathrm{C}$, 5 -methoxymethyl-2phenylpyridine 58 was isolated in $20 \%$ yield. Myers ${ }^{6 a, b}$ and co-workers reported a similar methanol addition product from the thermolyses of enyne allene $\mathbf{4 a}$ in methanol, and proposed that this is due to a partitioning between diradical and ionic reaction pathways. Recently, Carpenter ${ }^{23}$ and co-workers examined this reaction in detail by the combination
of experimental and computational means, and proposed that enyne-allene $4 \mathbf{4}$ must directly form the zwitterion $\mathbf{5 a} \pm$ and exhibit a post-rate-determining bifurcation to explain the formation of both 23 and trapping products 6, 19a,b (Scheme 1-11). Alternatively, the methoxymethyl pyridine 58 may arise from a different mechanism involving methanol addition to the aminoallene functionality of $\mathbf{4 d}$, followed by cyclization of the resulting azadienyne. ${ }^{21}$

Despite the overall similarity in trapping products from the thermal rearrangements of $\mathbf{4 a}$ and $\mathbf{4 d}$, there are differences between these two systems. In general, the isolated yields of trapped products from $\mathbf{4 d}$ are lower than those reported for $\mathbf{4 a}$. It may be that a competing aza-Schmittel pathway in the case of $\mathbf{4 d}$ leads to more complex reaction mixtures and lower isolated yields.

### 4.6 CONCLUSIONS

In conclusion, the formation of $\alpha, 5$-didehydro-3-picoline diradical intermediates from skipped aza-enediynes via isomerization to aza-enyne allenes and aza-Myers-Saito cyclization has been predicted on the basis of DFT calculations. Skipped aza-enediynes are easily prepared, and their facile aza-Myers cyclization provides a route to the previously unreported picolinyl diradicals. In addition, the isomerization of skipped azaenediynes and their subsequent rapid aza-Myers-Saito cyclization may have relevance to the recently reported DNA cleavage ability of cationic heterocyclic skipped azaenediynes such as AZB002 ${ }^{24}$ and AZB037 ${ }^{25}$ (Figure 1-6).

## REFERENCE

1. (a) Xi, Z.; Goldberg, I. H. in Comprehensive Natural Products Chemistry; Elsevier: New York, 1999, vol 7, pp 533-592. (b) Cosgrove, J. P.; Dedon, P. C. in Small molecule DNA and RNA binders 2003, vol 2, pp 609-642.
2. De Voss, J. J.; Townsend, C. A.; Ding, W. D.; Morton, G. O.; Ellestad, G. A.; Zein, N.; Tabor, A. B.; Schreiber, S. L. J. Am. Chem. Soc. 1990, 112, 9669.
3. Christner, D. F.; Frank, B. L.; Kozarich, J. W.; Stubbe, J.; Golik, J.; Doyle, T. W.; Rosenberg, I. E.; Kristnan, B. J. Am. Chem. Soc. 1992, 114, 8763.
4. Lopez-Larraza, D. M.; Moore, K.; Dedon, P. C. Chem. Res. Toxicol. 2001, 14, 528.
5. Wu, J. R.; Xu, J. H.; Dedon, P. C. Biochemistry 1999, 38, 15641.
6. (a) Myers, A. G.; Kuo, E. Y.; Finney, N. S. J. Am. Chem. Soc. 1989, 111, 8057. (b) Myers, A. G.; Dragovich P. S.; Kuo, E. Y. J. Am. Chem. Soc. 1992, 114, 9369. (c) Nagata, R.; Yamanaka H.; Okazaki, E.; Saito, I. Tetrahedron Lett. 1989, 30, 4995. (d) Nagata, R.; Yamanaka, H.; Murahashi, E.; Saito, I. Tetrahedron Lett. 1990, 31, 2907.
7. Schmittel, M.; Strittmatter, M.; Kiau, S. Angew. Chem., Int. Ed. Engl. 1996, 25, 1843.
8. Karlsson, J. O.; Nguyen, N. V.; Foland, L. D.; Moore, H. W. J. Am. Chem. Soc. 1985, 107, 3392.
9. Shi, C.; Wang, K. K. J. Org. Chem. 1998, 63, 3517.
10. Shi, C.; Zhang, Q.; Wang, K. K. J. Org. Chem. 1999, 64, 925.
11. Li, H.; Yang, H.; Peterson, J. L.; Wang, K. K. J. Org. Chem. 2004, 69, 4500.
12. All density functional calculations were performed using GAUSSIAN98 software package: Gaussian 98, Revision A.11.4, Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Zakrzewski, V. G.; Montgomery, Jr., J. A.; Stratmann, R. E.; Burant, J. C.; Dapprich, S.; Millam, J. M.; Daniels, A. D.; Kudin, K. N.; Strain, M. C.; Farkas, O.; Tomasi, J.; Barone, V.; Cossi, M.; Cammi, R.; Mennucci, B.; Pomelli, C.; Adamo, C.; Clifford, S.; Ochterski, J.; Petersson, G. A.; Ayala, P. Y.; Cui, Q.; Morokuma, K.; Rega, N.; Salvador, P.; Dannenberg, J. J.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Cioslowski, J.; Ortiz, J. V.; Baboul, A. G.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Gomperts, R.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Challacombe, M.; Gill, P. M. W.; Johnson, B.; Chen,
W.; Wong, M. W.; Andres, J. L.; Gonzalez, C.; Head-Gordon, M.; Replogle, E. S.; Pople, J. A. Gaussian, Inc., Pittsburgh PA, 2002.
13. Wenthold, P. G.; Squires, R. R.; Lineberger, W. C. J. Am. Chem. Soc. 1998, 120, 5279.
14. Wenthold, P. G.; Wierschke, S. G.; Nash, J. J.; Squires, R. R. J. Am. Chem. Soc. 1994, 116, 7378.
15. Schmittel, M.; Keller, M.; Kiau, S.; Strittmatter, M. Chem. Eur. J. 1997, 3, 807.
16. Engels, B.; Hanrath, M. J. Am. Chem. Soc. 1998, 120, 6356.
17. Schreiner, P. R.; Prall, M. J. Am. Chem. Soc. 1999, 121, 8615.
18. Krause, N.; Hashmi, A. S. K. Modern Allene Chemistry, chapter one, 2004, Wiley.
19. Wenthold, P. G.; Lipton, M. K. J. Am. Chem. Soc. 2000, 122, 9265.
20. Hughes, T. S.; Carpenter, B. K. J. Chem. Soc., Perkin Trans. 2 1999, 2291.
21. Feng, L.; Kumar, D.; Birney, D. M.; Kerwin, S. M. Org. Lett. 2004, 6, 2059.
22. (a) Musch, P.W.; Remenyi, C.; Helten, H.; Engels, B. J. Am.Chem. Soc. 2002, 124, 1823. (b) Cramer, C.J.; Kormos, B.L.; Seierstad, M.; Sherer, E.C.; Winget, P. Org. Lett. 2001, 3, 1881.
23. Cremeens, M. E.; Hughes, T. S.; Carpenter, B. K. J. Am. Chem. Soc. 2005, 127, 6652.
24. (a) David, W. M.; Kumar, D.; Kerwin, S. M. Bioorg. Med. Chem. Lett. 2000, 10, 2509. (b) Tuntiwechapikul. W; David, W. M.; Kumar, D.; Salazar, M.; Kerwin, S. M. Biochemistry 2002, 41, 5283.
25. Tuesuwan, B; Kerwin, S. M. unpublished data.

## Chapter 5 Aza-enediynes undergo both aza-Bergman rearrangement to nitriles and dimerization to enediynes simultaneously

### 5.1 Introduction

Naturally occurring enediynes (Scheme 1-1, Scheme 1-2) are a class of potent anticancer agents that undergo Bergman cyclization or Myers-Saito-type cyclization to reactive diradical intermediates (Scheme 1-5), ${ }^{1,2}$ which abstract hydrogen atoms from the sugar phosphate backbone of DNA, leading to oxidative DNA strand scission and ultimately cell death. ${ }^{1 b, c}$ The applications of these enediynes in cancer chemotherapy are limited due to their lack of tumor specificity and toxicity. ${ }^{3,4}$ In 1997, David and Kerwin ${ }^{5}$ first reported a new class of enediynes, the aza-enediynes, or $C, N$-dialkynyl imines (Scheme 1-14, 26e,f) and reported that these aza-enediynes undergo an aza-Bergman cyclization to produce 2,5 -didehydropyridine intermediate diradicals that may have the potential to cleave DNA (Scheme 1-14, 27e,f), since then, these compounds have been the subjects of a number of theoretical ${ }^{6,7,8,9,10}$ and experimental studies. ${ }^{10,11,12}$ In contrast to $1,4-\mathrm{ddb}, 2,5-\mathrm{ddp}$ diradicals are very short-lived un-reactive species and can undergo a rapid retro-aza-Bergman reaction to nitriles ${ }^{5,10,12}$ (Scheme 1-14, 28e,f). Chen ${ }^{10}$ and coworkers reported the trapped pyridine product of $2,5-\mathrm{ddp}$ from the thermolysis of azaenediyne 26e (Scheme 1-14) carried out under acidic conditions, and the isolation of this trapped pyridine product from samples of 26e stored at $-4^{\circ} \mathrm{C}$ for months. We have been unable to confirm this finding. Instead, we isolated $(E)$-, and $(Z)$-enediynes ${ }^{13} \mathbf{6 0 e}$ and bisnitrile 61 (Scheme 5-2) from samples of $\mathbf{2 6 e}$ stored at $-10{ }^{\circ} \mathrm{C}$ for 24 months. The transformation of aza-enediyne to enediynes has also been observed in the case of triphenyl aza-enediyne 26d,g (Scheme 2-3). The unusual behavior of compounds 26d,e,g trigged our interests to further study this remarkable transformation reaction. Here I
report our kinetic and mechanistic studies of aza-enediyne 26d,e,g including ${ }^{13} \mathrm{C}$ labeled $\left[4-{ }^{13} \mathrm{C}\right]-26 \mathrm{~d} .{ }^{26}$

### 5.2 SYNTHESIS OF $\left[4-{ }^{13} \mathrm{C}\right]-26 \mathrm{D}$

Aza-enediyne $\left[4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ was prepared according to the procedure analogous to that for the preparation of $\mathbf{2 6 d}$ (Scheme 2-3) by using ${ }^{13} \mathrm{C}$ labeled 1,3-diphenylpropynone $\left[1-{ }^{13} \mathrm{C}\right]-32 d$ (Scheme $5-1$ ). Briefly, the addition of the cuprate reagent derived from the phenyl acetylene to the mesitylate ${ }^{16}\left[1-{ }^{13} \mathrm{C}\right]-\mathbf{3 8 d}$, that was obtained from the condensation of the $\left[1-{ }^{13} \mathrm{C}\right]-\mathbf{3 2 d}$ with $O$-mesitylenesulfonyl hydroxylamine 37, afforded $\left[4-{ }^{13} \mathrm{C}\right]-26 d$ in $53 \%$ yield (Scheme 5-1).


Scheme 5-1 Synthesis of [4- $\left.{ }^{13} \mathrm{C}\right]-26 d$

Mesitylate $\left[1-{ }^{13} \mathrm{C}\right]-\mathbf{3 8 d}$ was isolated as predominant single isomer of undetermined stereochemistry as judged from ${ }^{1} \mathrm{H}$ NMR and ${ }^{13} \mathrm{C}$ NMR. The aza-enediyne [ $\left.4-{ }^{13} \mathrm{C}\right]-26 d$ can be purified by chromatography and isolated as single isomer based on ${ }^{1} \mathrm{H}$ NMR and ${ }^{13} \mathrm{C}$ NMR. Our previous calculations ${ }^{12}$ at B3LYP level demonstrated that the $(Z)$-isomer of aza-enediyne 40a (Scheme 2-5) is more stable than the $(E)$-isomer by 4.4 $\mathrm{kcal} / \mathrm{mol}$. We predict that the $(Z)$-isomer of $\mathbf{2 6 d}$ is more stable than the $(E)$-isomer.

Therefore, the single isomer observed for $\mathbf{2 6 d}$ is the more thermodynamically stable $(Z)$ isomer.

### 5.3 KINETIC AND MECHANISTIC STUDIES OF THE TRANSFORMATION OF AZAENEDIYNES 26D,E,G TO ENEDIYNES 60D,E

The neat aza-enediyne $\mathbf{2 6 d}$ is not stable even stored in a $-10{ }^{\circ} \mathrm{C}$ freezer. After being stored in a freezer for weeks, TLC of the samples of $\mathbf{2 6 d}$ showed that the small amounts of $\mathbf{2 6 d}$ converted to new unidentified compounds. A solution of 90 mg of azaenediyne $26 d(120 \mathrm{mM})$ in benzene was stirred at $55^{\circ} \mathrm{C}$ for 5 days until aza-enediyne 26d disappeared upon TLC analysis. The reaction generated a complex mixture from which $(E)$ - and ( $Z$ )-tetraphenyl enediynes $\mathbf{6 0 d}$ ( $68 \%, 1: 1$ ratio after chromatograph) and (E)-dicyano compound 61 (44\%) (Scheme 5-2) were isolated along with other unidentified more polar materials, presumably polymers. The structures of compounds 60d and 61 were confirmed by ${ }^{1} \mathrm{H}$ NMR, ${ }^{13} \mathrm{C}$ NMR and HRMS analysis. ${ }^{17,18}$


Scheme 5-2 Thermolyses of aza-enediynes $\mathbf{2 6 d}, \mathbf{e}, \mathbf{g}$

Thermolysis of aza-enediyne $\mathbf{2 6 g}$ with a 4 -(Me)Ph substituent ( 110 mM in benzene) at $50{ }^{\circ} \mathrm{C}$ afforded enediynes $(Z)-,(E)-60 d$ (1:1 after flash chromatography, 47\%), nitrile 28g with one 4-(Me)Ph substituent (14\%) and bisnitrile 62 with two 4(Me)Ph substituents (26\%) (Scheme 5-2). Compounds 28g and 62 were isolated as the mixtures (2:3 as judged by ${ }^{1} \mathrm{H}$ NMR). We propose that nitrile $\mathbf{2 8 g}$ was generated from the aza-Bergman-retro-aza-Bergman rearrangement of aza-enediyne $\mathbf{2 6 g}$. The small amount of pure bisnitrile $\mathbf{6 2}$ was obtained by re-crystallization of the mixtures of $\mathbf{2 8 g}$ and $\mathbf{6 2}$ from hexanes and ethyl acetate as white needle crystal. The structure of $\mathbf{6 2}$ was confirmed by ${ }^{1} \mathrm{H}$ NMR and CI-MS.

When the solution of aza-enediyne $\mathbf{2 6 d}(0.45 \mathrm{mM})$ in benzene was refluxed at 90 ${ }^{\circ} \mathrm{C}$ for 24 hours, the retro-aza-Bergman rearranged nitrile 28d was isolated as the sole product in $85 \%$ yield (Scheme 5-2). To gain further insight of this transformation of azaenediyne to enediynes, ${ }^{13} \mathrm{C}$ labeled aza-enediyne $\left[4-{ }^{13} \mathrm{C}\right]-26 d$ was prepared. Thermolysis of aza-enediyne $\left[4-{ }^{13} \mathrm{C}\right]$-26d ( 120 mM in benzene) at $70{ }^{\circ} \mathrm{C}$ afforded di- ${ }^{13} \mathrm{C}$ labeled enediynes $(Z)-,(E)-60 d$ (1:3 after flash chromatography, $60 \%$ ), mono- ${ }^{13} \mathrm{C}$ labeled retro-aza-Bergman rearranged nitrile 28d (24\%), and bisnitrile 61 (35\%) (Scheme 5-3). No label was detected in the bisnitrile 61. ${ }^{13} \mathrm{C}$-labeled nitrile 28d was generated from the aza-Bergman-retro-aza-Bergman rearrangement. NMR analysis of enediynes 60d demonstrated that both $(Z)-60 d$ and $(E)-60 d$ have two labels, one is alkynyl carbon $((E)-$ 60d: $\delta$ 98.56; (Z)-60d: $\delta 96.84$ ), and the other one is olefinic carbon $((E)-60 d: \delta 128.63$; $(Z)$-60d: $\delta 129.23$ ); the alkynyl carbon is coupled with the olefinic carbon to give the coupling constant 4.6 Hz for $(E) \mathbf{- 6 0 d}$ and 4.1 Hz for ( $Z$ ) $\mathbf{- 6 0 d}$, respectively. After comparing our coupling constants with the ones obtained from reported similar structures such as hept-1-yne $\left.{ }^{19,20}\left({ }^{3} J_{1,4}=3.1 \mathrm{~Hz}\right),{ }^{2} J_{1,3}=12.5 \mathrm{~Hz}\right)$ and pent-4-en-2-yn-1-ol ${ }^{19,21}\left({ }^{3} J_{1,4}\right.$
$=2.0 \mathrm{~Hz},{ }^{2} J_{1,3}=12.4 \mathrm{~Hz}$ and ${ }^{2} J_{2,4}=11.9 \mathrm{~Hz}$ ), we assign that two labels are at $1-$ and $4-$ positions.


Scheme 5-3 Thermolysis of $\left[4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ in benzene

The transformation of aza-enediyne 26d to enediynes $\mathbf{6 0 d}$ and the conversion of 26d to nitrile 28d can be followed by ${ }^{1} \mathrm{H}$ NMR. Briefly, a NMR tube containing degassed aza-enediyne $26 d$ solution in $d_{6}$-benzene $(120 \mathrm{mM})$ and 2,5-dimethylfuran (1.5 equiv, internal standard) was sealed under flame. ${ }^{1} \mathrm{H}$ NMR of the sample was taken as initial spectrum (reaction time equals zero second). After ${ }^{1} \mathrm{H}$ NMR, the tube was kept at $70{ }^{\circ} \mathrm{C}$. The conversion of aza-enediyne to enediyne and nitrile was followed spectroscopically by taking ${ }^{1} \mathrm{H}$ NMR spectra 15 times over a period of time. Intervals varied from 35 minutes to 5 hours. Every time, the ${ }^{1} \mathrm{H}$ NMR tube was taken out of a $70^{\circ} \mathrm{C}$ oil bath, and immediately inserted into a beaker containing ice-water to stop the reaction. Reaction time was measured as the time during which the tube maintained at $70^{\circ} \mathrm{C}$. Aza-enediyne 26d has a doublet resonance at 7.45 ppm which is well resolved from other peaks. The disappearance of the aza-enediyne $\mathbf{2 6 d}$ was monitored as the disappearance of the resonance at 7.45 ppm . The integrals of the resonance for $\mathbf{2 6 d}$ at 7.45 ppm were normalized against the resonance for the internal standard 2,5-dimethylfuran at 5.76 ppm . The relative concentrations of aza-enediyne 26d were calculated by normalizing the integrals of the resonance for $\mathbf{2 6 d}$ at 7.45 ppm obtained at different intervals against the
integral of the resonance for $\mathbf{2 6 d}$ at 7.45 ppm obtained at reaction time equals zero second. The kinetic data was obtained by fitting plots of the concentrations of azaenediyne 26d versus time. ${ }^{231}$ H NMR kinetic data demonstrated that the disappearance of 26d does not follow either first-order kinetics or second-order kinetics, but follows independent first-order and second-order kinetics simultaneously (Table 5-1). ${ }^{23}$ We have demonstrated that the conversion of aza-enediynes to nitriles follow first-order kinetics for both the disappearance of aza-enediynes and the appearance of nitriles. ${ }^{12}$ Here we discover that the conversion of aza-enediyne 26d to nitrile 28d in benzene follows firstorder kinetics with a half-life of 4.3 hours; the transformation of aza-enediyne 26d to enediyne 60d follows second-order kinetics (Table 5-1).

