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The Functional Neuroanatomy of Helplessness Vulnerability

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The Functional Neuroanatomy of Helplessness Vulnerability

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Dedication

with thanks and love to my parents Jack and Dixie Shumake

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The Functional Neuroanatomy of Helplessness Vulnerability

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This dissertation had three specific aims: 1) to behaviorally characterize the congenital helplessness rat model of vulnerability to depression and post-traumatic stress disorder (PTSD); 2) to metabolically map the brains of naïve congenitally helpless adults and newborns; and 3) to map the effects of the antidepressant fluoxetine on the brains of these rats. Congenitally helpless rats, selectively bred for vulnerability to learned helplessness, were assessed on tests of exploratory activity, anxiety, circadian activity, and interest in reward; they showed increased exploration and reduced anxiety in novel environments, but also insomnia and reduced sucrose preference. This pattern of results is discussed as resembling the temperamental profile of individuals with PTSD. The brains of rats from the congenitally helpless strain were metabolically mapped with quantitative cytochrome oxidase histochemistry. Adults showed widespread metabolic abnormalities in several brain regions, but these abnormalities were bidirectional and whole-brain metabolism was not altered; the paraventricular hypothalamic nucleus (PVH), habenula, ventral hippocampus, interpeduncular nucleus, and infralimbic cortex showed increases, and the dorsomedial prefrontal cortex, basal ganglia, septal nuclei, amygdala, and ventral

tegmental area (VTA) showed decreases. In contrast, newborns showed a global reduction in brain metabolism, but the largest decreases were seen in the two regions showing the largest increases in the adult: the PVH and habenula. Newborns also showed a marked reduction in interregional functional coupling, particularly between forebrain and brainstem regions. Fluoxetine produced anti-depressant effects in the adult brain, increasing VTA metabolism and decreasing habenula metabolism. Thus, the only common region affected in adults and newborns and by fluoxetine treatment was the habenula. Functional relationships among the implicated brain systems were considered in light of the literature on depression and PTSD. It is suggested that the hypometabolism and absence of functional connectivity observed in the newborn may reflect reduced ability of forebrain regions to regulate the response of brainstem regions to stress. Consequently, the PVH may become hyperactive in the adult, and other regions may follow suit in an unsuccessful attempt to constrain its activity. The habenula is discussed as a possible nexus through which antidepressants may influence stress-responsive networks.

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Chapter 1: Modeling the Predisposition to Stress-Related Psychopathology

Helplessness, a behavioral state of passivity accompanied by feelings of uncontrollability, is a common component of several comorbid psychiatric disorders: most notably, major depression, dysthymia, post-traumatic-stress disorder (PTSD), and substance abuse. Theories concerning the role of helplessness in these disorders stem from a series of animal learning studies conducted in the early 1960s in the lab of Richard L. Solomon. Overmier and Seligman (1967) initially found that exposure to inescapable electric shocks caused dogs to switch from an active behavior—running around in a distressed manner—to a passive one: most subjects eventually lay down in a corner and whined. Subsequently, when shocks could be avoided, the dogs did not acquire the avoidance response as efficiently as control animals. The dogs appeared to have lost the behavioral flexibility to respond effectively to stressful situations. Overmier and Seligman called this phenomenon "learned helplessness" because the dogs seemed to have learned that their responses were futile. Later, Seligman and Maier (1967) validated that uncontrollability was indeed the critical factor.

LEARNED HELPLESSNESS AS A DISEASE MODEL

Based on this and other work, Seligman (1975) argued that learned helplessness in animals modeled at least some forms of human depression, noting several similar features: decreased locomotion, poor performance in appetitive tasks, decreased aggression, and loss of appetite and weight. There is even convincing evidence that learned helpless animals show attention deficits which are largely responsible for their learning difficulties (Minor, Jackson, & Maier, 1984; Lee & Maier, 1988). Indeed, if learned helpless animals have impaired attention, then the number of symptomatic parallels would warrant a legitimate diagnosis of major depression (Table 1.1) except for one thing: most of the effects of uncontrollable shock are transient, with recovery in 2 to 3 days (Maier, 1984), whereas a diagnosis of depression requires at least 2 weeks of abnormality (American Psychiatric Association, 2000). However, learned helplessness remains a valid model of depression in other respects, including the most important one from a practical standpoint: it reliably discriminates drugs with antidepressant effects (Petty, Davis, Kabel, & Kramer, 1996; Sherman, Sacquitne, & Petty, 1982).

More recently, learned helplessness has been proposed as a model of PTSD (Foa, Zinbarg, & Rothbaum, 1992; Petty, Kramer, Wu, & Davis, 1997), a psychiatric disorder that develops following exposure to one or more stressful events. Table 1.2 shows that DSM criteria for PTSD diagnosis include four major symptom clusters: reexperiencing the traumatic event, avoidance of stimuli associated with the traumatic event, numbing of behavioral responsiveness, and increased arousal (American Psychiatric Association, 2000). Certainly the inescapable shock that leads to learned helplessness could be regarded as traumatic stress, but it would be premature to conclude that learned helplessness models PTSD exclusively. Indeed, it is not even clear that depression and PTSD are distinct entities themselves, since there is considerable phenomenological overlap between PTSD and chronic depression (Newport & Nemeroff, 2000). For example, PTSD and depression share anhedonia, sleep disturbance, and impaired concentration as core diagnostic criteria, and there is also overlap in terms of restricted

Table 1.1:Summary of Diagnostic and Statistical Manual of Mental Disorders (DSM-
IV-TR) criteria for major depression, reorganized according to signs and
symptoms.

At least five symptoms must be present for at least two weeks.

One of the symptoms must be depressed mood or loss of interest or pleasure. Behavioral Signs

Decreased interest in pleasurable stimuli (e.g., sex, food, social interactions) Insomnia or hypersomnia Psychomotor agitation or retardation Decreased or increased appetite, weight loss or weight gain Decreased ability to concentrate and think, indecisiveness <u>Subjective Symptoms</u> Depressed mood Low energy, fatigue Feelings of worthlessness, excessive or inappropriate guilt

Recurrent thoughts of death and suicide

Table 1.2:Summary of Diagnostic and Statistical Manual of Mental Disorders (DSM-
IV-TR) criteria for post-traumatic stress disorder.

A. Exposure to a traumatic event. Symptoms must be present for at least one month. B. Persistent reexperience of the trauma in at least one of the following ways:

(1) Distressing recollections

(2) Distressing dreams

(3) Acting or feeling as if the traumatic event were recurring

(4) Intense distress at exposure to internal or external cues of the trauma

(5) Physiological reactivity on exposure to internal or external cues of the trauma

C. Persistent avoidance of stimuli associated with the trauma and numbing of general responsiveness, as indicated by at least three of the following:

(1) Efforts to avoid thoughts, feelings, or conversations associated with the trauma

(2) Efforts to avoid activities, places, or people that arouse recollections of the trauma

(3) Inability to recall an important aspect of the trauma

(4) Markedly diminished interest or participation in significant activities

(5) Feeling of detachment or estrangement from others

(6) Restricted range of affect

(7) Sense of foreshortened future

D. Increased arousal, as indicated by at least two of the following:

(1) Difficulty falling or staying asleep

(2) Irritability or outbursts of anger

(3) Difficulty concentrating

(4) Hypervigilance

(5) Exaggerated startle response

range of affect, impairment in personal functioning, and guilt (Vermetten, Charney, & Bremner, 2001). Moreover, 48% of PTSD patients receive a comorbid diagnosis of major depression (Kessler, Sonnega, Bromet, Hughes, & Nelson, 1995).

Regardless of whether learned helplessness models depression, PTSD, or some aspects of both, two fundamental problems remain in using the model to understand the neurobiology underpinning human psychopathology. The first, already mentioned, is the insufficient duration of helpless behavior relative to human psychiatric disorders. The problem exists whether one is attempting to model PTSD or depression, and is even worse in the case of PTSD since its diagnosis requires a month of symptomatology as opposed to two weeks (Tables 1.1 and 1.2). The second problem is that stress per se is not sufficient to cause depression, and trauma per se is not sufficient to cause PTSD.