The conversion of aza-enediyne $\left[4-{ }^{13} \mathrm{C}\right]-26 d$ to enediyne $\left[1,4-{ }^{13} \mathrm{C}_{2}\right]-\mathbf{6 0 d}$ and nitrile $\left[5-{ }^{13} \mathrm{C}\right]-28 d$ can be followed by ${ }^{13} \mathrm{C}$ NMR by a procedure analogous to that described for ${ }^{1} \mathrm{H}$ NMR kinetics. The resonance for [4- $\left.{ }^{13} \mathrm{C}\right]-26 \mathrm{~d}$ at $159 \mathrm{ppm}\left({ }^{13} \mathrm{C}\right)$ diminished over time and was replaced by the resonance for $\left[1,4-{ }^{13} \mathrm{C}_{2}\right]-60 \mathrm{~d}$ at 97 ppm and $129 \mathrm{ppm}\left({ }^{13} \mathrm{C}\right)$ and the resonance for $\left[5-{ }^{13} \mathrm{C}\right]-\mathbf{2 8 d}$ at $100 \mathrm{ppm}\left({ }^{13} \mathrm{C}\right)$. The integrals of the resonance for $\left[4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ at 159 ppm were normalized against the resonance for the solvent (benzene, THF or DMSO). The relative concentrations of aza-enediyne $\left[4-{ }^{13} \mathrm{C}\right]$ $\mathbf{2 6 d}$ were calculated by normalizing the integrals of the resonance for $\left[4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ at 159 ppm obtained at different intervals against the integral of the resonance for $\left[4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ at 159 ppm obtained at reaction time equals zero second. The kinetic data was obtained by fitting plots of the concentrations of aza-enediyne $\left[4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ versus time. ${ }^{23}$ Kinetic data obtained from ${ }^{13} \mathrm{C}$ NMR studies were also independent first-order and second-order (Table 5-1). At $70{ }^{\circ} \mathrm{C}$, the second-order transformation reaction is faster than the firstorder aza-Bergman cyclization in either benzene, THF or DMSO. We have performed kinetic studies of 26d in different solvents to determine the solvent effect (Table 5-1). We
have reported that the conversion of aza-enediyne to nitrile is solvent dependent; proceeding more rapidly in less polar solvents. ${ }^{12}$ Here we report that the conversion of aza-enediyne 26d to nitrile 28d follows first-order kinetics and proceeds more rapidly in benzene than in more polar solvents such as THF and DMSO (Table 5-1). Importantly, ${ }^{13} \mathrm{C}$ NMR kinetic studies demonstrate that more than $90 \%$ of enediyne generated from the transformation is $(Z)$ - 60 d which slowly isomerizes to thermodynamic more stable $(E)$ 60d isomer during extended heating or chromatography on silica gel to mixtures of $(E)$-, and ( $Z$ )-60d.

Table 5-1 Rate constants for the first-order aza-Bergman rearrangement of 26d and the second-order transformation of $\mathbf{2 6 d}$ to $\mathbf{6 0 d}$ at $70 \pm 1^{\circ} \mathrm{C}$

| Aza-enediyne $^{\mathrm{a}}$ | First-order reaction $^{\text {Half-life }}$ |  | Second-order reaction |
| :---: | :---: | :---: | :---: |
|  | First-order rate constant <br> $\mathrm{k}_{1}\left(\mathrm{~s}^{-1}\right)$ | Second-order rate constant <br> $\mathrm{t}_{1 / 2}(\mathrm{~h})$ | $\mathrm{k}_{2}\left(\mathrm{~L} \mathrm{~mol}^{-1} \mathrm{~s}^{-1}\right)$ |

${ }^{\text {a }}$ Initial 26d or $\left[4-{ }^{13} \mathrm{C}\right]-26 d$ concentration of 120 mM . ${ }^{\text {b }}$ avarage values of one ${ }^{1} \mathrm{H}$ NMR run and one ${ }^{13} \mathrm{C}$ NMR run; ${ }^{\mathrm{c}}$ values from one ${ }^{13} \mathrm{C}$ NMR run.

Kamigata ${ }^{17}$ and co-workers reported the synthesis of $(Z)$-, $(E)$ - $\mathbf{6 0 d}$ from seleniumsubstituted allenes and proposed the formation of diphenyl propargyl carbene via the homolytic cleavage of the carbon-selenium bond of allene followed by the attack of the seleno radical to the selenium atom. Compound 60d generated from aza-enediyne 26d would also be formed through the diphenyl propargyl carbene. However, thermolyses of aza-enediyne $\mathbf{2 6 d}$ in benzene at $50^{\circ} \mathrm{C}$ in the presence of cyclohexene or 1,4-CHD (both are good carbene trapping reagents) only gave enediynes 60d and bisnitrile 61, and did
not afford any carbene trapping products. Based on the kinetic and mechanistic studies of aza-enediynes 26d,e,g, we proposed that the transformation of aza-enediynes to enediynes occurs through a "head-to-tail" coupling of the propargyl vinylidene units and a "tail-to-tail" coupling of the ynamine units. One possible mechanism is shown in Scheme 5-4. "Head-to-tail" $[2+2]$ cyclization affords an aza-cyclobutene which opens to give an $\alpha$-diimine. Diimine then leads to enediyne 60d,e and bisnitrile 61/62.


Scheme 5-4 Proposed mechanism of the formation of enediyne 60d,e from aza-enediyne 26d,e,g

It has been reported that certain transition metals such as osmium ${ }^{24(a)}$ and ruthenium ${ }^{24(b)}$ can catalyze olefin cross metathesis. The transformation of aza-enediyne $\mathbf{2 6 d}$ to enediyne $\mathbf{6 0 d}$ in the presence of 1.0 equiv of copper (I) bromide dimethyl sulfide complex in THF $(120 \mathrm{mM})$ at $50^{\circ} \mathrm{C}$ proceeded as slowly as the reaction in the absence of copper (I) salt. Therefore, trance amounts of $\mathrm{Cu}(\mathrm{I})$, if introduced from cuprate addition reaction, do not affect the transformation reaction.

### 5.4 CONCLUSIONS

$\mathrm{We}^{5,12}$ have demonstrated that aza-enediynes undergo aza-Bergman cyclization to 2,5-ddp diradical intermediates, which then undergo retro-aza-Bergman rearrangement to nitriles (Scheme 2-5). On the other hand, certain aza-enediynes ${ }^{25}$ can undergo reaction under acidic conditions to afford carbene intermediates which can be trapped efficiently (Scheme 3-5). Here we ${ }^{26}$ present the evidence for a remarkable transformation of azaenediynes 26d,e,g to enediynes 60d,e. Aza-enediynes (40a-c) that undergo rapid azaBergman cyclization do not undergo the transformation to enediynes. Aza-enediynes (26a) having bulky terminal substituents such as TIPS group do not undergo the transformation reaction. Aza-enediynes such as 26d,e,g that undergo aza-Bergman rearrangement slowly can convert to enediynes through a dimerization mechanism (Scheme 5-4). In this case, at high concentration of aza-enediynes, the transformation to enediynes can predominate.

### 5.5 SUMMARY AND FUTURE DIRECTIONS

Classes of $C, N$-dialkynyl imines (3-ene-3-aza-1,5-diynes, aza-enediynes) and 3-ene-4-aza-1,6-diynes (skipped aza-enediynes) have been designed and synthesized in the present study. The substitution of nitrogen for carbon in the enediyne system can have a profound impact on the facility of the Bergman rearrangement and on the nature of the intermediates that are involved. These aza-enediynes undergo a facile aza-Bergman cyclization to generate 2,5-didehydropyridine diradicals that undergo a rapid retro-azaBergman rearrangement to ( $Z$ )- $\beta$-alkynyl acrylonitriles. The convertion of aza-enediynes to the nitriles follows first-order kinetics. The rate of aza-Bergman cyclization is solvent dependent, proceeding more rapidly in less polar solvents. The kinetic studies of the azaBergman cyclization of a series of 6-triisopropylsilyl and 6-unsubstituted 4-aryl-3-aza-3-
ene-1,4-diynes demonstrate that there are two types of substituent effects in this system; the aza-Bergman reaction rate is not greatly affected by changing the 4 -aryl substituent on the aza-enediyne imine double bond, but is very sensitive to the nature of the 6 substitutent at the alkyne terminus.

Certain aza-enediynes (40a) can undergo reaction under acidic conditions to afford carbene intermediates which can be trapped efficiently. No pyridine products corresponding to trapping of $2,5-\mathrm{ddp}$ intermediates from the thermolysis of aza-enediynes 26a/40a under acidic conditions are detected, which is in contrast with Chen ${ }^{10}$ and coworkers' report of a pyridine product from the thermolysis of aza-enediyne 26e under acidic conditions or from the samples of $\mathbf{2 6 e}$ stored for months at $-4^{\circ} \mathrm{C}$. We were unable to isolate the pyridine product from the samples of $\mathbf{2 6 e}$ stored for two years at $-10^{\circ} \mathrm{C}$. On the other hand, aza-enediynes such as $\mathbf{2 6 d}, \mathbf{e}, \mathbf{g}$ that undergo aza-Bergman rearrangement slowly can undergo a remarkable transformation to produce enediynes through a dimerization mechanism. We also demonstrate that aza-enediynes (26a) having bulky terminal substituents such as TIPS group do not undergo the transformation to enediynes. The transformation of aza-enediynes to enediynes follows second-order kinetics.

The skipped aza-enediynes can isomerize to the corresponding $C$-alkynyl- $N$ allenyl imines (aza-enyne allenes) under basic conditions. The resulting aza-enyne allenes undergo a facile aza-Myers-Saito cyclization to generate previously un-reported $\alpha, 5$-didehydro-3-picoline diradicals that can be efficiently trapped. The isomerization of skipped aza-enediynes and their subsequent rapid aza-Myers-Saito cyclization may have relevance to the recently reported DNA cleavage ability of cationic heterocyclic skipped aza-enediynes such as AZB002 and AZB037 (Figure 1-6).

The 1,4-didehydrobenzenoid diradicals produced by the Bergman cyclization of enediynes are responsible for the DNA cleavage ability of these enediynes. ${ }^{1}$ However,
unlike enediynes, aza-enediynes studied cannot cleave DNA, since 2,5-didehydropyridine diradicals produced by the aza-Bergman cyclization of aza-enediynes are a very short lived and unreactive species, and these intermediates undergo a rapid retro-aza-Bergman rearrangement to ( $Z$ )- $\beta$-alkynyl acrylonitriles. ${ }^{5,12}$

Protonation of the nitrogen of the aza-enediyne is predicted to increase the diradical reactivity towards hydrogen atom abstraction, which would lead to the proposal of a pH -selective aza-enediyne. ${ }^{10}$ However, attempts to trap these 2,5-ddp diradicals under acidic conditions afforded cyclopropane derivatives derived from carbene intermediates (Scheme 3-6). ${ }^{25}$ It is possible that under acidic conditions the ynamine carbon of aza-enediyne can be protonated followed by addition of water affording carbene intermediate responsible for the formation of cyclopropane derivative. Increasing the $\mathrm{pK}_{\mathrm{a}}$ value of imine nitrogen would help protonate the nitrogen under acidic conditions. Based on our studies, the aza-Bergman cyclization rate is not greatly affected by the nature of the imine double bond substituent. ${ }^{12}$ We propose replacing the imine double bond of the aza-enediyne system with the amidine group to afford aza-enediynes such as 63 shown in Scheme 5-5 would facilitate nitrogen protonation due to the increased $\mathrm{pK}_{\mathrm{a}}$ value of nitrogen. In addition, incorporating amidine goup into azaenediyne system would prevent the hydrolysis of imine double bond. Therefore, it would be interesting to prepare aza-enediynes 63 (Scheme 5-5) and to study thermolyses of these aza-enediynes under acidic conditions.


Scheme 5-5

Since 2,5 -ddp diradical intermediates are a very short lived and unreactive species, these intermediates undergo a rapid retro-aza-Bergman rearrangement to produce thermodynamically more stable nitrile. We hypothesize that protonation of the nitrogen in aza-enediyne system through an intra-molecular proton transfer pathway may proceed much more rapidly than that through an inter-molecular proton transfer. ${ }^{27}$ Hence, under this conditions, 2,5 -ddp diradical intermediate may have potential to be protonated before it undergoes retro-aza-Bergman rearrangement to produce nitrile. Based on our hypothesis, we propose that aza-enediyne 67 undergoes aza-Bergman cyclization to produce diradical 68, whose nitrogen atom may be protonated by the adjacent acidic phenol group (Scheme 5-6). The protonated diradical 68 can abstract two hydrogen atoms from 1,4-CHD.


Scheme 5-6 Protonation of the nitrogen atom of 2,5-ddp diradical $\mathbf{6 8}$ via an intramolecular proton transfer from the adjacent acidic phenol group

As mentioned in Chapter 4, aza-enyne allenes $\mathbf{4 d}$,f undergo a facile aza-MyersSaito cyclization to produce $\alpha, 5$-didehydro-3-picoline diradicals which can be trapped efficiently (Scheme 4-4). We were unable to detect any products corresponding to the trapping of diradicals produced by Schmittel cyclization of aza-enyne allenes 4d,f. It has been reported that the enyne allenes with radical stabilizing substituents such as aryl or with bulky groups at the $\mathrm{C}^{7}$ position (alkyne terminus) favor the Schmittel cyclization (Scheme 1-12). ${ }^{28}$ We propose that an aza-enyne allene with a phenyl substituent at the alkyne terminus such as $\mathbf{7 0}$ may undergo aza-Schmittel cyclization to produce a pyrrolelike diradical 71, which may have potential to abstract two hydrogen atoms from hydrogen donor such as 1,4 -CHD to afford a pyrrole-like product 72 (Scheme 5-7, path A) or undergo intramolecular radical recombination followed by rearrangement of 73 to afford product 74 (Scheme 5-7, path B).

70

71
72

74
73

Scheme 5-7 Proposed aza-Schmittel cyclization of aza-enyne allene 70

The initial goal of this study was to design novel enediyne core structures that target cancer cell with minimum toxicity towards normal cells. However, all of the azaenediynes studied here are unsuitable for DNA cleavage. Our studies suggest that azaenyne allenes may have the potential to cleave DNA via diradical imtermediates produced by aza-Myers-Saito cyclization of aza-enyne allenes. We hope that the findings presented in this study lay a foundation for the design of a novel enediyne core structure that can target cancer cells.

## REFERENCE

1. (a) Xi, Z.; Goldberg, I. H. Comprehensive natural products chemistry. Elsevier, New York, 1999, vol 7, 533-592. (b) Cosgrove, J. P.; Dedon, P. C. Small molecule DNA and RNA binders. 2003, 2, 609-642. (c) De Voss, J. J.; Townsend, C. A.; Ding, W. D.; Morton, G. O.; Ellestad, G. A.; Zein, N.; Tabor, A. B.; Schreiber, S. L. J. Am. Chem. Soc. 1990, 112, 9669.
2. (a) Myers, A. G.; Kuo, E. Y.; Finney, N. S. J. Am. Chem. Soc. 1989, 111, 8057. (b) Myers, A. G.; Dragovich P. S.; Kuo, E. Y. J. Am. Chem. Soc. 1992, 114, 9369. (c) Nagata, R.; Yamanaka H.; Okazaki, E.; Saito, I. Tetrahedron Lett. 1989, 30, 4995. (d) Nagata, R.; Yamanaka, H.; Murahashi, E.; Saito, I. Tetrahedron Lett. 1990, 31, 2907.
3. Zhen, Y. S.; Ming, X. Y.; Yu, B.; Otani, T.; Saito, H.; Yamada, Y. J. Antibiot. 1989, 42, 1294.
4. Thorson, J. S.; Sievers, E. L.; Ahlert, J.; Shepard, E.; Whitwan, R. E.; Onwueme, K. C.; Ruppen, M. Curr. Pharm. Des. 2000, 6, 1841.
5. David, W. M.; Kerwin, S. M. J. Am. Chem. Soc. 1997, 119, 1464.
6. Cramer, C. J. J. Am. Chem. Soc. 1998, 120, 6261.
7. Kraka, E.; Cremer, D. J. Am. Chem. Soc. 2000, 122, 8245.
8. Kraka, E.; Cremer, D. J. Comput. Chem. 2001, 22, 216.
9. Schreiner, P. R.; Navarro-Vazquez, A.; Prall, M. Acc. Chem. Res. 2005, 38, 29.
10. Hoffer, J.; Schottelius, M. J.; Feichhtinger, D.; Chen, P. J. Am. Chem. Soc. 1998, 120, 376.
11. David, W. M.; Kumar, D.; Kerwin, S. M. Bioorg. Med. Chem. Lett. 2000, 10, 2509.
12. Feng, L.; Kumar, D.; Kerwin, S. M. J. Org. Chem. 2003, 68, 2234.
13. The assignment of the structures of $(E)$-, $(Z)$-3,6-dimethyl-1,4-diphenyl-3-hexene-1,5diynes 60 e is based on the comparison of ${ }^{1} \mathrm{H}$ NMR of $\mathbf{6 0 e}$ with ${ }^{1} \mathrm{H}$ NMR of reported 1,6-dimethyl-3,4-diphenyl-3-hexene-1,5-diyne (ref. 15) and 3,4-dimethyl-1,6-diphenyl-3-hexene-1,5-diyne (ref. 14,15); 60e: ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 2.01(\mathrm{~s}, 3 \mathrm{H}), 2.08$ $(\mathrm{s}, 3 \mathrm{H}), 2.09(\mathrm{~s}, 3 \mathrm{H}), 2.30(\mathrm{~s}, 3 \mathrm{H}), 7.23-7.36(\mathrm{~m}, 16 \mathrm{H}), 7.49-7.51(\mathrm{~m}, 2 \mathrm{H}), 7.77$ (d, $2 \mathrm{H}, J=7.6 \mathrm{~Hz}$ ); HRMS $m / z 257.1323$ (calcd 257.1330, $\mathrm{C}_{20} \mathrm{H}_{17}$ ).
14. Blackwell, J. M.; Figueroa, J. S.; Stephens, F, H.; Cummins, C. C. Organometallics 2003, 22, 3351.
15. Hayashi, M.; Saigo, K. Tetrahedron Lett. 1997, 38, 6241.
16. (a) Krause, J. G. Synthesis, 1972, 3, 140. (b) Carpino, L. A. J. Am. Chem. Soc. 1960, 82, 3133.
17. Shimizu, T.; Miyasaka, D.; Kamigata, N. Organic Lett. 2000, 2, 1923.
18. Refat, H. M.; Waggenspack, J.; Dutt, M.; Zhang, H.; Fadda, A. A.; Biehl, E. J. Org. Chem. 1995, 60, 1985.
19. Krivdin, L. B.; Della, E. W. Progress in NMR Spectroscopy 1991, 23, 301.
20. Porwoll, J. P.; Leete, E. J. Labelled. Comp. Radiopharm. 1985, 22, 257.
21. Lambert, J.; Klessinger, M. Magn. Reson. Chem. 1987, 25, 456.
22. Chaloner, P. A. J. Chem. Soc. Perkin Trans. 2 1980, 1028.
23. TableCurve $2 D^{\circledR}$ from SYSTAT Software Inc. was used to analyze the kinetic data. Kinetic data fits independent first-order and second-order kinetics with an equation: $\frac{d[A]}{d t}=-k_{1}[A]-k_{2}[A]^{2}, \quad[\mathrm{~A}]$ : concentration of aza-enediyne 26d; $\mathrm{k}_{1}$ : first-order constant; $\mathrm{k}_{2}$ : second-order constant.
24. (a) Castarlenas, R.; Esteruelas, M. A.; Onate, E. Organometallics 2005, 24, 4343. (b) Schmidt, B. J. Org. Chem. 2004, 69, 7672.
25. Feng, L.; Kerwin, S. M. Tetrahedron Lett. 2003, 44, 3463.
26. Feng, L.; Zhang, A.; Kerwin, S. M. submitted.
27. Zimmerman, H. E.; Cheng, J. Org. Lett. 2005, 7, 2595.
28. Engels, B.; Lennartz, C.; Hanrath, M.; Schmittel, M.; Strittmatter, M. Angew. Chem. Int. Ed. 1998, 37, 1960.

## Chapter 6 Experimental section

### 6.1 GENERAL

All commercial chemicals were used without further purification unless otherwise noted. Tetrahydrofuran (THF) was re-distilled from $\mathrm{Na} /$ benzophenone, methylene chloride, diethyl ether, and pyridine were re-distilled from calcium hydride prior to use. All reactions were performed under argon. Unless otherwise noted, organic extracts were dried with $\mathrm{Na}_{2} \mathrm{SO}_{4}$, filtered through a fritted glass funnel, and concentrated with a rotary evaporator ( $20-30 \mathrm{mmHg}$ ). $R_{f}$ values are reported for thin-layer chromatography (TLC) performed on silica gel TLC plates, using the indicated mobile phase. Flash chromatography was preformed with EM silica gel. ${ }^{1} \mathrm{H}$ NMR spectra were recorded at 500, 300, and 250 MHz and ${ }^{13} \mathrm{C}$ NMR spectra were recorded at 100,75 and 62 MHz . Spectroscopic studies were performed on a spectrophotometer with a Peltier thermostated cuvette holder. All mass spectra were obtained by chemical ionization with methane as the ionizing gas. Melting points are uncorrected. The yields for compounds 60d,e and 61/62 were calculated based on 2:1 stoichiometry between 26d,e,g and 60d,e, 61/62.

All calculations were carried out by using (U)B3LYP/6-31G* with Gaussian 98 program. ${ }^{35}$ Closed-shell singlet states were computed using B3LYP/6-31G*. Open-shell singlet diradicals and triplet diradicals were computed using unrestricted UB3LYP/6$31 G^{*}$, and the open-shell singlet states were specified by using the command GUESS $=$ ALTER in the input and switching the HOMO and LUMO for the $\beta$ electron. All stationary structures were identified as minima (number of imaginary frequencies $=0$ ) or transition states (number of imaginary frequencies $=1$ ) by computing vibrational frequencies. The energies reported here were corrected to zero-point vibrational energies obtained by computing vibration frequencies.

### 6.2 SYNTHESES

### 6.2.1 1-Phenyl-3-(triisopropylsilyl)-propynone 32a



To a mixture of triisopropylsilyl acetylene ( $0.5 \mathrm{~g}, 2.74 \mathrm{mmol}$ ) and cuprous iodide ( 26.6 $\mathrm{mg}, 0.14 \mathrm{mmol})$ in triethylamine $(9 \mathrm{~mL})$ was added benzoyl chloride $(0.5 \mathrm{~g}, 3.56 \mathrm{mmol})$. The reaction mixture was stirred at room temperature under argon for 30 h . After the removal of the solvent, methanol ( 3 mL ) was added and the mixture was stirred for an additional 5 min , during which time the mixture became clear. The methanol was distilled off under vacuum, and the residue was poured into water and extracted with dichloromethane $(2 \times 20 \mathrm{~mL})$ and dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$. The solvent was distilled off and the product was purified by Kugelrohr distillation $\left(110{ }^{\circ} \mathrm{C}, 1 \mathrm{mmHg}\right)$ to afford 0.69 g ( $87 \%$ ) of propynone 32a as a clear oil: ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 1.01-1.22(\mathrm{~m}, 21 \mathrm{H}), 7.46-7.55$ $(\mathrm{m}, 3 \mathrm{H}), 8.16-8.23(\mathrm{~m}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right)$ 811.07, 18.53, 97.91, 103.01, 128.53, $129.49,134.00,136.70,177.43 ;$ HRMS $m / z 287.1826$ (calcd 287.1831, $\mathrm{C}_{18} \mathrm{H}_{26} \mathrm{OSi}$ ).

### 6.2.2 1-(4-Methoxyphenyl)-3-(triisopropylsilyl)-propynone 32b



To a mixture of triisopropylsilyl acetylene ( $3.0 \mathrm{~g}, 16.45 \mathrm{mmol}$ ) and cuprous iodide (156.4 $\mathrm{mg}, 0.82 \mathrm{mmol})$ in triethylamine $(35 \mathrm{~mL})$ was added $p$-Anisoyl chloride ( $3.5 \mathrm{~g}, 20.57$ mmol ). The reaction mixture was stirred at room temperature under argon for 30 h . After the removal of the solvent, methanol $(10 \mathrm{~mL})$ was added and the mixture was stirred for an additional 5 min , during which time the mixture became clear. The methanol was distilled off under vacuum, and the residue was poured into water and extracted with dichloromethane $(2 \times 20 \mathrm{~mL})$ and dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$. The solvent was distilled off and the product was purified by flash chromatography $\left(0-30 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes) to afford $4.8 \mathrm{~g}(92 \%)$ of propynone $\mathbf{3 2 b}$ as a transparent oil: $R_{f} 0.85\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CD}_{2} \mathrm{Cl}_{2}\right)$ 81.11-1.30 (m, 21H), $3.86(\mathrm{~s}, 3 \mathrm{H}), 6.95(\mathrm{~d}, J=8.0 \mathrm{~Hz}, 2 \mathrm{H}), 8.13(\mathrm{~d}, J=8.0 \mathrm{~Hz}, 2 \mathrm{H}),{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CD}_{2} \mathrm{Cl}_{2}\right) \delta 11.54,18.74,55.99,97.21,103.87,114.24,130.80,132.12,165.04$, 176.23; HRMS m/z 317.1930 (calcd 317.1936, $\mathrm{C}_{19} \mathrm{H}_{28} \mathrm{O}_{2} \mathrm{Si}$ ).

### 6.2.3 1-(2-Methoxyphenyl)-3-(triisopropylsilyl)propynone 32c



To a mixture of triisopropylsilyl acetylene ( $0.5 \mathrm{~g}, 2.74 \mathrm{mmol}$ ) and cuprous iodide ( 26.6 $\mathrm{mg}, 0.14 \mathrm{mmol})$ in triethylamine ( 9 mL ) was added o-anisoyl chloride ( $0.5 \mathrm{~g}, 3.56$ mmol ). The reaction mixture was stirred at room temperature under argon for 30 h . After the removal of the solvent, methanol $(3 \mathrm{~mL})$ was added and the mixture was stirred for an additional 5 min , during which time the mixture became clear. The methanol was distilled off under vacuum, and the residue was poured into water and extracted with dichloromethane $(2 \times 20 \mathrm{~mL})$ and dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$. The solvent was distilled off and the product was purified by flash chromatography ( $10 \% \mathrm{EtOAc}$ in hexanes) to afford 4.62 $\mathrm{g}(74 \%)$ of 32c as a transparent oil: $R_{f} 0.85\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right) ;{ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 0.95-1.20(\mathrm{~m}$, $21 \mathrm{H}), 3.85(\mathrm{~s}, 3 \mathrm{H}), 6.90-7.01(\mathrm{~m}, 2 \mathrm{H}), 7.45(\mathrm{ddd}, J=8.2,6.0,2.0 \mathrm{~Hz}, 1 \mathrm{H}), 7.99(\mathrm{dd}, J=$ 8.2, 2.0 Hz, 1H); ${ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 10.94,18.34,55.51,95.50,105.11,111.84,119.96$, $126.25,132.56,134.72,159.96,176.11$; HRMS $m / z 317.1944$ (calcd 317.1936, $\left.\mathrm{C}_{19} \mathrm{H}_{28} \mathrm{O}_{2} \mathrm{Si}\right)$.

### 6.2.4 1,3-Diphenyl-1- ${ }^{13} \mathrm{C}$-propynone 32 d



To a mixture of phenyl acetylene $(0.22 \mathrm{~g}, 2.2 \mathrm{mmol})$ and cuprous iodide ( $17.0 \mathrm{mg}, 0.08$ $\mathrm{mmol})$ in triethylamine $(5.0 \mathrm{~mL})$ was added benzoyl chloride $(0.20 \mathrm{~g}, 1.4 \mathrm{mmol})$. The reaction mixture was stirred at room temperature under argon for 48 h . After filtering off the salt, the solvent was distilled off and the product was purified by flash chromatography (hexanes/EtOAc, 20:1) to afford $0.35 \mathrm{~g}(60 \%)$ of 1,3 -diphenyl-1- ${ }^{13} \mathrm{C}$ propynone 32d as a yellow oil: $R_{f} 0.42(E t O A c / h e x a n e s, 1: 9) .{ }^{3}{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.31-$ $7.53(\mathrm{~m}, 5 \mathrm{H}), 7.56-7.70(\mathrm{~m}, 3 \mathrm{H}), 8.15-8.23(\mathrm{~m}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 86.78(\mathrm{~d}, J=$ $91.2 \mathrm{~Hz}), 93.01(\mathrm{~d}, J=13.7 \mathrm{~Hz}), 119.98(\mathrm{~d}, J=1.7 \mathrm{~Hz}), 128.54(\mathrm{~d}, J=5.0 \mathrm{~Hz}), 128.60$, $129.46(\mathrm{~d}, J=3.3 \mathrm{~Hz}), 130.73,132.96,134.05,136.74(\mathrm{~d}, J=61.1 \mathrm{~Hz}), 177.89\left({ }^{13} \mathrm{C}\right)$; HRMS $m / z 208.0842$ (calc'd 208.0843, ${ }^{12} \mathrm{C}_{14}{ }^{13} \mathrm{CH}_{11} \mathrm{O}$ ).

### 6.2.5 1-Phenyl-3-(triisopropylsilyl)-propynone oxime 33a



To a solution of $\mathbf{3 2} \mathbf{a}(0.64 \mathrm{~g}, 2.23 \mathrm{mmol})$ in 8 mL of EtOH was added hydroxylamine hydrochloride ( $186 \mathrm{mg}, 2.68 \mathrm{mmol}$ ) and sodium acetate $(220 \mathrm{mg}, 2.68 \mathrm{mmol})$. The mixture was stirred at room temperature under argon for 10 days. The reaction mixture was poured into water $(20 \mathrm{~mL})$, and the water layer was extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}(3 \times 15$ mL ). The residue upon drying and concentration of the organic layer was purified by flash chromatography $\left(0-20 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes) to afford $0.51 \mathrm{~g}(77 \%)$ of oxime 33a as a transparent oil: $R_{f} 0.54\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 1.04-1.30(\mathrm{~m}, 21 \mathrm{H}), 7.38-7.42$ $(\mathrm{m}, 3 \mathrm{H}), 7.80-7.89(\mathrm{~m}, 2 \mathrm{H}), 8.55(\mathrm{~s}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 11.13,18.61,94.61$, 107.42, 126.46, 128.46, 129.82, 133.12, 141.97; HRMS m/z 302.1946 (calcd 302.1940, $\mathrm{C}_{18} \mathrm{H}_{27} \mathrm{NOSi}$.

### 6.2.6 1-(4-Methoxyphenyl)-3-(triisopropylsilyl)propynone oxime 33b



To a solution of $\mathbf{3 2 b}(2.46 \mathrm{~g}, 7.77 \mathrm{mmol})$ in 15 mL of EtOH was added hydroxylamine hydrochloride ( $647 \mathrm{mg}, 9.32 \mathrm{mmol}$ ) and sodium acetate $(764 \mathrm{mg}, 9.32 \mathrm{mmol})$. The mixture was stirred at room temperature under argon for 10 days. The reaction mixture was poured into water $(40 \mathrm{~mL})$, and the water layer was extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}(3 \times 20$ $\mathrm{mL})$. The residue upon drying and concentration of the organic layer was purified by flash chromatography on silica gel $\left(0-20 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes) to afford $2.01 \mathrm{~g}(79 \%)$ of oxime 33b as a light yellow oil: $R_{f} 0.50\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CD}_{2} \mathrm{Cl}_{2}\right) \delta 0.96-1.35(\mathrm{~m}$, $21 \mathrm{H}), 3.85(\mathrm{~s}, 3 \mathrm{H}), 6.93(\mathrm{~d}, J=8.1 \mathrm{~Hz}, 2 \mathrm{H}), 7.82(\mathrm{~d}, J=8.1 \mathrm{~Hz}, 2 \mathrm{H}), 8.80(\mathrm{~s}, 1 \mathrm{H}),{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CD}_{2} \mathrm{Cl}_{2}\right) \delta 11.58,18.80,55.74,97.90,107.23,114.26,126.14,128.26,142.00$, 161.52; HRMS $m / z 332.2036$ (calcd 332.2045, $\mathrm{C}_{19} \mathrm{H}_{29} \mathrm{NO}_{2} \mathrm{Si}$ ).

### 6.2.7 1-(2-Methoxyphenyl)-3-(triisopropylsilyl)propynone oxime 33c



To a solution of $\mathbf{3 2 c}(0.64 \mathrm{~g}, 2.23 \mathrm{mmol})$ in 8 mL of EtOH was added hydroxylamine hydrochloride ( $186 \mathrm{mg}, 2.68 \mathrm{mmol}$ ) and sodium acetate $(220 \mathrm{mg}, 2.68 \mathrm{mmol})$. The mixture was stirred at room temperature under argon for 10 days. The reaction mixture was poured into water $(20 \mathrm{~mL})$, and the water layer was extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}(3 \times 15$ mL ). The residue upon drying and concentration of the organic layer was purified by flash chromatography $\left(0-20 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes) to afford $1.24 \mathrm{~g}(79 \%)$ of oxime 33 c as a white solid: $R_{f} 0.40\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right) ; \mathrm{mp} 120.2-121.0{ }^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H} \mathrm{NMR}\left(\mathrm{CD}_{2} \mathrm{Cl}_{2}\right) \delta 1.10-1.22(\mathrm{~m}$, $21 \mathrm{H}), 3.89(\mathrm{~s}, 3 \mathrm{H}), 6.97-7.08(\mathrm{~m}, 2 \mathrm{H}), 7.39(\mathrm{ddd}, J=8.0,7.0,2.0 \mathrm{~Hz}, 1 \mathrm{H}), 7.80(\mathrm{dd}, J=$ $8.0,2.0 \mathrm{~Hz}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CD}_{2} \mathrm{Cl}_{2}\right) \delta 11.57,18.78,55.71,96.46,105.56,111.84$, 120.77, 122.20, 131.01, 131.39, 140.42, 157.68; HRMS m/z 332.2058 (calcd 332.2045, $\mathrm{C}_{19} \mathrm{H}_{29} \mathrm{NO}_{2} \mathrm{Si}$ ).

### 6.2.8 1-Phenyl-3-(triisopropylsilyl)-propynone mesylate 34a



To a solution of oxime $\mathbf{3 3 a}(2.7 \mathrm{~g}, 8.96 \mathrm{mmol})$ in 60 mL of dry $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ was added dry pyridine ( $3.6 \mathrm{~mL}, 44.8 \mathrm{mmol}$ ). The reaction mixture was cooled to $0^{\circ} \mathrm{C}$, followed by dropwise addition of a solution of methanesulfonyl chloride ( $1.28 \mathrm{~g}, 11.2 \mathrm{mmol}$ ) in 5 mL of dry $\mathrm{CH}_{2} \mathrm{Cl}_{2}$. The reaction mixture was stirred at room temperature for 3 days. The solvent volume was reduced by rotary evaporation, and the residue was poured into icewater. The aqueous layer was extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ three times. The residue upon drying and concentration of the organic layer was purified by flash chromatography ( $0-20 \%$ $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ in hexanes) to afford $2.6 \mathrm{~g}(78 \%)$ of mesylate $\mathbf{3 4 a}$ as a yellow oil: $R_{f} 0.76$ in $\mathrm{CH}_{2} \mathrm{Cl}_{2} ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 1.04-1.28(\mathrm{~m}, 21 \mathrm{H}), 3.25(\mathrm{~s}, 3 \mathrm{H}), 7.39-7.52(\mathrm{~m}, 3 \mathrm{H}), 7.94$ $(\mathrm{dd}, J=8.0,1.7 \mathrm{~Hz}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 11.01,18.50,36.71,93.37,111.53$, 127.52, 128.68, 130.89, 131.79, 148.58; HRMS m/z 380.1718 (calcd 380.1715, $\left.\mathrm{C}_{19} \mathrm{H}_{29} \mathrm{NOSSi}\right)$.

### 6.2.9 1-(4-Methoxyphenyl)-3-(triisopropylsilyl)-propynone mesylate 34b



To a solution of oxime $\mathbf{3 3 b}(0.77 \mathrm{~g}, 2.33 \mathrm{mmol})$ in 30 mL of dry $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ was added dry pyridine ( $0.94 \mathrm{~mL}, 11.6 \mathrm{mmol}$ ). The reaction mixture was cooled to $0{ }^{\circ} \mathrm{C}$, followed by dropwise addition of a solution of methanesulfonyl chloride ( $0.33 \mathrm{~g}, 2.92 \mathrm{mmol}$ ) in 5 mL of dry $\mathrm{CH}_{2} \mathrm{Cl}_{2}$. The reaction mixture was stirred at room temperature for 3 days. The solvent volume was reduced by rotary evaporation, and the residue was poured into icewater. The aqueous layer was extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ three times. The residue upon drying and concentration of the organic layer was purified by flash chromatography on silica gel $\left(0-30 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes) to afford $450 \mathrm{mg}(50 \%)$ of mesylate $\mathbf{3 4 b}$ as a yellow solid: mp 85.8-86.9 ${ }^{\circ} \mathrm{C} ; R_{f} 0.73\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 1.05-1.20(\mathrm{~m}, 21 \mathrm{H}), 3.20(\mathrm{~s}, 3 \mathrm{H})$, $3.81(\mathrm{~s}, 3 \mathrm{H}), 6.91(\mathrm{~d}, J=8.7 \mathrm{~Hz}, 2 \mathrm{H}), 7.87(\mathrm{~d}, J=8.7 \mathrm{~Hz}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta$ $10.98,18.46,36.59,55.35,93.53,110.61,114.05,123.27,129.19,148.20,162.53$; HRMS $m / z 410.1828$ (calcd 410.1821, $\mathrm{C}_{20} \mathrm{H}_{31} \mathrm{NO}_{4} \mathrm{SSi}$ ).

### 6.2.10 1-(2-Methoxyphenyl)-3-(triisopropylsilyl)-propynone mesylate 34c



To a solution of oxime $\mathbf{3 3 c}(2.7 \mathrm{~g}, 8.96 \mathrm{mmol})$ in 60 mL of dry $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ was added dry pyridine ( $3.6 \mathrm{~mL}, 44.8 \mathrm{mmol}$ ). The reaction mixture was cooled to $0^{\circ} \mathrm{C}$, followed by dropwise addition of a solution of methanesulfonyl chloride ( $1.28 \mathrm{~g}, 11.2 \mathrm{mmol}$ ) in 5 mL of dry $\mathrm{CH}_{2} \mathrm{Cl}_{2}$. The reaction mixture was stirred at room temperature for 3 days. The solvent volume was reduced by rotary evaporation, and the residue was poured into icewater. The aqueous layer was extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ three times. The residue upon drying and concentration of the organic layer was purified by flash chromatography ( $0-15 \%$ $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ in hexanes) to afford 90 mg of a yellow oil consisting of an inseparable mixture of mesylate 34c along with Beckman rearrangement product 3-(triisopropylsilyl)propynoic acid (2-methoxyphenyl)amide in 6 to 1 ratio, as judged by ${ }^{1} \mathrm{H}$ NMR. A pure sample of 3-(triisopropylsilyl)-propynoic acid (2-methoxyphenyl)amide was obtained as a yellow solid in $72 \%$ yield after chromatography from a reaction allowed to proceed for 6 days. Mesylate 34c: $R_{f} 0.65\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right) ;{ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 1.01-1.19(\mathrm{~m}, 21 \mathrm{H}), 3.18(\mathrm{~s}$, $3 \mathrm{H}), 3.83(\mathrm{~s}, 3 \mathrm{H}), 6.93-7.05(\mathrm{~m}, 2 \mathrm{H}), 7.43$ (ddd, $J=9.3,7.5,1.8 \mathrm{~Hz}, 1 \mathrm{H}), 7.55(\mathrm{dd}, J=$ $7.5,1.5 \mathrm{~Hz}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 10.93,18.38,36.57,55.55,94.39,110.12,111.55$, $120.45,120.59,130.64,132.43,147.87,157.91 ;$ HRMS $m / z 410.1825$ (calcd 410.1821, $\mathrm{C}_{20} \mathrm{H}_{31} \mathrm{NO}_{4} \mathrm{SSi}$ ). 3-(Triisopropylsilyl)-propynoic acid (2-methoxyphenyl)amide: $R_{f} 0.67$ $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right) ; \mathrm{mp} 32.0-32.9{ }^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 1.00-1.19(\mathrm{~m}, 21 \mathrm{H}), 3.88(\mathrm{~s}, 3 \mathrm{H}), 6.82-$ $6.98(\mathrm{~m}, 2 \mathrm{H}), 7.00-7.08(\mathrm{~m}, 1 \mathrm{H}), 8.05(\mathrm{~s}, 1 \mathrm{H}), 8.29(\mathrm{dd}, J=8.7,2.0,1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 10.98,18.45,55.76,89.10,100.31,109.86,120.21,121.08,124.32,127.15$, 147.43, 149.81; HRMS $m / z 332.2042$ (calcd 332.2045, $\mathrm{C}_{19} \mathrm{H}_{29} \mathrm{NO}_{2} \mathrm{Si}$ ).