THE IMPORTANCE OF PREDISPOSITIONAL FACTORS

Most people do not develop depression or PTSD following stress or a traumatic event. Only 6 - 15% of women develop depression following a stressful event, while 1% develop depression even in the absence of significant stress (Kendler et al., 1995). And, while nearly half of all U.S. adults experience at least one traumatic event in their lifetime, only 10% of women and 5% of men develop PTSD (Kessler et al., 1995). This points to individual differences in susceptibility to stress, and epidemiological studies show that 40%-50% of the risk for depression is genetic (Sanders, Detera-Wadleigh, & Gershon, 1999; Fava & Kendler, 2000). This makes depression a highly heritable disorder—as heritable as type II diabetes, hypertension, asthma, and certain cancers (Nestler et al., 2002). PTSD is also partially hereditary, with at least 30% of symptoms based on genetic factors (True et al., 1993). Clearly, greater knowledge of these predispositional factors would be immensely beneficial, both to identify at-risk individuals and to develop more effective treatments. However, most studies, both animal and human, have examined their subjects only after a stressful episode has taken its toll. These studies have accumulated a vast wealth of data, but without prospective biological measurements taken before illness onset, it is impossible to tease apart primary etiological factors from what are likely a voluminous number of irrelevant or compensatory factors that are consequential, not causal, in nature.

While it is possible to address these questions in humans—through studies of discordant identical twins or other healthy relatives of depressed patients, or through prospective longitudinal studies of at-risk subjects—the requisite experiments are difficult to execute in practice. Identical twins discordant for depression are not common, and longitudinal studies are inherently burdensome; they demand large amounts of time, funding, and manpower, and are inevitably plagued by high rates of subject drop-out. And clearly it would be unethical to genetically manipulate human subjects in an attempt to induce psychopathology. It is because of these practical and ethical limitations to human experimentation that animal models are vitally needed for elucidating the predisposing factors for depression and other stress-related affective disorders. The aim of this dissertation project is to employ one such model, the congenitally helpless rat, to map the brain systems underlying vulnerability to affective disorders.

CONGENITAL HELPLESSNESS AS A VULNERABILITY MODEL

Animals also show individual differences in vulnerability to stress. Henn, Johnson, Edwards, and Anderson (1985) showed that only 5% - 20% of rats develop learned helplessness following 20 minutes of 0.8 mA footshock distributed randomly within a 40 minute session (Henn, Johnson, Edwards, & Anderson, 1985). These percentages are remarkably in line with the prevalence rates of depression and PTSD in humans; moreover, helpless behavior in these susceptible rats can persist up to a month following training (Henn et al., 1985). Thus, by focusing on especially vulnerable rats, both of the problems discussed earlier—prevalence and endurance—are simultaneously addressed.

Hypothesizing that individual differences in stress vulnerability had a genetic basis, Fritz Henn and Emmeline Edwards began to interbreed animals that demonstrated both greater susceptibility and resistance to helplessness. Twenty-four hours after the inescapable shock session, the rats were tested in an escape paradigm where foot shock could be eliminated with a single lever press. Animals with more than 10 failures out of 15 trials were considered helpless, and animals with less than 5 failures were considered non-helpless. Helpless and non-helpless animals, respectively, were mated for the subsequent generations, avoiding sib crosses. The first four generations of selective breeding more than doubled the percentage of helplessness-susceptible offspring and rendered the non-helpless line of rats entirely resistant to helplessness (Henn et al., 1985). Genetic vulnerability continued to increase in the helplessness-susceptible line until a plateau was reached in the 25th generation, when 95% of rats began to show spontaneous helpless behavior in the absence of a training session (Lachman et al., 1992). Henn and Edwards named their susceptible and resistant strains "congenital learned-helpless" and "congenital non-learned-helpless," respectively. For simplicity and clarity, we have shortened these to "congenitally helpless" and "congenitally non-helpless" to emphasize that the rats used in our studies did not undergo learned helplessness training. The choice of the word "congenital" as opposed to "genetic" is important because, while the helplessness-susceptibility trait appears inborn, one cannot rule out a possible contribution of the prenatal environment to the phenotype.