### 6.2.11 t-Butyl $\mathbf{N}$-mesitylenesulfonyl oxycarbamate 36



TEA ( $1.9 \mathrm{~mL}, 13.7 \mathrm{mmol}$ ) was added dropwise to a mixture of 2-mesitylenesulfonyl chloride ( $3.0 \mathrm{~g}, 13.7 \mathrm{mmol}$ ) and $t$-butyl $N$-hydroxycarbamate ( $1.8 \mathrm{~g}, 13.7 \mathrm{mmol}$ ) in 40 mL dry $\mathrm{Et}_{2} \mathrm{O}$ at $0^{\circ} \mathrm{C}$. The resulting mixture was stirred at the same temperature for 55 min . Upon filtering off TEA hydrochloride salt, the organic layer was washed by water (20 $\mathrm{mL} x 2$ ). The residue upon drying and concentration of the organic layer was purified by flash chromatography ( $10 \%$ EtOAc in hexanes) to afford $3.9 \mathrm{~g}(90 \%)$ of oxycarbamate 36 as a white solid: $R_{f} 0.50$ (hexanes/EtOAc, 1:9); m.p $100.2-102.1{ }^{\circ} \mathrm{C}$, lit. ${ }^{4} 102-104{ }^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR ( $\left.\mathrm{CDCl}_{3}\right) \delta 1.28(\mathrm{~s}, 9 \mathrm{H}), 2.29(\mathrm{~s}, 3 \mathrm{H}), 2.64(\mathrm{~s}, 6 \mathrm{H}), 6.96(\mathrm{~s}, 2 \mathrm{H}), 7.80(\mathrm{br}, 1 \mathrm{H}, \mathrm{NH})$; ${ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 21.05,23.07,27.66,83.74,128.43,131.59,1414.88,144.37,154.27$; HRMS $m / z 316.1226$ (calc'd 316.1218, $\mathrm{C}_{14} \mathrm{H}_{21} \mathrm{NO}_{5} \mathrm{~S}$ ).

### 6.2.12 Mesitylenesulfonyl hydroxylamine 37



Trifluoroacetic acid ( 8 mL ) was added to $t$-Butyl $N$-mesitylenesulfonyloxycarbamate (2.0 g) at $5{ }^{\circ} \mathrm{C}$. The mixture was stirred at $5^{\circ} \mathrm{C}$ for $5-10 \mathrm{~min}$, and then poured into ice-water $(80 \mathrm{~mL})$. After filtration, the white solid was collected and dissolved in a minimum amount of $\mathrm{Et}_{2} \mathrm{O}(2 \mathrm{~mL})$. The white crystal was precipitated by the addition of petroleum ether $\left(30-60^{\circ} \mathrm{C}, 10 \mathrm{~mL}\right)$. The crystal was filtered to afford $0.99 \mathrm{~g}(73 \%)$ of compound $\mathbf{3 7}$ as white needles: ${ }^{5,6}{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 2.30(\mathrm{~s}, 3 \mathrm{H}), 2.61(\mathrm{~s}, 6 \mathrm{H}), 5.53\left(\mathrm{br}, 2 \mathrm{H} . \mathrm{NH}_{2}\right)$, 6.97 (s, 2H); ${ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 21.05,22.86,128.99,131.67,140.93,143.79$; HRMS $m / z 216.0689$ (calc'd 216.0694, $\mathrm{C}_{9} \mathrm{H}_{14} \mathrm{NO}_{3} \mathrm{~S}$ ).

### 6.2.13 1,3-Diphenylpropynone mesitylate 38d



To a solution of compound $\mathbf{3 2 d}(0.25 \mathrm{~g}, 1.21 \mathrm{mmol})$ in 6 mL dry $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ was added compound 37 ( $0.48 \mathrm{~g}, 2.32 \mathrm{mmol}$ ). The reaction mixture was stirred at room temperature for 30 h , and poured into ice-water. The aqueous layer was extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}(15 \mathrm{~mL} x$ 3). The residue upon drying and concentration of the organic layer was purified by flash chromatography ( $5 \%$ EtOAc in hexanes) to afford $0.34 \mathrm{~g}(70 \%)$ of mesitylate 38d as a white solid: $R_{f} 0.34\left(10 \%\right.$ EtOAc in hexanes); m.p 111.0-113.0 ${ }^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta$ 2.33 (s, 3H), 2.79 (s, 6H), 7.01 (s, 2H), 7.39-7.51 (m, 6H), 7.68 (dd, 2H, $J=7.8,1.5 \mathrm{~Hz}$ ), $7.87(\mathrm{dd}, 2 \mathrm{H}, J=7.8,1.5 \mathrm{~Hz}){ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 20.99,22.77,77.79,104.29,120.57$, $127.29,128.51,128.59,130.32,130.47,131.29,131.40,131.61,132.44,140.70,143.69$, 147.59; HRMS $m / z 404.1304$ (calc'd 404.1320, $\mathrm{C}_{24} \mathrm{H}_{22} \mathrm{NO}_{3} \mathrm{~S}$ ).

### 6.2.14 1,3-diphenyl-1- ${ }^{13} \mathrm{C}$-propynone mesitylate 38 d



To a solution of 1,3 -diphenyl-1- ${ }^{13}$ C-propynone $\mathbf{3 2 d}(0.11 \mathrm{~g}, 0.53 \mathrm{mmol})$ in 4 mL dry $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ was added mesitylenesulfonyl hydroxyamine 37 ( $0.30 \mathrm{~g}, 1.39 \mathrm{mmol}$ ). The reaction mixture was stirred at room temperature for 48 h , and poured into ice-water. The aqueous layer was extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}(15 \mathrm{~mL} x 3)$. The residue upon drying and concentration of the organic layer was purified by flash chromatography (5\% EtOAc in hexanes) to afford $0.15 \mathrm{~g}(70 \%)$ of mesitylate $\mathbf{3 8 d}$ as a white solid: $R_{f} 0.34(10 \% \mathrm{EtOAc}$ in hexanes); m.p $112.0-114.0{ }^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 2.29(\mathrm{~s}, 3 \mathrm{H}), 2.77(\mathrm{~s}, 6 \mathrm{H}), 6.97(\mathrm{~s}$, $2 \mathrm{H})$, 7.37-7.49 (m, 6H), $7.64(\mathrm{dd}, 2 \mathrm{H}, J=7.8,1.5 \mathrm{~Hz}), 7.82-7.88(\mathrm{~m}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 20.99,22.77,77.78(\mathrm{~d}, J=95.1 \mathrm{~Hz}), 104.28(\mathrm{~d}, J=11.5 \mathrm{~Hz}), 120.56(\mathrm{~d}, J=$ $2.2 \mathrm{~Hz}), 127.29(\mathrm{~d}, J=2.2 \mathrm{~Hz}), 128.51(\mathrm{~d}, J=5.0 \mathrm{~Hz}), 128.59,130.32,130.47,131.27$ $(\mathrm{d}, J=63.8 \mathrm{~Hz}), 131.40,131.61,132.44,140.70,143.69,147.59\left({ }^{13} \mathrm{C}\right) ;$ HRMS $\mathrm{m} / \mathrm{z}$ 405.1354 (calc'd $405.1354,{ }^{12} \mathrm{C}_{23}{ }^{13} \mathrm{CH}_{22} \mathrm{NO}_{3} \mathrm{~S}$ ).

### 6.2.15 1,4-Diphenyl-6-(triisopropylsilyl)-3-aza-3-ene-1,5-diyne 26a



To a solution of phenylacetylene ( $410 \mathrm{mg}, 4.02 \mathrm{mmol}$ ) in 5 mL of dry ether at $-20^{\circ} \mathrm{C}$ was added dropwise a solution of $n$-butyllithium ( $2.89 \mathrm{~mL}, 1.6 \mathrm{M}$ in hexanes, 4.02 mmol ) such that the temperature was maintained at $-20^{\circ} \mathrm{C}$. The reaction mixture was stirred for 20 min at the same temperature. In a separate flask, a suspension of cuperous iodide (251 $\mathrm{mg}, 1.32 \mathrm{mmol}$ ) in 5 mL of dry ether was cooled to $-40^{\circ} \mathrm{C}$. The phenylacetylide solution was added slowly via cannula to this suspension. After the addition was complete, the mixture was stirred at room temperature for 40 min . The resulting light yellow suspension was added to a solution of mesylate 38a ( $500 \mathrm{mg}, 1.32 \mathrm{mmol}$ ) in 5 mL of dry ether at $0{ }^{\circ} \mathrm{C}$. The reaction mixture was stirred at $0{ }^{\circ} \mathrm{C}$ for 1 h , and then at room temperature for 2 h . The solvent was evaporated, and the product was purified by flash chromatography on silica gel (hexanes) to afford $110 \mathrm{mg}(26 \%)$ of aza-enediyne 26a as a yellow oil: $R_{f} 0.56\left(40 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes); ${ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 1.10-1.25(\mathrm{~m}, 21 \mathrm{H})$, 7.28-7.39 (m, 3H), 7.39-7.55 (m, 5H), $8.14(\mathrm{dd}, J=7.6,1.7 \mathrm{~Hz}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right)$ $\delta 11.55,18.79,93.75,100.27,100.60,106.79,124.90,128.26,128.52,128.66,128.92$, 131.77, 132.49, 136.22, 158.96; HRMS $m / z 386.2310$ (calcd 386.2304, $\mathrm{C}_{26} \mathrm{H}_{31} \mathrm{NSi}$ ).

### 6.2.16 1-Phenyl-4-(4-methoxyphenyl)-6-(triisopropylsilyl)-3-aza-3-ene-1,5-diyne 26b



To a solution of phenylacetylene ( $78 \mathrm{mg}, 0.76 \mathrm{mmol}$ ) in 2 mL of dry ether at $-20^{\circ} \mathrm{C}$ was added dropwise a solution of $n$-butyllithium ( $0.64 \mathrm{~mL}, 1.6 \mathrm{M}$ in hexanes, 0.76 mmol ) such that the temperature was maintained at $-20^{\circ} \mathrm{C}$. The reaction mixture was stirred for 20 min at the same temperature. In a separate flask, a suspension of cuperous iodide (48 $\mathrm{mg}, 0.25 \mathrm{mmol}$ ) in 2 mL of dry ether was cooled to $-40^{\circ} \mathrm{C}$. The phenylacetylide solution was added slowly via cannula to this suspension. After the addition was complete, the mixture was stirred at room temperature for 40 min . The resulting light yellow suspension was added to a solution of mesylate $\mathbf{3 8 b}(100 \mathrm{mg}, 0.24 \mathrm{mmol})$ in 2 mL of dry ether at $0{ }^{\circ} \mathrm{C}$. The reaction mixture was stirred at $0{ }^{\circ} \mathrm{C}$ for 1 h , and then at room temperature for 1 h . The solvent was evaporated, and the product was purified by preparative TLC on silica gel (hexanes) to afford 12 mg (15\%) of aza-enediyne $\mathbf{2 6 b}$ as a yellow oil: $R_{f} 0.35\left(30 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes); ${ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 1.08-1.25(\mathrm{~m}, 21 \mathrm{H})$, $3.85(\mathrm{~s}, 3 \mathrm{H}), 6.92(\mathrm{~d}, J=9.0 \mathrm{~Hz}, 2 \mathrm{H}), 7.25-7.34(\mathrm{~m}, 3 \mathrm{H}), 7.44(\mathrm{dd}, J=8.4,1.5 \mathrm{~Hz}, 2 \mathrm{H})$, $8.08(\mathrm{~d}, J=9.0 \mathrm{~Hz}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 11.16,18.65,55.47,93.60,97.86,100.00$, $105.35,113.94,124.90,127.82,128.14,128.99,129.86,131.38,131.92,163.00$; HRMS $m / z 416.2407$ (calcd 416.2409, $\mathrm{C}_{27} \mathrm{H}_{33} \mathrm{NOSi}$ ).

### 6.2.17 1-Phenyl-4-(2-methoxyphenyl)-6-(triisopropylsilyl)-3-aza-3-ene-1,5-diyne 26c



To a solution of phenylacetylene ( $410 \mathrm{mg}, 4.02 \mathrm{mmol}$ ) in 5 mL of dry ether at $-20^{\circ} \mathrm{C}$ was added dropwise a solution of $n$-butyllithium ( $2.89 \mathrm{~mL}, 1.6 \mathrm{M}$ in hexanes, 4.02 mmol ) such that the temperature was maintained at $-20^{\circ} \mathrm{C}$. The reaction mixture was stirred for 20 min at the same temperature. In a separate flask, a suspension of cuperous iodide (251 $\mathrm{mg}, 1.32 \mathrm{mmol}$ ) in 5 mL of dry ether was cooled to $-40^{\circ} \mathrm{C}$. The phenylacetylide solution was added slowly via cannula to this suspension. After the addition was complete, the mixture was stirred at room temperature for 40 min . The resulting light yellow suspension was added to a solution of mesylate $\mathbf{3 8 c}(500 \mathrm{mg}, 1.32 \mathrm{mmol})$ in 5 mL of dry ether at $0{ }^{\circ} \mathrm{C}$. The reaction mixture was stirred at $0{ }^{\circ} \mathrm{C}$ for 1 h , and then at room temperature for 2 h . The solvent was evaporated, and the product was purified by flash chromatography ( $0-20 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}$ in hexanes) to afford $22 \mathrm{mg}(31 \%)$ of aza-enediyne $\mathbf{2 6 c}$ as a yellow oil: $R_{f} 0.45\left(40 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes); ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 1.05-1.19(\mathrm{~m}$, $21 \mathrm{H}), 3.88(\mathrm{~s}, 3 \mathrm{H}), 6.95-7.02(\mathrm{~m}, 2 \mathrm{H}), 7.26-7.34(\mathrm{~m}, 3 \mathrm{H}), 7.36-7.48(\mathrm{~m}, 3 \mathrm{H}), 7.86(\mathrm{~d}, J=$ $7.6,1.5 \mathrm{~Hz}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 11.19,18.60,55.70,93.21,100.66,101.82,105.05$, $111.77,120.44,124.86,125.99,127.77,128.11,130.81,131.27,132.64,157.66,158.60 ;$ HRMS m/z 416.2404 (calcd 416.2409, $\mathrm{C}_{27} \mathrm{H}_{33} \mathrm{NOSi}$ ).

### 6.2.18 1,4,6-triphenyl-3-aza-3-ene-1,5-diyne 26d



To a solution of phenyl acetylene ( $262 \mathrm{mg}, 2.57 \mathrm{mmol}$ ) in 3 mL of dry $\mathrm{Et}_{2} \mathrm{O}$ at $-20^{\circ} \mathrm{C}$ was added dropwise a solution of $n$-butyllithium ( $2.57 \mathrm{~mL}, 1.0 \mathrm{M}$ in hexanes, 2.67 mmol ) such that the temperature was maintained at $-20^{\circ} \mathrm{C}$. The reaction mixture was stirred for 20 min at the same temperature. In a separate flask, a suspension of cuperous iodide (161 $\mathrm{mg}, 0.85 \mathrm{mmol}$ ) in 3 mL of dry $\mathrm{Et}_{2} \mathrm{O}$ was cooled to $-40^{\circ} \mathrm{C}$. The phenylacetylide solution was added slowly via cannula to this suspension. After the addition was complete, the mixture was stirred at room temperature for 40 min . The resulting light yellow suspension was added to a solution of mesitylate $\mathbf{3 8 d}(280 \mathrm{mg}, 0.51 \mathrm{mmol})$ in 3 mL of dry $\mathrm{Et}_{2} \mathrm{O}$ at $0{ }^{\circ} \mathrm{C}$. The reaction mixture was stirred at $0{ }^{\circ} \mathrm{C}$ for 1 h , and then at room temperature for 24 h . The solvent was evaporated, and the product was purified by flash chromatography on silica gel (hexanes) to afford 76 mg (50\%) of aza-enediyne $\mathbf{2 6 d}$ as a yellow oil: $R_{f} 0.68\left(50 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes); ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.31-7.58(\mathrm{~m}, 11 \mathrm{H})$, $7.70(\mathrm{dd}, 2 \mathrm{H}, J=7.5,1.5 \mathrm{~Hz}), 8.22(\mathrm{dd}, 2 \mathrm{H}, J=7.5,1.5 \mathrm{~Hz}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 84.36$, 93.94, $99.83,102.07,121.14,124.70,127.97,128.24,128.40,128.55,128.68,130.33$, $131.60,132.09,132.61,135.82,159.03 ;$ HRMS $m / z 306.1272\left(\right.$ calcd 306.1282, $\left.\mathrm{C}_{23} \mathrm{H}_{16} \mathrm{~N}\right)$.

### 6.2.19 1,4, 6-triphenyl-4- ${ }^{13}$ C-3-aza-3-ene-1,5-diyne 26d



To a solution of phenyl acetylene ( $124 \mathrm{mg}, 1.22 \mathrm{mmol}$ ) in 2 mL of dry $\mathrm{Et}_{2} \mathrm{O}$ at $-20^{\circ} \mathrm{C}$ was added dropwise a solution of $n$-butyllithium ( $0.87 \mathrm{~mL}, 1.6 \mathrm{M}$ in hexanes, 1.22 mmol ) such that the temperature was maintained at $-20^{\circ} \mathrm{C}$. The reaction mixture was stirred for 20 min at the same temperature. In a separate flask, a suspension of cuperous iodide (70 $\mathrm{mg}, 0.37 \mathrm{mmol}$ ) in 2 mL of dry $\mathrm{Et}_{2} \mathrm{O}$ was cooled to $-40^{\circ} \mathrm{C}$. The phenylacetylide solution was added slowly via cannula to this suspension. After the addition was complete, the mixture was stirred at room temperature for 40 min . The resulting light yellow suspension was added to a solution of mesitylate $\left[1-{ }^{13} \mathrm{C}\right]-\mathbf{3 8 d}(140 \mathrm{mg}, 0.34 \mathrm{mmol})$ in 2 mL of dry $\mathrm{Et}_{2} \mathrm{O}$ at $0^{\circ} \mathrm{C}$. The reaction mixture was stirred at $0^{\circ} \mathrm{C}$ for 1 h , and then at room temperature for 20 h . The solvent was evaporated, and the product was purified by flash chromatography on silica gel (hexanes) to afford $55 \mathrm{mg}(53 \%)$ of aza-enediyne $\left[4-{ }^{13} \mathrm{C}\right]$ 26d as a red-yellow oil: $R_{f} 0.70\left(50 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes $) ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta$ 7.29-7.57 $(\mathrm{m}, 11 \mathrm{H}), 7.69(\mathrm{dd}, 2 \mathrm{H}, J=8.1,1.8 \mathrm{~Hz}), 8.18-8.23(\mathrm{~m}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 84.34$ $(\mathrm{d}, J=91.2 \mathrm{~Hz}), 93.94(\mathrm{~d}, J=6.6 \mathrm{~Hz}), 99.83(\mathrm{~d}, J=7.0 \mathrm{~Hz}), 102.06(\mathrm{~d}, J=10.4 \mathrm{~Hz})$, $121.13(\mathrm{~d}, J=2.2 \mathrm{~Hz}), 124.70(\mathrm{~d}, J=2.2 \mathrm{~Hz}), 127.96(\mathrm{~d}, J=2.8 \mathrm{~Hz}), 128.23,128.40$, $128.54(\mathrm{~d}, J=5.0 \mathrm{~Hz}), 128.68,130.32,131.60(\mathrm{~d}, J=1.7 \mathrm{~Hz}), 132.07,132.61,135.79(\mathrm{~d}$, $J=67.6 \mathrm{~Hz}), 159.03\left({ }^{13} \mathrm{C}\right)$; HRMS $m / z 307.1323\left(\operatorname{calcd} 307.1316,{ }^{12} \mathrm{C}_{22}{ }^{13} \mathrm{CH}_{16} \mathrm{~N}\right)$.

### 6.2.20 1-(Methylphenyl)-4,6-diphenyl-3-ene-3-aza-1,5-diyne 26g



To a solution of 4-methylphenyl acetylene ( $152 \mathrm{mg}, 1.30 \mathrm{mmol}$ ) in 3 mL of dry $\mathrm{Et}_{2} \mathrm{O}$ at $20^{\circ} \mathrm{C}$ was added dropwise a solution of $n$-butyllithium $(1.04 \mathrm{~mL}, 1.6 \mathrm{M}$ in hexanes, 1.30 mmol ) such that the temperature was maintained at $-20^{\circ} \mathrm{C}$. The reaction mixture was stirred for 20 min at the same temperature. In a separate flask, a suspension of cuperous iodide ( $82 \mathrm{mg}, 0.43 \mathrm{mmol}$ ) in 3 mL of dry $\mathrm{Et}_{2} \mathrm{O}$ was cooled to $-40{ }^{\circ} \mathrm{C}$. The phenylacetylide solution was added slowly via cannula to this suspension. After the addition was complete, the mixture was stirred at room temperature for 40 min . The resulting light yellow suspension was added to a solution of mesitylate $\mathbf{3 8 d}$ ( $165 \mathrm{mg}, 0.41$ $\mathrm{mmol})$ in 3 mL of dry $\mathrm{Et}_{2} \mathrm{O}$ at $0^{\circ} \mathrm{C}$. The reaction mixture was stirred at $0^{\circ} \mathrm{C}$ for 1 h , and then at room temperature for 20 h . The solvent was evaporated, and the product was purified by flash chromatography on silica gel (hexanes) to afford $67 \mathrm{mg}(55 \%)$ of azaenediyne $\mathbf{2 6 g}$ as a yellow oil: $R_{f} 0.70\left(50 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes); ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 2.38$ (s, 3H), 7.17 (d, 2H, $J=8.1 \mathrm{~Hz}$ ), 7.42-7.4 (m, 8H), 7.69 (dd, 2H, $J=8.1,1.5$ ), 8.20 (dd, $2 \mathrm{H}, J=8.1,1.5) ;{ }^{13} \mathrm{C} \mathrm{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 21.53,84.40,93.69,100.42,101.86,121.22$, $121.65,127.88,128.52,128.66,129.20,130.25,131.60,131.94,132.59,135.89,138.54$, 158.44; HRMS m/z 320.1445 (calcd 320.1439, $\mathrm{C}_{24} \mathrm{H}_{18} \mathrm{~N}$ ).

### 6.2.21 (Z)-2,5-Diphenyl-pent-2-en-4-ynenitrile 28a



A solution of $\mathbf{2 6 a}(68 \mathrm{mg}, 0.176 \mathrm{mmol})$ in dry THF $(6 \mathrm{~mL})$ was cooled to $-78^{\circ} \mathrm{C}$ and then a 1 M solution of TBAF ( 0.194 mmol ) was added slowly. After being stirred at $-78{ }^{\circ} \mathrm{C}$ for 5-10 min the mixture was poured into ice-water $(10 \mathrm{~mL})$ and extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}(2 \times 15$ $\mathrm{mL})$. The dry organic solution was plugged through a small column before evaporation. The conversion of aza-enediyne 40a to (Z)-nitrile 28a was followed by ${ }^{1} \mathrm{H}$ NMR and/or UV-vis spectroscopy. After the conversion was complete, the ( $Z$ )-nitrile 28a was purified by flash chromatography on silica gel $\left(0-10 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes) to afford $36 \mathrm{mg}(89 \%)$ of nitrile 28a as a yellow solid: $R_{f} 0.41\left(50 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes); mp 79.3-80.7 ${ }^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CD}_{2} \mathrm{Cl}_{2}\right) \delta 6.92(\mathrm{~s}, 1 \mathrm{H}), 7.38-7.50(\mathrm{~m}, 6 \mathrm{H}), 7.55-7.67(\mathrm{~m}, 4 \mathrm{H}) ;{ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CD}_{3} \mathrm{CN}\right)$ $\delta 7.1(\mathrm{~s}, 1 \mathrm{H}), 7.41-7.50(\mathrm{~m}, 6 \mathrm{H}), 7.54-7.61(\mathrm{~m}, 2 \mathrm{H}), 7.65-7.71(\mathrm{~m}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CD}_{2} \mathrm{Cl}_{2}\right) \delta 86.45,102.23,116.90,121.87,122.43,124.09,126.03,128.99,129.54$, $130.15,130.52,132.48,133.04 ;$ HRMS $m / z 230.0972\left(\right.$ calcd $\left.230.0969, \mathrm{C}_{17} \mathrm{H}_{11} \mathrm{~N}\right)$.

### 6.2.22 (Z)-5-(4-Methoxyphenyl)-2-phenyl-pent-2-en-4-ynenitrile 28b



A solution of $\mathbf{2 6 b}(9 \mathrm{mg}, 0.022 \mathrm{mmol})$ in dry THF ( 2 mL ) was cooled to $-78{ }^{\circ} \mathrm{C}$ and then a 1 M solution of TBAF $(0.024 \mathrm{mmol})$ was added slowly. After being stirred at $-78^{\circ} \mathrm{C}$ for 5-10 min the mixture was poured into ice-water $(10 \mathrm{~mL})$ and extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}(2 \times 10$ mL ). The dry organic solution was plugged through a small column before evaporation. The conversion of imine $\mathbf{4 0 b}$ to nitrile $\mathbf{2 8 b}$ was followed by ${ }^{1} \mathrm{H}$ NMR and/or UV-vis. The nitrile 28b was purified by flash chromatography on silica gel $\left(0-5 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes) to afford $4 \mathrm{mg}(72 \%)$ of ( $Z$ )-nitrile 28b as a light yellow solid: $R_{f} 0.35\left(40 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes); mp 85.5-86.6 ${ }^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 3.85(\mathrm{~s}, 3 \mathrm{H}), 6.84(\mathrm{~s}, 1 \mathrm{H}), 6.88(\mathrm{~d}, J=$ $8.9 \mathrm{~Hz}, 2 \mathrm{H}), 7.36-7.44(\mathrm{~m}, 3 \mathrm{H}), 7.52(\mathrm{~d}, J=8.9 \mathrm{~Hz}, 2 \mathrm{H}), 7.57-7.62(\mathrm{dd}, J=7.7,1.5 \mathrm{~Hz}$, 2H); HRMS $m / z 260.1070$ (calcd 260.1075, $\mathrm{C}_{18} \mathrm{H}_{13} \mathrm{NO}$ ).

### 6.2.23 (Z)-5-(2-Methoxyphenyl)-2-phenyl-pent-2-en-4-ynenitrile 28c



A solution of $\mathbf{2 6 c}(68 \mathrm{mg}, 0.176 \mathrm{mmol})$ in dry THF $(6 \mathrm{~mL})$ was cooled to $-78^{\circ} \mathrm{C}$ and then a 1 M solution of TBAF $(0.194 \mathrm{mmol})$ was added slowly. After being stirred at $-78^{\circ} \mathrm{C}$ for 5-10 min the mixture was poured into ice-water $(10 \mathrm{~mL})$ and extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}(2 \times 15$ $\mathrm{mL})$. The dry organic solution was plugged through a small column before evaporation. The conversion of imine $\mathbf{4 0 c}$ to nitrile 28c was followed by ${ }^{1} \mathrm{H}$ NMR and/or UV-vis. The nitrile 28c was purified by flash chromatography $\left(0-10 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes) to afford 10 $\mathrm{mg}(91 \%)$ of ( $Z$ )-nitrile 28c as a light yellow solid: $R_{f} 0.30\left(40 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes); mp 57.7-58.6 ${ }^{\circ} \mathrm{C}$; ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 3.92(\mathrm{~s}, 3 \mathrm{H}), 6.88-6.98(\mathrm{~m}, 3 \mathrm{H}), 7.33-7.48(\mathrm{~m}, 4 \mathrm{H})$, 7.50-7.57 (dd, $J=7.8,1.8 \mathrm{~Hz}, 1 \mathrm{H}), 7.58-7.62(\mathrm{~m}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 55.87,90.44$, $99.41,110.74,111.21,120.65,120.82,121.56,122.95,125.61,129.14,129.94,131.41$, 132.50, 134.27, 160.54; HRMS m/z 260.1077 (calcd 260.1075, $\mathrm{C}_{18} \mathrm{H}_{13} \mathrm{NO}$ ).

### 6.2.24 (E)-2,5-Diphenyl-3-(triisopropylsilyl)-pent-2-ene-4-ynenitrile 39



To a solution of the aza-enediyne $\mathbf{2 6 a}(20 \mathrm{mg}, 0.0518 \mathrm{mmol})$ in chlorobenzene $(1.5 \mathrm{~mL})$ was added 1,4-cyclohexadiene ( $83.19 \mathrm{mg}, 1.036 \mathrm{mmol}, 20$ equiv). The mixture was heated at $150{ }^{\circ} \mathrm{C}$ for 3 days. The solvent was evaporated and the residue purified by chromatography to afford nitrile $39(7 \mathrm{mg}, 35 \%)$ as a yellow oil: $R_{f} 0.51\left(40 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes); ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 1.10-1.22(\mathrm{~d}, J=7.5 \mathrm{~Hz}, 18 \mathrm{H}), 1.68-1.85(\mathrm{p}, J=7.5 \mathrm{~Hz}$, $3 \mathrm{H}), 7.24-7.34(\mathrm{~m}, 5 \mathrm{H}), 7.37-7.48(\mathrm{~m}, 3 \mathrm{H}), 7.84(\mathrm{dd}, J=7.7,1.9,2 \mathrm{H}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right)$ $\delta 12.27,18.78,91.31,108.25,120.38,122.98,128.12,128.51,128.70,129.23,129.42$, 131.44, 134.10, 136.07, 139.69; HRMS $m / z 386.2317$ (calcd 386.2304, $\mathrm{C}_{26} \mathrm{H}_{31} \mathrm{NSi}$ ).
6.2.25 (E) and (Z)-1,3,4,6-tetraphenyl-3-hexene-1,5-diynes 60 d and (E)-1,2-dicyano-

## 1,2-diphenylethylene 61





A benzene solution ( 2.5 mL ) of compound $\mathbf{2 6 d}(90 \mathrm{mg}, 0.29 \mathrm{mmol})$ was stirred at $55^{\circ} \mathrm{C}$ for 5 days under argon. The residue upon concentration of the organic layer was purified by flash chromatography to afford $38 \mathrm{mg}(70 \%)$ of $(Z) \mathbf{- 6 0 d}$ and $(E)-\mathbf{6 0 d}$ mixtures (1:1 ratio judged by ${ }^{1} \mathrm{H}$ NMR) as a yellow solid and $15 \mathrm{mg}(45 \%)$ of ( $E$ )-dicyano compound 61 as a white solid. Re-crystallization of the mixtures of $(Z)$ and $(E)$ tetraphenyl enediynes 60 d from hexanes/EtOAc afforded pure $(E)$-isomer (purity $>95 \%$ as judged by ${ }^{1} \mathrm{H}$ NMR) and ( $Z$ )-isomer rich fraction (containing $20 \%$ of $(E)$-isomer). ( $E$ )-60d: yellow prisms from hexanes/EtOAc; $R_{f} 0.65$ (10\% EtOAc in hexanes); m.p. $155.0-157.0^{\circ} \mathrm{C}$, lit. ${ }^{7}$ $156.2-157.2{ }^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.23-7.25(\mathrm{~m}, 10 \mathrm{H}), 7.39(\mathrm{t}, 2 \mathrm{H}, J=8.0 \mathrm{~Hz}), 7.45(\mathrm{t}$, $4 \mathrm{H}, J=8.0 \mathrm{~Hz}), 7.97(\mathrm{dd}, 4 \mathrm{H}, J=8.0,1.2 \mathrm{~Hz}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 90.90,98.53,123.21$, $127.83,128.28,128.45,128.60,129.23,131.40,139.01$; $(Z)$-60d: yellow needles from hexanes/EtOAc, after 3 times of re-crystallization, there is still at least $\mathbf{2 0 \%}$ of $(E) \mathbf{- 6 0 d}$ in the sample; $R_{f} 0.65\left(10 \%\right.$ EtOAc in hexanes); m.p. 96.0-99.0 ${ }^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta$ 7.18-7.21 (m, 6H), 7.26-7.32 (m, 10H), 7.52-7.55 (m, 4H); Mixture of (E)-60d and (Z)60d (1:1): yellow solid; $R_{f} 0.65$ ( $10 \%$ EtOAc in hexanes); m.p. $95.0-99.0{ }^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.19-7.23(\mathrm{~m}, 6 \mathrm{H}), 7.26-7.40(\mathrm{~m}, 20 \mathrm{H}), 7.39(\mathrm{t}, 2 \mathrm{H}, J=8.0 \mathrm{~Hz}), 7.45(\mathrm{t}, 4 \mathrm{H}, J$ $=8.0 \mathrm{~Hz}), 7.53-7.57(\mathrm{~m}, 4 \mathrm{H}), 7.97(\mathrm{dd}, 4 \mathrm{H}, J=8.0,1.2 \mathrm{~Hz}) ;{ }^{13} \mathrm{C}$ NMR $\delta 90.90,91.77$, 96.84, $98.53,123.21,123.37,127.83,128.04,128.28,128.36,128.45,128.52,128.60$, 129.23, 129.72, 131.40, 131.65, 137.45, 139.01; HRMS m/z (60d) 381.1636 (calcd 381.1643, $\mathrm{C}_{30} \mathrm{H}_{21}$ ); (E)-61: white needle from hexanes; $R_{f} 0.40$ ( $10 \%$ EtOAc in hexanes);
m.p. 161.5-162.5 ${ }^{\circ} \mathrm{C}$, lit. $.^{8} 161-162{ }^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.50-7.55(\mathrm{~m}, 6 \mathrm{H}), 7.80-7.84$ (m, 4H); ${ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 116.6,125.57,128.67,129.27,131.70,131.95 ;$ HRMS $m / z$ 231.0924 (calcd 231.0922, $\mathrm{C}_{16} \mathrm{H}_{11} \mathrm{~N}_{2}$ ).

### 6.2.26 2,3,5-Triphenyl-pent-2-en-4-ynenitrile 28d



The solution of aza-enediyne $\mathbf{2 6 d}(22 \mathrm{mg}$ ) in benzene ( 0.45 mM ), kept in dark, was stirred at $90^{\circ} \mathrm{C}$ for 24 hours. The solvent was evaporated, and the product was purified by flash chromatography on silica gel (1\% EtOAc in hexanes) to afford $18.6 \mathrm{mg}(85 \%)$ of nitrile 28d as a light-yellow solid: $R_{f} 0.50\left(10 \%\right.$ EtOAc in hexanes); mp: $132-134{ }^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.22-7.40(\mathrm{~m}, 13 \mathrm{H}), 7.62-7.66(\mathrm{~m}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 89.47$, $100.48,118.24,119.24,121.91,128.49,128.60,129.05,129.42,129.48,129.65,132.23$, 132.87, 135.32, $137.34,138.26 ;$ FT-IR: $2185 \mathrm{~cm}^{-1}$; HRMS m/z 306.1284 (calcd 306.1283, $\left.\mathrm{C}_{23} \mathrm{H}_{16} \mathrm{~N}\right)$.

### 6.2.27 $\left[1,4-{ }^{13} \mathrm{C}_{2}\right]-60 \mathrm{~d}$, and $\left[5-{ }^{13} \mathrm{C}\right]-28 \mathrm{~d}$





A benzene solution ( 1.1 mL ) of compound $\mathbf{2 6 d}(42 \mathrm{mg}, 0.14 \mathrm{mmol})$ was stirred at $70{ }^{\circ} \mathrm{C}$ for 2 days under argon. The residue upon concentration of the organic layer was purified by flash chromatography to afford $16 \mathrm{mg}(60 \%)$ of $(Z),(E)-\left[1,4-{ }^{13} \mathrm{C}_{2}\right]-60 \mathrm{~d}$ mixtures (1:3 ratio judged by ${ }^{1} \mathrm{H}$ NMR) as a yellow solid, $6 \mathrm{mg}(35 \%)$ of ( $E$ )-dicyano compound $\mathbf{6 1}$ as a white solid and $10 \mathrm{mg}(24 \%)$ of nitrile $\left[5-{ }^{13} \mathrm{C}\right]-\mathbf{2 8 d}$ as yellow solid. $(E)-\left[1,4-{ }^{13} \mathrm{C}_{2}\right]-\mathbf{6 0 d}$ : ${ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 96.84\left({ }^{13} \mathrm{C}\right), 129.23\left({ }^{13} \mathrm{C}\right) ;(Z)-\left[1,4-{ }^{13} \mathrm{C}_{2}\right]-60 \mathrm{~d}:{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta$ $98.56\left({ }^{13} \mathrm{C}\right), 128.63\left({ }^{13} \mathrm{C}\right)$; HRMS $m / z 383.1708$ (calcd 383.1710); [5- $\left.{ }^{13} \mathrm{C}\right]-28 d$ : ${ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 100.48\left({ }^{13} \mathrm{C}\right)$; HRMS m/z 307.1314 (calcd 307.1316, ${ }^{12} \mathrm{C}_{22}{ }^{13} \mathrm{CH}_{16} \mathrm{~N}$ ).

### 6.2.28 (E), (Z)-3,6-Dimethyl-1,4-diphenyl-3-hexene-1,5-diynes 60 e



Flash chromatography of the samples of aza-enediyne $\mathbf{2 6 e}(20 \mathrm{mg})$ stored in a $-10{ }^{\circ} \mathrm{C}$ freezer for 24 months affords $1.2 \mathrm{mg}(14 \%)$ of (Z), (E)-60e (1:1 ratio as judged by ${ }^{1} \mathrm{H}$ NMR) as a yellow solid, 2.8 mg ( $35 \%$ ) of ( $E$ )-1,2-dicyano-1,2-diphenylethylene $\mathbf{6 1}$ as a white solid. (Z), (E)-60e: $R_{f} 0.60\left(10 \%\right.$ EtOAc in hexanes); ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 2.01$ (s, $3 \mathrm{H}), 2.08(\mathrm{~s}, 3 \mathrm{H}), 2.09(\mathrm{~s}, 3 \mathrm{H}), 2.30(\mathrm{~s}, 3 \mathrm{H}), 7.23-7.36(\mathrm{~m}, 16 \mathrm{H}), 7.49-7.51(\mathrm{~m}, 2 \mathrm{H}), 7.77$ (d, $2 \mathrm{H}, J=7.6 \mathrm{~Hz}$ ); HRMS $m / z 257.1323$ (calcd 257.1330, $\mathrm{C}_{20} \mathrm{H}_{17}$ ).

### 6.2.29 2-(Methylphenyl)-3,5-diphenyl-pent-2-en-4-ynenitrile 28g (E)-1,2-dicyano-1,2-di-(methylphenyl)ethylene 62




A benzene solution ( 1.4 mL ) of compound $\mathbf{2 6 g}(48 \mathrm{mg}, 0.14 \mathrm{mmol})$ was stirred at $50{ }^{\circ} \mathrm{C}$ for 6 days under argon. The residue upon concentration of the organic layer was purified by flash chromatography to afford $12.5 \mathrm{mg}(47 \%)$ of $(Z),(E)$ - $\mathbf{6 0 d}$ mixtures (1:1 ratio judged by ${ }^{1} \mathrm{H}$ NMR) as a yellow solid, 16.5 mg of the mixtures of $(E)$-dicyano compound $62\left(26 \%\right.$, based on ${ }^{1} \mathrm{H}$ NMR) and nitrile 28d ( $14 \%$, based on ${ }^{1} \mathrm{H}$ NMR) as light yellow solid. The small amount of pure $\mathbf{6 2}$ was obtained by re-crystallization of the mixtures from hexanes and ethyl acetate. 62: ${ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 2.42(\mathrm{~s}, 6 \mathrm{H}), 7.31(\mathrm{~d}, J=7.5 \mathrm{~Hz}$, $4 \mathrm{H}), 7.71(\mathrm{~d}, J=8.1 \mathrm{~Hz}, 4 \mathrm{H})$; CIMS $m / z 259\left(\mathrm{MH}^{+}\right)$.