Behavioral Characteristics of the Congenitally Helpless Rat

Outside of their spontaneous helpless response to electric shock, congenitally helpless rats have not been extensively characterized in terms of behavior. As one might expect, following exposure to inescapable shock, vulnerable rats display all the vegetative behaviors traditionally associated with learned helplessness (Henn et al., 1985). However, prior to this year, almost no published work has examined the premorbid temperament of congenitally helpless rats by assessing behaviors early in development and prior to significant stress.

This question is the focus of the next chapter, but there are two potential criticisms of the model that can be addressed here. In breeding for escape failures following inescapable shock, one would obviously like the selected trait to be something like a depressive phenotype. However, there are two other possibilities which must be ruled out. One is that congenitally helpless rats have a general learning deficit which has nothing to do with emotional state. Two studies have dispelled this notion by showing that, under baseline conditions, congenitally helpless rats perform as well as controls in the Morris water maze (King, Abend, & Edwards, 2001; Vollmayr et al., 2004). The other alternative is that the congenital helplessness trait is somehow specific to withstanding footshock and does not reflect a generalized coping response. This possibility was dispelled by a study showing that congenitally helpless rats express elevated levels of behavioral despair in the forced swim test (Patel, Bartoszyk, Edwards, & Ashby, Jr., 2004). Behavioral despair is similar to learned helplessness in that it entails increased immobility 24 hours after inescapable stress. However, in the forced swim test the stressor is 15 minutes of swimming instead of 20 minutes of footshock. Thus, helpless rats show increased depressive behavior in another well-validated model of depression (Porsolt, 2000; Willner, 1990), which they were not specifically bred for. This, together

with the absence of a general learning deficit, argues that the selected phenotype does correspond to an emotional response to stress. This hypothesis is further supported by other behavioral tests, which are reported in Chapter 2.

Biological Characteristics of the Congenitally Helpless Rat

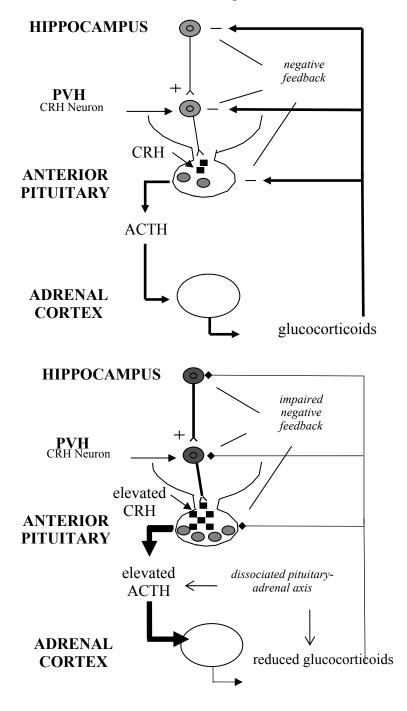
Several published studies have examined the biological characteristics of the congenitally helpless strain. Their findings have fallen into one of three major themes: 1) disruption of the hypothalamic-pituitary-adrenal (HPA) axis, 2) enhanced opiod analgesia, and 3) abnormal intracellular signaling.

Disrupted Hypothalamic-Pituitary-Adrenal Axis

Several studies suggest that congenitally helpless rats have deficient glucocorticoid signaling in response to stress, and Figure 1.1 illustrates the nature of HPA disruption in congenitally helpless rats. Lachman et al. (1993) observed that, unlike control rats, congenitally helpless rats do not alter glucocorticoid receptor mRNA expression in the hippocampus or hypothalamus in response to corticosterone administration, even though their glucocorticoid receptors are normal in terms of prevalence and binding characteristics. The exact cause of this dysfunction is unknown, but appears to involve a functional modification that affects some, but not all, glucocorticoid receptors since another glucocorticoid-inducible gene, metallothionein-1 (MT-1), does increase normally in response to corticosterone administration. There is evidence that depressed humans possess similarly aberrant glucocorticoid receptors (Calfa et al., 2003) and improve their functioning (Pariante & Miller, 2001).