### 6.2.30 2-Benzyl-5-bicyclo[4.1.0]hept-3-en-7-yl-4-phenyl-oxazole 44



To a solution of 1,4-diphenyl-6-(triisopropylsilyl)-3-aza-3-ene-1,5-diyne 26a ( 60 mg , 0.156 mmol ) in 5 ml dry THF cooled to $-78^{\circ} \mathrm{C}$ was added slowly 1 M solution of TBAF ( 0.172 mmol ). After stirred at $-78^{\circ} \mathrm{C}$ for $5-10 \mathrm{~min}$, the mixture was poured into ice-water $(10 \mathrm{ml})$ and extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}(2 \times 15 \mathrm{ml})$. The dry organic solution was plugged through a small column before evaporation. The residue was further dried under vacuum for $10-15 \mathrm{~min}$, and then re-dissolved in 4 ml dry $\mathrm{CH}_{2} \mathrm{Cl}_{2}$. To this red solution cooled to $78^{\circ} \mathrm{C}$ was added 20 equiv freshly distilled $1,4-\mathrm{CHD}$, followed by 0.1 equiv trifluoromethane sulfonic acid. The mixture was stirred at $-78^{\circ} \mathrm{C}$ for 30 min , and then at $10{ }^{\circ} \mathrm{C}$ for 4 days. The oxazole derivative 44 and nitrile $\mathbf{2 8 a}$ were purified by flash chromatography on silica gel $\left(0-30 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes) to afforded 15 mg oxazole 44 as white solid: $R_{f}=0.25\left(50 \% \mathrm{CH}_{2} \mathrm{Cl}_{2}\right.$ in hexanes $)$ and 12 mg nitrile 28a as yellow solid. The product 44 was re-crystallized in hexanes and afforded colorless lathes and needles crystal: $\mathrm{mp}=85.0-86.4{ }^{\circ} \mathrm{C}$. Oxazole $44:{ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 7.88(\mathrm{dd}, J=8.5,1.4 \mathrm{~Hz}$, $2 \mathrm{H}), 7.37(\mathrm{t}, J=8.1 \mathrm{~Hz}, 2 \mathrm{H}), 7.29(\mathrm{dd}, J=7.1,1.0 \mathrm{~Hz}, 2 \mathrm{H}), 7.28-7.15(\mathrm{~m}, 4 \mathrm{H}), 4.89$ (s(br), 2H), $4.04(\mathrm{~s}, 2 \mathrm{H}), 2.22(\mathrm{~d}(\mathrm{br}), J=19.8 \mathrm{~Hz} 2 \mathrm{H}), 2.18-2.00(\mathrm{~m}, 3 \mathrm{H}), 1.60(\mathrm{dd}, J=$ $7.7,4.5 \mathrm{~Hz}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 13.77,15.79,20.78,34.85,123.15,126.00,126.83$, 126.84, 128.39, 128.45, 128.86, 132.77, 135.67, 136.01, 145.72, 160.45; HRMS m/z 328.1711 (Calc. $328.1702, \mathrm{C}_{23} \mathrm{H}_{21} \mathrm{NO}$ ). The data crystal was cut from a long lathe and had approximate dimension: $0.27 \times 0.17 \times 0.04 \mathrm{~mm}$. The data were collected on a Nonius Kappa CCD diffractimeter using a graphite monochromator with $\mathrm{MoK} \alpha$ radiation ( $\lambda=$
$0.71073 \AA$ ). A total of 400 frames of data were collected using $\omega$-scans with a scan range of $1^{\circ}$ and a counting time of 136 seconds per frame. The data were collected at 153 K using an Oxford Cryostream low temperature device. Crystallographic data for compound 44 has been deposited with the Cambridge Crystallographic Data Center as CCDC 204592 and can be obtained free of charge from CCDC at deposit@ccdc.cam.ac.uk.

### 6.2.31 2,5-Diphenylpyridinium triflate salt 43



To a solution of 2,5-diphenylpyridine $42(8 \mathrm{mg}, 0.04 \mathrm{mmol})$ in $1 \mathrm{~mL} \mathrm{CH} \mathrm{Cl}_{2}$ at $-40^{\circ}$ was added methyl triflate ( $6.5 \mathrm{mg}, 0.05 \mathrm{mmol}$ ) in 1 mL ether solution via cannula. The reaction mixture was stirred at $-20^{\circ} \mathrm{C}$ for 1 h , and then at $0^{\circ} \mathrm{C}$ for 12 h . The triflate salt $43(10 \mathrm{mg}, 75 \%)$, precipitated from ether as light yellow solid, was collected by filtering off solvent. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 4.25(\mathrm{~s}, 3 \mathrm{H}), 7.40-7.85(\mathrm{~m}, 11 \mathrm{H}), 8.51(\mathrm{~d}, \mathrm{~J}=1.8 \mathrm{~Hz}$, 1H), 9.17 (s, 1H).

### 6.2.32 1-Triisopropylsilyl-3-phenyl-3-ene-4-aza-1,6-diyne 53e



Propargylamine ( $200.0 \mathrm{mg}, 3.49 \mathrm{mmol}$ ) was added dropwise to a mixture of 1-phenyl-3-(triisopropylsilyl)-propynone ( $500.0 \mathrm{mg}, 1.75 \mathrm{mmol}$ ) and titanium (IV) ethoxide ( 600.0 $\mathrm{mg}, 2.62 \mathrm{mmol}$ ) in 10 mL dry THF. The resulting yellow mixture was heated to reflux under argon for 1.5 h and then cooled to room temperature. The mixture was poured into water $(30 \mathrm{~mL})$, and the water layer was extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}(2 \times 20 \mathrm{~mL})$. The residue upon drying and concentration of the organic layer was purified by flash chromatography ( $0-2 \%$ ethyl acetate in hexanes) to afford 290.0 mg (52\%) of compound 53e as a yellow oil: $R_{f} 0.50$ (ethyl acetate : hexanes 1:9); ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 1.12-1.21(\mathrm{~m}, 21 \mathrm{H}), 2.28(\mathrm{t}$, $1 \mathrm{H}, J=2.8 \mathrm{~Hz}), 4.58(\mathrm{~d}, 2 \mathrm{H}, J=2.8 \mathrm{~Hz}), 7.36-7.44(\mathrm{~m}, 3 \mathrm{H}), 8.07(\mathrm{dd}, 2 \mathrm{H}, J=8.0,1.6$ $\mathrm{Hz}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 11.09,18.61,45.07,70.87,81.26,97.15,105.18,127.69$, 128.24, 130.92, 136.59, 153.72; HRMS $m / z 324.2153$ (calcd 324.2147, $\mathrm{C}_{21} \mathrm{H}_{29} \mathrm{NSi}$ ).

### 6.2.33 3-Phenyl-prop-2-ynylidene)-prop-2-ynyl-amine 53f



Propargylamine ( $169.0 \mathrm{mg}, 3.08 \mathrm{mmol}$ ) was added dropwise to a mixture of phenyl propynal ( $200.0 \mathrm{mg}, 1.54 \mathrm{mmol}$ ) and titanium (IV) ethoxide $(600.0 \mathrm{mg}, 2.62 \mathrm{mmol})$ in 3 mL dry THF. The resulting yellow mixture was stirred at room temperature for 30 min . The mixture was poured into water ( 30 mL ), and the water layer was extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ $(2 \times 20 \mathrm{~mL})$. The residue upon drying and concentration of the organic layer was purified by re-crystallization (in hexanes) to afford 155.0 mg ( $60 \%$ ) of compound $\mathbf{5 3 f}$ as a white cube crystal (two isomers, $1: 3$ ratio as judged by ${ }^{1} \mathrm{H}$ NMR): $R_{f} 0.40$ (ethyl acetate : hexanes 1:9); ${ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CD}_{2} \mathrm{Cl}_{2}\right) \delta 2.34(\mathrm{t}, J=2.4 \mathrm{~Hz}, 0.3 \mathrm{H}), 2.63(\mathrm{t}, J=2.4 \mathrm{~Hz}, 0.7 \mathrm{H})$, 4.45-4.51 (m, 2H), 7.36-7.44 (m, 3H), 7.52-7.58 (m, 2H), $7.76(\mathrm{t}, J=3.0 \mathrm{~Hz}, 0.3 \mathrm{H}), 8.08$ $(\mathrm{t}, J=1.8 \mathrm{~Hz}, 0.7 \mathrm{H}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(\mathrm{CD}_{2} \mathrm{Cl}_{2}\right) \delta 44.95,48.04,71.22,76.84,78.15,81.21$, $81.50,86.78,92.88,99.53,121.11,121.71,128.86,128.96,130.01,130.49,132.52$, 132.54, 145.67, 146.87; HRMS $m / z 168.0807$ (calcd 168.0807, $\mathrm{C}_{12} \mathrm{H}_{10} \mathrm{~N}$ ).

### 6.2.34 3-Cyclohexa-2,4-dienylmethyl-4-phenylpyridine 59a and 3-Cyclohexa-2,5-dienylmethyl-4-phenylpyridine 59b




To a solution of $\mathbf{5 3 f}(190 \mathrm{mg}, 1.14 \mathrm{mmol})$ in 2 mL THF and 2 mL 1,4-CHD was added $\mathrm{Et}_{3} \mathrm{~N}(580 \mathrm{mg}, 5.69 \mathrm{mmol})$ at room temperature. The reaction mixture was then stirred at $45^{\circ} \mathrm{C}$ for 40 h . The compounds $\mathbf{5 9}$ a,b were purified by flash chromatography ( $0-10 \%$ $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ in hexanes) to afford $47 \mathrm{mg}(27 \%)$ of $\mathbf{5 9} \mathbf{a}, \mathbf{b}$ as a light yellow wax product: $R_{f}$ 0.25 (25\% ethyl acetate in hexanes); ${ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right)(\mathbf{5 9 a} / \mathbf{5 9 b}) \delta 1.95-2.08$ and 2.092.22 (two m, 1H), 2.35-2.40(m, 0.5H), 2.58-2.65 (m, 1H), 3.05-3.18 (m, 0.5H), 3.55-3.68 $(\mathrm{m}, 0.5 \mathrm{H}) 3.80-3.96(\mathrm{~m}, 1 \mathrm{H}), 5.45-5.55(\mathrm{~m}, 2 \mathrm{H}), 5.68-5.5 .78(\mathrm{~m}, 2 \mathrm{H}), 5.88-5.95(\mathrm{~m}$, $0.5 \mathrm{H}), 7.12-7.7 .32(\mathrm{~m}, 5 \mathrm{H}), 7.52-7.60(\mathrm{~m}, 1 \mathrm{H}), 8.42(\mathrm{t}, 1 \mathrm{H}, J=4.8 \mathrm{~Hz}), 8.55($ brs, 1 H$)$ HRMS $m / z 248.1437$ (calcd 248.1439, $\mathrm{C}_{18} \mathrm{H}_{18} \mathrm{~N}$ ).

### 6.2.35 (1-Phenyl-prop-2-ynylidene)-propa-1,2-dienyl-amine 4d



A 1 M solution of TBAF ( 1.46 mmol ) was slowly added to a solution of the compound $\mathbf{5 3 e}(130.0 \mathrm{mg}, 0.40 \mathrm{mmol})$ in dry THF $(3 \mathrm{~mL})$ cooled to $-78^{\circ} \mathrm{C}$. After being stirred at $78^{\circ} \mathrm{C}$ for 5 min , the mixture was poured into ice-water $(10 \mathrm{~mL})$ and extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ $(2 \times 15 \mathrm{~mL})$. The organic layer upon drying, concentration, and removal of trace of solvent under vacuum $(0.05-0.1 \mathrm{mmHg} / 5 \mathrm{~min})$ afforded aza-enyne allene $\mathbf{4 d}$ as a yellow oil. $R_{f} 0.45$ (ethyl acetate : hexanes 1:9); ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 3.82(\mathrm{~s}, 1 \mathrm{H}), 5.36(\mathrm{~d}, 2 \mathrm{H}, J$ $=6.0 \mathrm{~Hz}), 7.38-7.41(\mathrm{~m}, 3 \mathrm{H}), 7.60(\mathrm{t}, 1 \mathrm{H}, J=6.0 \mathrm{~Hz}), 8.07(\mathrm{dd}, 2 \mathrm{H}, J=7.6,2.0 \mathrm{~Hz}),{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 68.17,81.06,89.72,91.43,110.11,127.99,128.54,131.12,136.72$, 147.10, 213.72. CIMS $m / z 168\left(\mathrm{MH}^{+}\right)$.
6.2.36 5-Cyclohexa-2,4-dienylmethyl-2-phenylpyridine 56a, 5-cyclohexa-2,5-dienylmythyl-2-phenylpyridine 56b, and 6-phenyl-3-picoline 55




Compound $\mathbf{4 d}$ ( 0.40 mmol ) prepared as above was placed in a round bottle flask that was then cooled to $0{ }^{\circ} \mathrm{C}$. To this was added freshly distilled 1,4 -chd $(2 \mathrm{~mL})$. The reaction mixture was then kept at $4{ }^{\circ} \mathrm{C}$ for 8 h . The residue upon concentration was purified by flash chromatography ( $0-2 \%$ ethyl acetate in hexanes) to afford $16.5 \mathrm{mg}(17 \%)$ of $1: 1$ ratio mixture of $\mathbf{5 6 a}$ and $\mathbf{5 6 b}$ as a yellow oil and $13.0 \mathrm{mg}(20 \%)$ of compound $\mathbf{5 5}$ as a light yellow solid: $\left.R_{f} \mathbf{( 5 6 a / 5 6 b}\right) 0.40$ (ethyl acetate : hexanes 1:9); ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right)$ (56a/56b) $\delta 1.98-2.08$ and 2.20-2.31 (two m, 1H), 2.50-2.60 (m, 0.5H), 2.60-2.71 (m, $1 \mathrm{H}), 2.72(\mathrm{~d}, 2 \mathrm{H}, J=6.9 \mathrm{~Hz}), 2.97-3.10(\mathrm{~m}, 0.5 \mathrm{H}), 5.58-5.78(\mathrm{~m}, 3 \mathrm{H}), 5.88-5.94(\mathrm{~m}, 1 \mathrm{H})$, 7.37-7.48 (m, 3H), 7.52-7.58(m, 1H), $7.63(\mathrm{dd}, 1 \mathrm{H}, J=8.4,1.8 \mathrm{~Hz}), 7.96(\mathrm{~d}, 2 \mathrm{H}, J=7.2$ Hz ), 8.49 (brs, 1H); CI-MS $m / z(\mathbf{5 6 a} / \mathbf{5 6 b}) 248 ; R_{f} \mathbf{( 5 5 )} 0.36$ (ethyl acetate : hexanes, 1:9); $\mathrm{mp}(\mathbf{5 5}) 52-54{ }^{\circ} \mathrm{C}$, Lit: ${ }^{1} \mathrm{mp} 56{ }^{\circ} \mathrm{C}$; ${ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right)(55) \delta 2.35(\mathrm{~s}, 3 \mathrm{H}), 7.35-7.40(\mathrm{~m}$, $1 \mathrm{H}), 7.41-7.47(\mathrm{~m}, 2 \mathrm{H}), 7.54(\mathrm{dd}, 1 \mathrm{H}, J=7.6,2.0 \mathrm{~Hz}), 7.60(\mathrm{~d}, 1 \mathrm{H}, J=7.6 \mathrm{~Hz}), 7.93(\mathrm{dd}$, $2 \mathrm{H}, J=8.4,1.6 \mathrm{~Hz}), 8.50(\mathrm{brs}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right)(6 \mathrm{~b}) \delta 17.67,120.09,126.68$, $128.58,128.68,131.61,137.34,139.36,150.02,154.80 ;$ CI-MS m/z (55) 170.

### 6.2.37 5-Cyclohexylmethyl-2-phenylpyridine 57



A 5 mL empty round bottle flask was charged with $10 \%$ palladium on activated carbon $(10.0 \mathrm{mg}, 0.0063 \mathrm{mmol})$; the flask was purged and then filled with hydrogen gas (this process was repeated three times). To this flask was added $\mathbf{5 6 a} / \mathbf{5 6 b}(15.0 \mathrm{mg}, 0.063$ mmol ) in degassed ethyl acetate ( 1 mL ). The reaction mixture was stirred under an atmosphere of $\mathrm{H}_{2}$ at room temperature for 2 h . The reaction mixture was filtered, concentrated, and purified by flash chromatography ( $0.5 \%$ ethyl acetate in hexanes) to afford $13.0 \mathrm{mg}(87 \%)$ of compound 57 as a white solid: $\mathrm{mp} 60-62{ }^{\circ} \mathrm{C} ; R_{f} 0.47$ (ethyl acetate : hexanes, 1:9); ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 0.90-1.02(\mathrm{~m}, 2 \mathrm{H}), 1.14-1.1 .24(\mathrm{~m}, 4 \mathrm{H}), 1.48-$ $1.58(\mathrm{~m}, 1 \mathrm{H}), 1.60-1.74(\mathrm{~m}, 4 \mathrm{H}), 2.50(\mathrm{~d}, 2 \mathrm{H}, J=7.2 \mathrm{~Hz}), 7.37(\mathrm{t}, 1 \mathrm{H}, J=7.2 \mathrm{~Hz}), 7.40-$ $7.48(\mathrm{~m}, 2 \mathrm{H}), 7.51(\mathrm{dd}, 1 \mathrm{H}, J=8.0,2.0 \mathrm{~Hz}), 7.63(\mathrm{~d}, 1 \mathrm{H}, J=8.0 \mathrm{~Hz}), 7.96(\mathrm{dd}, 2 \mathrm{H}, J=$ $8.8,1.6 \mathrm{~Hz}), 8.46(\mathrm{~d}, 1 \mathrm{H}, J=2.0 \mathrm{~Hz}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 26.19,26.42,32.96,39.53$, 40.69, 119.98, 126.68, 128.62, 128.69, 134.99, 137.42, 139.29, 150.23, 154.89; HRMS $m / z 252.1747$ (calcd 252.1752, $\mathrm{C}_{18} \mathrm{H}_{21} \mathrm{~N}$ ).

### 6.2.38 5-Methoxymethyl-2-phenylpyridine 58



Compound $\mathbf{4 d}(0.17 \mathrm{mmol})$ prepared as above was placed in a round bottle flask that was then cooled to $0{ }^{\circ} \mathrm{C}$. To this was added freshly distilled methanol $(2 \mathrm{~mL})$. The reaction mixture was then kept at $0^{\circ} \mathrm{C}$ for 14 h . The residue upon concentration was purified by flash chromatography ( $0-10 \%$ ethyl acetate in hexanes) to afford $6.7 \mathrm{mg}(20 \%)$ of compound 58 as a yellow oil: $R_{f} 0.20$ (ethyl acetate : hexanes, 1:9); ${ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta$ 3.41 (s, 3H), 4.50 (s, 2H), 7.38-7.49 (m, 3H), 7.71-7.74 (m, 2H), 7.94-7.99 (m, 2H), 8.62 (brs, 1 H$) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 58.32,71.97,120.32,126.91,128.75,128.98,131.94$, 136.41, 139.09, 149.05, 156.97; HRMS $m / z 200.1074$ (calcd 200.1075, $\mathrm{C}_{13} \mathrm{H}_{13} \mathrm{NO}$ ).

### 6.2.39 6-phenylpyridine-3-carbaldehyde 54



Compound $4 \mathbf{d}$ ( 0.31 mmol ) prepared above was dissolved in 1 mL of $\mathrm{CDCl}_{3}$. The solution was stored at $10{ }^{\circ} \mathrm{C}$ in a fridge for 12 h . The residue upon concentration was purified by flash chromatography ( $10 \%$ ethyl acetate in hexanes) to afford $1.0 \mathrm{mg}(1.8 \%)$ of compound 54 as a yellow wax product: ${ }^{2} R_{f} 0.12$ (ethyl acetate : hexanes, 1:9); ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.45-7.52(\mathrm{~m}, 3 \mathrm{H}), 7.90(\mathrm{~d}, 1 \mathrm{H}, J=8.4 \mathrm{~Hz}), 8.03-8,08(\mathrm{~m}, 2 \mathrm{H}), 8.22(\mathrm{dd}$, $1 \mathrm{H}, J=8.4,2.0 \mathrm{~Hz}$ ), $9.11(\mathrm{dd}, 1 \mathrm{H}, J=2.0,0.4 \mathrm{~Hz}), 10.13(\mathrm{~s}, 1 \mathrm{H}) ;$ HRMS $m / z 184.0765$ (calcd 184.0762, $\mathrm{C}_{12} \mathrm{H}_{9} \mathrm{NO}$ ).

### 6.3 SPECTROSCOPIC (UV-VIS) KINETICS (40A-C)

To a solution of aza-enediyne 26a-c ( 0.0052 mmol ) in dry THF ( 2 mL ) cooled to $-78{ }^{\circ} \mathrm{C}$ was added slowly a 1 M solution of tetrabutylammonium floride in dry THF (0.0059 mmol ). The reaction mixture was allowed to stir at $-78^{\circ} \mathrm{C}$ for $5-10 \mathrm{~min}$, and the mixture was poured into ice-water $(10 \mathrm{~mL})$ and extracted by $\mathrm{CH}_{2} \mathrm{Cl}_{2}(2 \times 15 \mathrm{~mL})$. The dry organic solution was chromatographed through a small plug of silica gel to remove tetrabutylammonium species before evaporation. The residue was stored at $-80^{\circ} \mathrm{C}$ and quickly redissolved in $\mathrm{CHCl}_{3}$ or other solvents (THF, $i \mathrm{PrOH}$, chlorobenzene, hexanes, $\mathrm{CH}_{3} \mathrm{CN}$ ) and the resulting solution transferred to a thermostated cuvette at $20,25,30,37$, or $45{ }^{\circ} \mathrm{C}$. The conversion of aza-enediyne 40a-c to (Z)-nitrile 28a-c was followed spectrophotometrically by periodically acquiring UV absorbance spectra (250-500 nm) over a period of at least 2 half-lives. The first-order rate constants were obtained by fitting plots of the normalized absorbance at $390 \mathrm{~nm}(\mathbf{4 0 a})$ or $415 \mathrm{~nm}(\mathbf{4 0 b} / \mathbf{4 0 c})$ versus time to an exponential decay.

### 6.4 SPECTROSCOPIC ( ${ }^{\mathbf{1}} \mathrm{H}$ NMR AND ${ }^{13} \mathrm{C}$ NMR) KINETICS (26D)

A NMR tube containing the degassed (freeze-pump-thaw) solution of aza-enediyne 26d ( 120 mM ) and 2,5-dimethylfuran (1.5 equiv, internal standard) in $d_{6}$-benzene was sealed under flame. ${ }^{1} \mathrm{H}$ NMR of the sample was taken as initial spectrum (reaction time equals zero second). After ${ }^{1} \mathrm{H}$ NMR, the tube was kept at $70^{\circ} \mathrm{C}$. The conversion of aza-enediyne to enediyne and nitrile was followed spectroscopically by taking ${ }^{1} \mathrm{H}$ NMR spectra 15 times over a period of time. Intervals varied from 35 minutes to 5 hours. Every time, the ${ }^{1} \mathrm{H}$ NMR tube was taken out of a $70{ }^{\circ} \mathrm{C}$ oil bath, and immediately inserted into a baker containing ice-water to stop the reaction. Reaction time was measured as the time during which the tube maintained at $70^{\circ} \mathrm{C}$. Aza-enediyne $\mathbf{2 6 d}$ has a doublet resonance at 7.45 ppm which is well resolved from other peaks. The disappearance of the aza-enediyne 26d was monitored as the disappearance of the resonance for $\mathbf{2 6 d}$ at 7.45 ppm . The integrals of the resonance for $\mathbf{2 6 d}$ at 7.45 ppm were normalized against the resonance for the internal standard 2,5-dimethylfuran at 5.76 ppm . The relative concentrations of azaenediyne 26d were calculated by normalizing the integrals of the resonance for 26d at 7.45 ppm obtained at different intervals against the integral of the resonance for $\mathbf{2 6 d}$ at 7.45 ppm obtained at reaction time equals zero second. The kinetic data was obtained by fitting plots of the relative concentrations of aza-enediyne $\mathbf{2 6 d}$ versus time. TableCurve $2 D^{\circledR}$ from SYSTAT Software Inc. was used to analyze the kinetic data. Kinetic data fits independent first-order and second-order kinetics with an equation: $\frac{d[A]}{d t}=-k_{1}[A]-k_{2}[A]^{2},[\mathrm{~A}]$ : concentration of aza-enediyne 26d; $\mathrm{k}_{1}$ : first-order constant; $\mathrm{k}_{2}$ : second-order constant.

The conversion of aza-enediyne $\left[4-{ }^{13} \mathrm{C}\right]-26 d$ to enediyne $\left[1,4-{ }^{13} \mathrm{C}_{2}\right]-\mathbf{6 0 d}$ and nitrile $\left[5-{ }^{13} \mathrm{C}\right]-28 d$ can be followed by ${ }^{13} \mathrm{C}$ NMR by a procedure analogous to that described for ${ }^{1} \mathrm{H}$ NMR kinetics. The resonance for $\left[4-{ }^{13} \mathrm{C}\right]-26 d$ at $159 \mathrm{ppm}\left({ }^{13} \mathrm{C}\right)$
diminished over time and was replaced by the resonance for $\left[1,4-{ }^{13} \mathrm{C}_{2}\right]-60 \mathrm{~d}$ at 97 ppm $\left({ }^{13} \mathrm{C}\right)$ and the resonance for $\left[5-{ }^{13} \mathrm{C}\right]-\mathbf{2 8 d}$ at $100 \mathrm{ppm}\left({ }^{13} \mathrm{C}\right)$. The integrals of the resonance for [4- $\left.{ }^{13} \mathrm{C}\right]-26 d$ at 159 ppm were normalized against the resonance for solvent (benzene, THF or DMSO). The relative concentrations of aza-enediyne [ $\left.4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ were calculated by normalizing the integrals of the resonance for $\left[4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ at 159 ppm obtained at different intervals against the integral of the resonance for $\left[4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ at 159 ppm obtained at reaction time equals zero second. The kinetic data was obtained by fitting plots of the relative concentration versus time. TableCurve $2 \mathrm{D}^{\circledR}$ from SYSTAT Software Inc. was used to analyze the kinetic data. Kinetic data fits independent first-order and second-order kinetics with an equation: $\frac{d[A]}{d t}=-k_{1}[A]-k_{2}[A]^{2}$, [A]: concentration of aza-enediyne 26d; $\mathrm{k}_{1}$ : first-order constant; $\mathrm{k}_{2}$ : second-order constant.

## REFERENCE

1. Kagabu, S.; Ando, C.; Ando, J.; J. Chem. Soc. Perkin Trans 1, 1994, 6, 739.
2. Tanaka, A.; Terasawa, T.; Hagihara, H.; Sakuma, Y.; Ishibe, N.; Sawada, M.; Takasugi, H.; Tanaka, H. J. Med. Chem. 1998, 41, 2390.
3. Alonso, D. A.; Najera, C.; Pacheco, C. J. Org. Chem. 2004, 65, 1615.
4. Carpino, L. A. J. Am. Chem. Soc. 1960, 82, 3133-3135.
5. Krause, J.G. Synthesis, 1972, 3, 140.
6. Caution: O-Mesitylenesulfonylhydroxyamine is potentially explosive: Ning, R. Y. Chem. Engineer News 1973, 51, 36.
7. Shimizu, T.; Miyasaka, D.; Kamigata, N. Organic Lett. 2000, 2, 1923.
8. Refat, H.M.; Waggenspack, J.; Dutt, M.; Zhang, H.; Fadda, A.A.; Biehl, E. J. Org. Chem. 1995, 60, 1985

## Appendices

## APPENDIX A ENERGIES CALCULATED AT THE (U)B3LYP/6-31* LEVEL OF THEORY

Table 1 Energies calculated on Myers-Saito and Schmittel cyclizations of $\mathbf{4 a}$ at the (U)B3LYP/6-31* level

|  | Electronic energy ${ }^{\text {a }}$ | ZPE ${ }^{\text {b }}$ | Relative energy ${ }^{\text {c }}$ |
| :---: | :---: | :---: | :---: |
| (Z)-Enyne allene (S-cis) 4a | -270.20169 | 62.2 | 4.0 |
| (Z)-Enyne allene (S-trans) 4a | -270.20832 | 62.3 | 0 |
| (E)-Enyne allene (S-cis) 4a | -270.20436 | 62.1 | -2.2 |
| (E)-Enyne allene (S-trans) 4a | -270.20816 | 62.1 | -0.1 |
| $\alpha, 3$-Didehydrotoluene triplet diradical 5a | -270.22722 | 64.0 | -10.2 |
| $\alpha, 3$-Didehydrotoluene singlet diradical 5a | -270.22906 | 63.7 | -11.6 |
| Zwitterion ${ }^{\text {d }}{ }^{ \pm}$ | -270.17919 | 65.0 | 20.9 |
| Cyclic allene 24a | -270.21139 | 64.4 | 0.1 |
| $\begin{aligned} & \hline \text { Myers-Saito transition } \text { state }^{\mathrm{e}} \\ & \text { TS1a } \end{aligned}$ | -270.16603 | 62.1 | 26.3 |
| Schmittel triplet diradical 52a, anti-addition | -270.19203 | 62.6 | 10.5 |
| Schmittel singlet diradical 52a, anti-addition | -270.18754 | 62.4 | 13.1 |
| Schmittel transition state $^{\mathrm{e}}$ TS2a, anti-addition | -270.1521095 | 61.5 | 34.5 |

${ }^{\text {a }}$ Absolute energies in Hartrees; ${ }^{b} \mathrm{ZPE}$ in $\mathrm{kcal} / \mathrm{mol}$; ${ }^{\mathrm{c}}$ Energies relative to ( $Z$ )-Enyne allene (S-trans) $\mathbf{4 a}$ in $\mathrm{kcal} / \mathrm{mol}$; ${ }^{\mathrm{d}} 1$ imaginary frequency at the $\mathrm{HF} / 6-31 \mathrm{G}^{*}$ level; ${ }^{\mathrm{e}} 1$ imaginary frequency at the B3LYP/6-31G* level.

Table 2 Energies calculated on aza-Myers-Saito and aza-Schmittel cyclizations of $\mathbf{4 b}$ at the (U)B3LYP/6-31G* level

|  | Electronic energy ${ }^{\text {a }}$ | ZPE ${ }^{\text {b }}$ | Relative energy ${ }^{\text {c }}$ |
| :---: | :---: | :---: | :---: |
| (Z)-Aza-enyne allene (S-cis) 4b | -286.23233 | 54.7 | 3.5 |
| (Z)-Aza-enyne allene (S-trans) 4b | -286.23795 | 54.8 | 0 |
| (E)-Aza-enyne allene (S-cis) 4b | -286.23454 | 54.7 | -2.0 |
| (E)-Aza-enyne allene (S-trans) 4b | -286.23766 | 54.5 | -0.1 |
| $\alpha, 5$-Didehydro-3-picoline triplet diradical 5b | -286.26314 | 56.6 | -14.0 |
| a,5-Didehydro-3-picoline singlet diradical 5b | -286.26503 | 56.4 | -15.4 |
| Aza-zwitterion ${ }^{\text {d }} \mathbf{5 b}^{ \pm}$ | -286.21523 | 57.0 | 16.5 |
| Aza-cyclic allene 24b | -286.24552 | 56.9 | -2.6 |
| (Z)-Aza-skipped enediyne 53b | -286.21899 | 54.9 | 12.0 |
| ( $E$ )-Aza-skipped enediyne 53b | -286.22186 | 54.7 | 10.1 |
| Aza-Myers transition state ${ }^{\text {d }}$ TS1b | -286.20081 | 54.6 | 23.2 |
| Aza-Schmittel singlet diradical 52b, syn-addition | -286.22656 | 54.8 | 7.2 |
| Aza-Schmittel triplet diradical 52b, syn-addition | -286.23075 | 55.0 | 4.8 |
| Aza-Schmittel transition state ${ }^{\text {d }}$ TS2b, syn-addition, | -286.18804 | 53.8 | 30.4 |
| Aza-Schmittel singlet diradical 52b, anti-addition | -286.22630 | 54.8 | 7.3 |
| Aza-Schmittel triplet diradical 52b, anti-addition | -286.23001 | 54.9 | 5.2 |
| Aza-Schmittel transition state ${ }^{\text {d }}$ TS2b, anti-addition | -286.19269 | 54.1 | 27.7 |

${ }^{\text {a }}$ Absolute energies in Hartrees; ${ }^{b} \mathrm{ZPE}$ in $\mathrm{kcal} / \mathrm{mol}$; ${ }^{\text {c }}$ Energies relative to ( $Z$ )-Aza-enyne allene (S-trans) 4b in $\mathrm{kcal} / \mathrm{mol} ;{ }^{\mathrm{d}} 1$ imaginary frequency at the B3LYP/6-31G* level.

## APPENDIX B SPECTROSCOPIC KINETICS FOR AZA-ENEDIYNE 26D



Figure 1 Kinetic plot of the transformation of aza-enediyne $\mathbf{2 6 d}$ to enediyne $\mathbf{6 0 d}$ and the conversion of $\mathbf{2 6 d}$ to nitrile $\mathbf{6 1}$ followed by ${ }^{1} \mathrm{H}$ NMR in $d_{6}$-benzene. The data fit independent first-order and second-order kinetics simultaneously by using TableCurve $2 D^{\circledR}$ from SYSTAT Software Inc. The x-axis represents reaction time in seconds; the yaxis represents the concentrations of $\mathbf{2 6 d}$ in $\mathrm{mol} / \mathrm{L}$ calculated from the ratio of the integals of resonance at 7.45 ppm from $\mathbf{2 6 d}$ to the one at 5.76 ppm from internal standard 2,5-dimethyl furan (see section 6.4 for details). The first-order rate constant $\mathrm{k}_{1}=4.15 \mathrm{x}$ $10^{-5} \mathrm{~s}^{-1}$, the second-order rate constant $\mathrm{k}_{2}=4.29 \times 10^{-4} \mathrm{~s}^{-1} \mathrm{~mol}^{-1} \mathrm{~L}$.


Figure 2 Kinetic plot of the transformation of aza-enediyne [ $\left.4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ to enediyne [1,4$\left.{ }^{13} \mathrm{C}_{2}\right]$-60d and the conversion of $\left[4-{ }^{13} \mathrm{C}\right]-26 d$ to nitrile 61 followed by ${ }^{13} \mathrm{H}$ NMR in $d_{6^{-}}$ benzene. The data fit independent first-order and second-order kinetics simultaneously by using TableCurve $2 D^{\circledR}$ from SYSTAT Software Inc. The $x$-axis represents reaction time in seconds; the y-axis represents the concentrations of $\left[4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ in $\mathrm{mol} / \mathrm{L}$ calculated from the ratio of the integals of resonance at 159 ppm from $\left[4-{ }^{13} \mathrm{C}\right]-26 d$ to the one from $d_{6}$-benzene (see section 6.4 for details). The first-order rate constant $\mathrm{k}_{1}=4.71 \times 10^{-5} \mathrm{~s}^{-1}$, the second-order rate constant $\mathrm{k}_{2}=3.83 \times 10^{-4} \mathrm{~s}^{-1} \mathrm{~mol}^{-1} \mathrm{~L}$.


Figure 3 Kinetic plot of the transformation of aza-enediyne [ $\left.4-{ }^{13} \mathrm{C}\right]$-26d to enediyne [1,4$\left.{ }^{13} \mathrm{C}_{2}\right]-60 \mathrm{~d}$ and the conversion of $\left[4-{ }^{13} \mathrm{C}\right]-26 d$ to nitrile 61 followed by ${ }^{13} \mathrm{H}$ NMR in $d_{8^{-}}$ THF. The data fit independent first-order and second-order kinetics simultaneously by using TableCurve $2 D^{\circledR}$ from SYSTAT Software Inc. The x -axis represents reaction time in seconds; the $y$-axis represents the concentrations of $\left[4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ in $\mathrm{mol} / \mathrm{L}$ calculated from the ratio of the integals of resonance at 159 ppm from $\left[4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ to the one from $d_{8}$-THF (see section 6.4 for details). The first-order rate constant $\mathrm{k}_{1}=1.79 \times 10^{-5} \mathrm{~s}^{-1}$, the second-order rate constant $\mathrm{k}_{2}=2.04 \times 10^{-4} \mathrm{~s}^{-1} \mathrm{~mol}^{-1} \mathrm{~L}$.


Figure 4 Kinetic plot of the transformation of aza-enediyne [ $\left.4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ to enediyne [1,4$\left.{ }^{13} \mathrm{C}_{2}\right]-60 \mathrm{~d}$ and the conversion of $\left[4-{ }^{13} \mathrm{C}\right]-26 \mathrm{~d}$ to nitrile 61 followed by ${ }^{13} \mathrm{H}$ NMR in $d_{6^{-}}$ DMSO. The data fit independent first-order and second-order kinetics simultaneously by using TableCurve $2 \mathrm{D}^{\circledR}$ from SYSTAT Software Inc. The x -axis represents reaction time in seconds; the $y$-axis represents the concentrations of $\left[4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ in $\mathrm{mol} / \mathrm{L}$ calculated from the ratio of the integals of resonance at 159 ppm from $\left[4-{ }^{13} \mathrm{C}\right]-\mathbf{2 6 d}$ to the one from $d_{6}$-DMSO (see section 6.4 for details). The first-order rate constant $\mathrm{k}_{1}=7.18 \times 10^{-6} \mathrm{~s}^{-1}$, the second-order rate constant $\mathrm{k}_{2}=1.56 \times 10^{-3} \mathrm{~s}^{-1} \mathrm{~mol}^{-1} \mathrm{~L}$.

## APPENDIX C SELECTED ${ }^{\mathbf{1}} \mathbf{H}$ NMRS FOR COMPOUNDS PREPARED IN EXPERIMENTAL SECTION

Compound 32a


Compound 33a


Compound 34a


## Compound 26a




Compound 28a


Compound 28a



Compound 28a after 4.5 h irradiation at 371 nm



Compound 33b



Compound 33b


Compound 34b


Compound 26b


Compound 28b


Compound 39


Compound 32c


Compound 33c



## Compound 34c



Compound 26c


## Compound 28c



Compound 43


Compound 38d


## Compound 26d



Compound (E)-60d


## Compound 60d



Compound (E)-61


Compound 28d


Compound 60e



1:1




Compound 26g


Compound 62


Compound 44


## Compound 53e



Compound 4d


Compound 4d (decoupled ${ }^{1} \mathrm{H}$ NMR)


## Compound 4d ( ${ }^{13} \mathrm{C}$ NMR)



Compound 55


Compound 56a,b


Compound 57


Compound 58


Compound 54


Compound 53f


Compound 59a,b


## Bibliography

Ahlert, J.; Shepard, E.; Lomovskaya, N.; Zazopoulos, E.; Staffa, A.; Bachmann, B. O.; Huang, K.; Fonstein, L.; Czisny, A.; Whitwan, R. E.; Farnet, C. M.; Thorson, J. S. Science 2002, 297, 1173.

Alonso, D. A.; Najera, C.; Pacheco, C. J. Org. Chem. 2004, 65, 1615.
Ando, T.; Ishii, M.; Kajiura, T.; Kameyama, T.; Miwa, K.; Sugiura, Y. Tetrahedron Lett. 1998, 39, 6495.

Basak, A.; Bdour, H. M.; Shain, J. C.; Mandal, S.; Rudra, K. R., Nag, S. Bioorg. Med. Chem. Lett. 2000, 10, 1321.

Basak, A.; Shain, J. C. Tetrahedron Lett. 1998, 30, 3029.
Basak, A.; Shain, J. C.; Khamrai, U. K.; Rudra, K. R. J. Chem. Soc., Perkin Trans. I 2000, 1955.

Bates, R. B.; Taylor, S. R. J. Org. Chem. 1993, 58, 4469.
Becke, A. D. J. Chem. Phys. 1993, 98, 5648.
Birney, D. M. J. Am. Chem. Soc. 2000, 122, 10917.
Blackwell, J. M.; Figueroa, J. S.; Stephens, F, H.; Cummins, C. C. Organometallics 2003, 22, 3351.

Brukner, I. Curr. Opin. Oncol. Endocr. Met. Invest. Drugs 2000, 2, 344 and references therein.

Carpino, L. A. J. Am. Chem. Soc. 1960, 82, 3133.
Carter, P. A.; Magnus, P. J. Am. Chem. Soc. 1988, 110, 6921.
Castarlenas, R.; Esteruelas, M. A.; Onate, E. Organometallics 2005, 24, 4343.
Chaloner, P. A. J. Chem. Soc. Perkin Trans. 2 1980, 1028.
Christner, D. F.; Frank, B. L.; Kozarich, J. W.; Stubbe, J.; Golik, J.; Doyle, T. W.; Rosenberg, I. E.; Kristnan, B. J. Am. Chem. Soc. 1992, 114, 8763.

Cosgrove, J. P.; Dedon, P. C. in Small molecule DNA and RNA binders 2003, vol 2, pp 609-642.

Cramer, C. J. J. Am. Chem. Soc. 1998, 120, 6261.
Cramer, C. J.; Kormos, B. L.; Seierstad, M.; Sherer, E. C.; Winget, P. Org. Lett. 2001, 3, 1881.

Cramer, C. J.; Squires, R. R. Org. Lett. 1999, 1, 215.
Cremeens, M. E.; Hughes, T. S.; Carpenter, B. K. J. Am. Chem. Soc. 2005, 127, 6652.
David, W. M. Ph. D Dissertation 2000, The University of Texas at Austin.
David, W. M.; Kerwin, S. M. J. Am. Chem. Soc. 1997, 119, 1464.
David, W. M.; Kumar, D.; Kerwin, S. M. Bioorg. Med. Chem. Lett. 2000, 10, 2509.
De Voss, J. J.; Townsend, C. A.; Ding, W. D.; Morton, G. O.; Ellestad, G. A.; Zein, N.; Tabor, A. B.; Schreiber, S. L. J. Am. Chem. Soc. 1990, 112, 9669.