8

Figure 1.1: Dynamics of the hypothalamic-pituitary-adrenal axis in response to stress for the normal (TOP) vs. the congenitally helpless rat (BOTTOM). PVH = paraventricular hypothalamic nucleus; CRH = corticotropin-releasing hormone; ACTH = adrenocorticotropin hormone



In addition to glucocorticoid receptor dysfunction, several studies have shown reduced corticosterone secretion in congenitally helpless rats, either at baseline (Edwards, King, & Fray, 1999), in response to stress (Edwards, King, & Fray, 2000; King, Campbell, & Edwards, 1993; King & Edwards, 1999), or in response to cues associated with a prior stressful experience (King et al., 2001), but only if the animals had encountered previous stress. (Stress-naïve rats showed the same hormonal response as controls.) However, the stress does not have to be severe or protracted. For example, a mere 2 hours of isolation on postnatal day 21 was sufficient to induce a robust reduction of corticosterone secretion on postnatal day 90 (King & Edwards, 1999). While most of these studies inflicted stress early in life (Edwards et al., 1999; King et al., 1993; King & Edwards, 1999), developmental timing does not appear critical since stress first encountered in adulthood also leads to adrenal hypoactivity within as few as 24 hours (Edwards et al., 2000; King et al., 2001). However, corticosterone hyposecretion in this case is not indicative of overall HPA activity since congenitally helpless rats actually show abnormally high ACTH secretion in response to stress (King & Edwards, 1999). Similar to the congenitally helpless rats, PTSD has been linked to a dissociation between the hypothalamus and the adrenal glands. At the hypothalamic level, there is hypersecretion of CRH (Baker et al., 1999), which is similar to that reported in depression. However, cortisol levels are normal to low in PTSD (Yehuda et al., 1993; Yehuda & Antelman, 1993; Yehuda et al., 1996; Yehuda, McFarlane, & Shalev, 1998; Yehuda, 1999). In conclusion, HPA function in congenitally helpless rats resembles evidence from human studies of both PTSD and depression.

Enhanced Opiod Analgesia

King, Abend, and Edwards (2001) showed that exposure to 20 min of inescapable footshock approximately doubled the pain tolerance of congenitally helpless animals

when they were re-exposed to the shock chamber or to an odor cue associated with the shock chamber. In contrast, control rats did not experience any conditioned analgesia from the chamber or odor. Similarly, Vietnam veterans re-exposed to combat trauma through videotapes experienced a 30% decrease in pain sensitivity (van der Kolk, Greenberg, Orr, & Pitman, 1989), which was reversible with naloxone, a μ -opiod receptor antagonist (Pitman, van der Kolk, Orr, & Greenberg, 1990). Interestingly, congenitally helpless rats show 3-5 times the number of μ -opiod receptors compared to congenitally non-helpless rats (Henn, Edwards, & Muneyyirci, 1993), and suicide victims show 9 times the number of μ -opiod receptors compared to controls (Gross-Isseroff, Dillon, Israeli, & Biegon, 1990).

Abnormal Intracellular Signaling

Kohen, Neumaier, Hamblin, and Edwards (2003) assessed gene expression levels of several intracellular signaling molecules and found that congenitally helpless rats showed reduced hippocampal expression of cyclic adenosine monophosphate (cAMP) response element binding protein (CREB) mRNA. This is consistent with a helpless/depressive disposition, given that hippocampal levels of CREB mRNA increase following chronic administration of antidepressants, lithium, or electroconvulsive therapy (Nibuya, Nestler, & Duman, 1996), and over-expressing CREB in the dentate gyrus of rats reduces learned helplessness and shortens immobility time in the forced swim test (Chen, Shirayama, Shin, Neve, & Duman, 2001). Kohen et al. (2003) also found increased bcl-2 mRNA levels in the prefrontal cortex. Bcl-2 is an antiapoptotic protein that promotes neuronal survival, differentiation, and neurite outgrowth (Zhang, Westberg, Holtta, & Andersson, 1996).