De Voss, J. J.; Townsend, C. A.; Ding, W. D.; Morton, G. O.; Ellestad, G. A.; Zein, N.; Tabor, A. B.; Schreiber, S. L. J. Am. Chem. Soc. 1990, 112, 9669.

Debbert, S. L.; Cremer, D. J. Int. J. Mass Spectrom. 2000, 201, 1.
Dedon, P. C. Biol. Ther. Leukemia 2001, 2, 2.
Dedon, P. C.; Goldberg, I. H. Chem. Res. Toxicol. 1992, 5, 311.
Dedon, P. C.; Goldberg, I. H. J. Biol. Chem. 1990, 265, 14713.
Engels, B.; Hanrath, M. J. Am. Chem. Soc. 1998, 120, 6356.
Engels, B.; Lennartz, C.; Hanrath, M.; Schmittel, M.; Strittmatter, M. Angew. Chem. Int. Ed. 1998, 37, 1960.

Feng, L.; Kerwin, S. M. Tetrahedron Lett. 2003, 44, 3463.
Feng, L.; Kumar, D.; Birney, D. M.; Kerwin, S. M. Org. Lett. 2004, 6, 2059.
Feng, L.; Kumar, D.; Kerwin, S. M. J. Org. Chem. 2003, 68, 2234.
Feng, L.; Zhang, A.; Kerwin, S. M. submitted.
Feustel, M.; Himbert, G. Tetrahedron Lett. 1983, 24, 2165.

Foland, L. D.; Karlsson, J. O.; Perri, S. T.; Schwade, R.; Xu, S. L.; Patil, S.; Moore, H. W. J. Am. Chem. Soc. 1989, 111, 975.

Frey, L. F.; Tillyer, R. D.; Ouellet, S. G.; Reamer, R. A.; Grabowski, E. J. J.; Reider, P. J. J. Am. Chem. Soc. 2000, 122, 1215.

Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Zakrzewski, V. G.; Montgomery, Jr., J. A.; Stratmann, R. E.; Burant, J. C.; Dapprich, S.; Millam, J. M.; Daniels, A. D.; Kudin, K. N.; Strain, M. C.; Farkas, O.; Tomasi, J.; Barone, V.; Cossi, M.; Cammi, R.; Mennucci, B.; Pomelli, C.; Adamo, C.; Clifford, S.; Ochterski, J.; Petersson, G. A.; Ayala, P. Y.; Cui, Q.; Morokuma, K.; Rega, N.; Salvador, P.; Dannenberg, J. J.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Cioslowski, J.; Ortiz, J. V.; Baboul, A. G.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Gomperts, R.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Challacombe, M.; Gill, P. M. W.; Johnson, B.; Chen, W.; Wong, M. W.; Andres, J. L.; Gonzalez, C.; Head-Gordon, M.; Replogle, E. S.; Pople, J. A. Gaussian, Inc., Pittsburgh PA, 2002.

Frutos, O. D.; Echavarren, A. M. Tetrahedron Lett. 1997, 38, 7941.
Galm, U.; Hager, M. H.; Van Lanen, S. G.; Ju, J.; Throson, J. S.; Shen, B. Chem. Rev. 2005, 105, 739.

Garcia, J. G.; Ramos, B.; Pratt, L.; Rodriguez, A. Tetrahedron Lett. 1995, 36, 7391.
Giles, F. J.; Estey, E.; O’Brien, S. Cancer 2003, 98, 2095.
Hagopian, R. A.; Therien, M. J.; Murdoch, J. R. J. Am. Chem. Soc. 1984, 106, 5753.
Halter, R. J.; Fimmen, R. L.; McMahon, R. J.; Peebles, S. A.; Kuczkowski, R. L.; Stanton, J. F. J. Am. Chem. Soc. 2001, 123, 12353.

Hayashi, M.; Saigo, K. Tetrahedron Lett. 1997, 38, 6241.
Hayashi, S.-I.; Nair, M.; Houser, D. J.; Shechter, H. Tetrahedron Lett. 1979, 20, 2961.
Herges, R. Angew. Chem. Int. Ed. Engl. 1994, 33, 255.
Hirama, M.; Akiyama, K.; Tanaka, T.; Noda, T.; Iida, K.; Sato, I.; Hanaishi, R.; FukudaIshisaka, S.; Ishiguro, M.; Otani, T.; Leet, J. E. J. Am. Chem. Soc. 2000, 122, 720.

Hoffer, J.; Schottelius, M. J.; Feichhtinger, D.; Chen, P. J. Am. Chem. Soc. 1998, 120, 376.

Hughes, T. S.; Carpenter, B. K. J Chem. Soc., Perkin Trans. 2 1999, 2291.

Jones, G. B.; Fouad, F. S. Curr. Pharm. Des. 2002, 8, 2415.
Jones, G. B.; Plourde, G. W. Org. Lett. 2000, 2, 1757.
Jones, G. B.; Warner, P. M. J. Am. Chem. Soc. 2001, 123, 2134.
Jones, R. R.; Bergman, R. G. J. Am. Chem. Soc. 1972, 94, 660.
Kagabu, S.; Ando, C.; Ando, J.; J. Chem. Soc. Perkin Trans 1, 1994, 6, 739.
Karlsson, J. O.; Nguyen, N. V.; Foland, L. D.; Moore, H. W. J. Am. Chem. Soc. 1985, 107, 3392.

Khasanova, T.; Sheridan, R. S. J. Am. Chem. Soc. 1998, 120, 233.
Kim, C.; Russell, K. C. Tetrahedron Lett. 1999, 40, 3835.
Kimball, D. B.; Herges, R.; Haley, M. M. J. Am. Chem. Soc. 2002, 124, 1572.
Koide, Y.; Ishii, F.; Hasuda, K.; Koyama, Y.; Edo, K.; Katamine, S.; Kitame, F.; Ishida, N. J. Antibiot. 1980, 33, 342.

Konishi, M.; Ohkuma, H.; Matsumoto, K.; Tsuno, T.; Kamei, H.; Miyaki, T.; Oki, T.; Kawaguchi, H.; Van Duyne, G. D.; Clardy, J. J. Antibiot. 1989, 42, 1449.

Konishi, M.; Ohkuma, H.; Saitoh, K.; Kawaguchi, H.; Golik, J.; Dubay, G.; Groenewold, G.; Krishnan, B.; Doyle, T. W. J. Antibiot. 1985, 38, 1605.

Kraka, E.; Cramer, D. J. Am. Chem. Soc. 2000, 122, 8245.
Kraka, E.; Cramer, D. J. Comput. Chem. 2001, 22, 216.
Krause, J. G. Synthesis, 1972, 3, 140.
Krause, N.; Hashmi, A. S. K. Modern Allene Chemistry, chapter one, 2004, Wiley.
Krivdin, L. B.; Della, E. W. Progress in NMR Spectroscopy 1991, 23, 301.
Lambert, J.; Klessinger, M. Magn. Reson. Chem. 1987, 25, 456.
Lee, M. D.; Dunne, T. S.; Siegel, M. M.; Chang, C. C.; Morton, G. O.; Borders, D. B. J. Am. Chem. Soc. 1987, 109, 3464.

Leet, J. E.; Schroeder, D. R.; Hofstead, S. J.; Golik, J.; Colson, K. L.; Huang, S.; Klohr, S. E.; Doyle, T. W.; Matson, J. A. J. Am. Chem. Soc. 1992, 114, 7946.

Li, H.; Peterson, J. L.; Wang, K. K. J. Org. Chem. 2003, 68, 5512.

Li, H.; Yang, H.; Peterson, J. L.; Wang, K. K. J. Org. Chem. 2004, 69, 4500.
Liu W.; Christenson, S. D.; Standage, S.; Shen, B. Science, 2002, 297, 1170.
Logan, C. F.; Chen, P. J. Am. Chem. Soc. 1996, 118, 4896.
Lopez-Larraza, D. M.; Moore, K.; Dedon, P. C. Chem. Res. Toxicol. 2001, 14, 528.
Magnus, P.; Fortt, S.; Pitterna, T.; Snyder, J. P. J. Am. Chem. Soc. 1990, 112, 4986.
Magnus, P.; Kewis, R. T.; Huffman, J. C. J. Am. Chem. Soc. 1988, 110, 1626.
Maier, M. E.; Greiner, B. Liebigs Ann. Chem. 1992, 855.
Moore, H. W.; Chow, K.; Nguyen, N. V.; J. Org. Chem. 1987, 52, 2530.
Murrell, J. M.; Liu, W.; Shen, B. J. Nat, Prod. 2004, 67, 206.
Musch, P.W.; Remenyi, C.; Helten, H.; Engels, B. J. Am.Chem. Soc. 2002, 124, 1823.
Myers, A. G.; Dragovich, P. S.; Kuo, E. Y. J. Am. Chem. Soc. 1992, 114, 9369.
Myers, A. G.; Kuo, E. Y.; Finney, N. S. J. Am. Chem. Soc. 1989, 111, 8057.
Nadipuram, A. K.; David, W. M.; Kumar, D.; Kerwin, S. M. Org. Lett. 2002, 4, 4543.
Nagata, R.; Yamanaka H.; Okazaki, E.; Saito, I. Tetrahedron Lett. 1989, 30, 4995.
Nagata, R.; Yamanaka, H.; Murahashi, E.; Saito, I. Tetrahedron Lett. 1990, 31, 2907.
Nam, H. H.; Leroi, G. E.; Harrison, J. F. J. Phys. Chem. 1991, 95, 6514.
Negishi, E.-I.; King, A. O.; Okukado, N. J. Org. Chem. 1977, 42, 1821.
Newell, K.; Wood, P.; Stratford, I.; Tannock, I. Br. J. Cancer 1992, 66, 311.
Nicolaou, K. C.; Smith, A. L.; Yue, E. E. Proc. Natl. Acad. Sci. USA 1993, 90, 5881.
Nicolaou, K. C.; Zuccarello, G.; Ogawa, Y.; Schweiger, E. T.; Kumazawa, T. J. Am. Chem. Soc. 1988, 110, 4866.

Nicolaou, K. C.; Zuccarello, G.; Riemer, C.; Estevez, V. A.; Dai, W-M. J. Am. Chem. Soc. 1992, 114, 7360.

Ning, R. Y. Chem. Engineer News 1973, 51, 36.
Oku, N.; Matsunaga, S.; Fusetani, N. J. Am. Chem. Soc. 2003, 125, 2044.

Otani, T.; Minami, Y.; Sakawa, K.; Yoshida, K. J. Antibiot. 1991, 44, 564.
Porwoll, J. P.; Leete, E. J. Labelled. Comp. Radiopharm. 1985, 22, 257.
Prall, M.; Wittkopp, A.; Fokin, A. A.; Schreiner, P. R. J. Computational Chem. 2001, 22, 1605.

Priyadarsini, K. I.; Dennis, M. F.; Naylor, M. A.; Stratford, M. R.; Wardman, P. J. Am. Chem. Soc. 1996, 118, 5648 and references therein.

Raghunand, N.; Gillies, R. J. Drug Resis. Updates 2000, 3, 39 and references therein.
Rahm, R; Maas, G. Chem. Ber. 1994, 127, 1295.
Rawat, D. S.; Zaleski, J. M. J. Am. Chem. Soc. 2001, 123, 9675.
Refat, H. M.; Waggenspack, J.; Dutt, M.; Zhang, H.; Fadda, A. A.; Biehl, E. J. Org. Chem. 1995, 60, 1985.

Roth, W.R.; Hopf, H.; Horn, C. Chem. Ber. 1994, 127, 1765.
Rule, J. D.; Wilson, S. R.; Moore, J. S. J. Am. Chem. Soc. 2003, 125, 12992.
Schlegel, J. Mass, G. Synthesis 1999, 1, 100.
Schmidt, B. J. Org. Chem. 2004, 69, 7672.
Schmittel, M.; Keller, M.; Kiau, S.; Strittmatter, M. Chem. Eur. J. 1997, 3, 807.
Schmittel, M.; Kiau, S. Chem. Lett. 1995, 953.
Schmittel, M.; Kiau, S.; Siebert, T.; Strittmatter, M. Tetrahedron Lett. 1996, 37, 7691.
Schmittel, M.; Maywald, M.; Strittmatter, M. Synlett 1997, 165.
Schmittel, M.; Steffen, J.-P.; Auer, D.; Maywald, M. Tetrahedron lett. 1997, 38, 6177.
Schmittel, M.; Strittmatter, M.; Kiau, S. Angew. Chem., Int. Ed. Engl. 1996, 25, 1843.
Schmittel, M.; Strittmatter, M.; Kiau, S. Tetrahedron Lett. 1995, 36, 4975.
Schmittel, M.; Strittmatter, M.; Vollman, K.; Kiau, S. Tetrahedron Lett. 1996, 37, 999.
Schor, N. F. Current Therapeutics 1997, 229.
Schreiner, P. R.; Navarro-Vazquez, A.; Prall, M. Acc. Chem. Res. 2005, 38, 29.

Schreiner, P. R.; Prall, M. J. Am. Chem. Soc. 1999, 121, 8615.
Schroeder, D. R.; Colson, K. L.; Klohr, S. E.; Zein, N.; Langley, D. R.; Lee, M. S.; Marson, J. A.; Doyle, T. W. J. Am. Chem. Soc. 1994, 116, 9351.

Semmelhack, M. F.; Neu, T.; Foubelo, F. J. Org. Chem. 1994, 59, 5038.
Shabat, D.; Rader, C.; List, B.; Lerner, R. A.; Barbas, C. F.; Proc. Natl. Acad. Sci. USA 1999, 96, 6925.

Shen, B.; Liu, W.; Nonaka, K. Curr. Med. Chem. 2003, 10, 2317.
Shi, C.; Wang, K. K. J. Org. Chem. 1998, 63, 3517.
Shi, C.; Zhang, Q.; Wang, K. K. J. Org. Chem. 1999, 64, 925.
Shimizu, T.; Miyasaka, D.; Kamigata, N. Organic Lett. 2000, 2, 1923.
Sievers, E. L.; Appelbaum, F. R.; Spielberger, R. T.; Forman, S. J.; Flowers, D.; Smith, F. O.; Shannon-Dorcy, K.; Berger, M. S.; Bernstein, I. D. Blood 1999, 93, 3678.

Snyder, J. P. J. Am. Chem. Soc. 1989, 111, 7630.
Snyder, J. P. J. Am. Chem. Soc. 2000, 122, 7208.
Song, C. W.; Kim, G. E.; Lyons, J. C.; Makepeace, C. M.; Griffin, R. J.; Rao, G. H.; Cragoe, E. J. Jr. Int. J. Radiat. Oncol. Biol. Phys. 1994, 30, 1161.

Tanaka, A.; Terasawa, T.; Hagihara, H.; Sakuma, Y.; Ishibe, N.; Sawada, M.; Takasugi, H.; Tanaka, H. J. Med. Chem. 1998, 41, 2390.

Thorson, J. S. Curr. Pharm. Design 2000, 6, 1841.
Thorson, J. S.; Shen, B.; Whitwan, R. E.; Liu, W.; Li, Y.; Ahlert, J. Bioorg. Chem. 1999, 27, 172.

Thorson, J. S.; Sievers, E. L.; Ahlert, J.; Shepard, E.; Whitwan, R. E.; Onwueme, K. C.; Ruppen, M. Curr. Pharm. Des. 2000, 6, 1841.

Tilley, J. W.; Zawoiski, S. J. Org. Chem. 1988, 53, 386.
Totsuka, R.; Aizawa, Y; Uesugi, M.; Okune, Y.; Matsumoto, T.; Sugiura, Y. Biochem. Biophys. Res. Commun 1995, 208, 168.

Tsutsui, H.; Hayashi, Y.; Narasaka, K. Chem. Lett. 1997, 317.
Tsutsui, H.; Ichikawa, T.; Narasaka, K. Bull. Chem. Soc. Jan. 1999, 72, 1869.

Tuntiwechapikul. W; David, W. M.; Kumar, D.; Salazar, M.; Kerwin, S. M. Biochemistry 2002, 41, 5283.

Tusuwan, B; Kerwin, S. M. unpublished data.
Wang, J.-X.; Wei, B.; Hu, Y.; Liu, Z.; Fu, Y. Synth. Commun. 2001, 31, 3527.
Warner, B. P.; Millar, S. P.; Broene, R. D.; Buchwald, S. L. Science 1995, 269, 814.
Wenthod, P. G.; Hu, J.; Squires, R. R.; Lineberger, W. C. J. Am. Chem. Soc. 1998, 120, 5279.

Wenthold, P. G.; Lipton, M. K. J. Am. Chem. Soc. 2000, 122, 9265.
Wenthold, P. G.; Squires, R. R. J. Am. Chem. Soc. 1994, 116, 6961.
Wenthold, P. G.; Squires, R. R.; Lineberger, W. C. J. Am. Chem. Soc. 1998, 120, 5279.
Wenthold, P. G.; Wierschke, S. G.; Nash, J. J.; Squires, R. R. J. Am. Chem. Soc. 1994, 116, 7378.

Wu, J. R.; Xu, J. H.; Dedon, P. C. Biochemistry 1999, 38, 15641.
Wurthwein, E.-U.; Weigmann, R. Angew. Chem. 1987, 99, 918.
Xi, Z.; Goldberg, I. H. in Comprehensive Natural Products Chemistry; Elsevier: New York, 1999, vol 7, pp 533-592.

Xu, Y. J.; Xi, Z.; Zhen, Y. S.; Goldberg, I. H. Biochemistry 1995, 34, 12451.
Xu, Y. J.; Zhen, Y. S.; Goldberg, I. H. Biochemistry 1994, 33, 5947.
Yoshida, K.; Minami, Y.; Otani, T.; Tada, Y.; Hirama, M. Tetrahedron Lett. 1994, 35, 5253.

Yu, H.-T.; Hurley, L. H.; Kerwin, S. M. J. Am. Chem. Soc. 1996, 118, 7040.
Zein, N.; Schroeder, D. R. Adv. DNA Seq.-Spec. Agents 1998, 3, 201.
Zhen, Y. S.; Ming, X. Y.; Yu, B.; Otani, T.; Saito, H.; Yamada, Y. J. Antibiot. 1989, 42, 1294.

## Vita

Liping Feng was born in Yanchi, China on December 10, 1972, the daughter of Guanghou Feng and Zongfang Chang. After completing the degree of B.S. and M.S. from Beijing Normal University, China, in 1998, she attended Memorial University of Newfoundland, Canada, and received the degree of M.S. in 2001. In January 2001, she entered the Graduate School of The University of Texas at Austin.

## Publications:

Feng, L.; Zhang, A.; Kerwin, S. M. submitted.
Feng, L.; Kumar, D.; Birney, D. M.; Kerwin, S. M. Organic Letters 2004, 6, 2059-2062.
Feng, L; Kumar, D.; Kerwin, S. M. Journal of Organic Chemistry 2003, 68, 2234-2242.
Feng, L.; Kerwin, S. M. Tetrahedron Letters 2003, 44, 3463-3466.

Permanent address: 20 Xinde Street, Apt 1206, Xicheng District, Beijing, China, 100088

This dissertation was typed by Liping Feng.


[^0]:    ${ }^{\text {a }}$ Based on one ${ }^{1} \mathrm{H}$ NMR run in $\mathrm{CDCl}_{3}$. ${ }^{\mathrm{b}}$ Average value for two ${ }^{1} \mathrm{H}$ NMR runs in $\mathrm{CDCl}_{3}$.
    ${ }^{\mathrm{c}}$ Based on two ${ }^{1} \mathrm{H}$ NMR runs in $\mathrm{CDCl}_{3}$ and two spectrophotometric runs in $\mathrm{CHCl}_{3}$.
    ${ }^{\mathrm{d}}$ Average of two spectrophotometric runs in $\mathrm{CHCl}_{3}$. ${ }^{\text {e }}$ Based on one ${ }^{1} \mathrm{H}$ NMR run in $\mathrm{CDCl}_{3}$ and two spectrophotometic runs in $\mathrm{CHCl}_{3}$. ${ }^{\mathrm{f}}$ Data from reference 1 in $\mathrm{CH}_{3} \mathrm{CN}$.