CANDIDATE BRAIN SYSTEMS FOR CONFERRING DEPRESSION VULNERABILITY

Regions of Interest Suggested by Human Studies

Since congenitally helpless rats display a temperament similar to that seen in depression or PTSD (see Chapter 2), we surveyed the literature on the functional neuroanatomy of these disorders. A similar set of regions has been implicated in both, including the prefrontal cortex (PFC), anterior cingulate cortex, basal ganglia, and hippocampus, which have tended to show reduced activity, and the amygdala, medial diencephalon, and cerebellum, which have tended to show increased activity. However, the results are not completely consistent from study to study.

Studies of Depression

Frontal-cingulate cortex. Abnormalities in activation of prefrontal regions in depression have been reported more frequently than for any other brain region. Depression occurs more frequently following strokes that affect the left PFC (Morris, Robinson, Raphael, & Hopwood, 1996), and structural MRI has confirmed that depression is associated with a specific reduction of PFC volume (Coffey et al., 1993). Even depressed children and adolescents show these region-specific volume reductions (Steingard et al., 1996). However, the primary evidence linking frontal cortex to depression has come from functional neuroimaging studies. PET and fMRI have consistently shown abnormalities in frontal metabolism in depressed patients, although the specific nature of these abnormalities varies as a function of the specific subregions and the symptomatic profiles of the patients assessed.

PFC regions where dysfunction might impair the ability to modulate emotional responses in mood disorders include the dorsomedial PFC (area 9), dorsolateral PFC (areas 9/46), and the area of anterior cingulate immediately dorsal to the corpus callosum

(area 24). Abnormally decreased blood flow and metabolism in these areas have been the most extensively replicated findings in depression neuroimaging (Baxter, Jr. et al., 1989a; Bench et al., 1992; Biver et al., 1994; Ebert, Feistel, & Barocka, 1991; Mayberg et al., 1997), and these metabolic deficits appear to relate to the negative thoughts (Dunn et al., 2002) and subtle impairments of attention and memory (Bench, Friston, Brown, Frackowiak, & Dolan, 1993) that accompany major depression episodes.

Located ventral to the genu of the corpus callosum, subgenual anterior cingulate has shown both metabolic decrements (Drevets et al., 1997) and mild hyperactivity (Mayberg et al., 1999) in depression. Pregenual anterior cingulate cortex, located anterior to the genu of the corpus callosum, has shown both elevated and suppressed blood flow and metabolism in untreated depression; elevated activity predicts that a patient will respond well to antidepressant treatment, and suppressed activity predicts that a patient will respond poorly (Mayberg et al., 1997).

Similar to pregenual cingulate cortex, a complex relationship exists between depression severity and metabolic activity in the orbitofrontal cortex. Although metabolic activity is abnormally increased in outpatient or treatment-responsive depressives, more severely ill or treatment-refractory inpatients either do not differ in orbitofrontal metabolism or are decreased relative to control samples (Drevets et al., 1997; Mayberg et al., 1997). In addition, remitted depressed patients who relapse under tryptophan depletion show reduced metabolism restricted to the most medial portion of orbitofrontal cortex (gyrus rectus and medial orbital gyrus; Bremner et al., 1997a).

Basal ganglia. Many human neuroimaging studies have shown reduced blood flow or metabolism in the basal ganglia of depressed patients (Austin et al., 1992; Baxter, Jr. et al., 1989b; Buchsbaum et al., 1986; Mayberg, Lewis, Regenold, & Wagner, Jr., 1994; Rogers, Bradshaw, Pantelis, & Phillips, 1998; Videbech, 2000), and structural imaging has found reduced volume of the caudate and putamen in depressed patients (Krishnan et al., 1992; Parashos, Tupler, Blitchington, & Krishnan, 1998). Lacerda et al. (2003) further found that left putamen volumes decreased as a function of length of depressive illness, and that left globus pallidum volume increased as a function of the number of depressive episodes. Basal ganglia deficits may relate to the motor changes frequently observed in depression, and psychomotor-anhedonia symptoms correlated with decreased metabolism in the anteroventral caudate-putamen (Dunn et al., 2002).

Paraventricular hypothalamus. A prominent mechanism by which the brain reacts to acute and chronic stress is activation of the hypothalamic-pituitary-adrenal axis. Neurons in the paraventricular nucleus of the hypothalamus (PVH) secrete corticotropin-releasing hormone (CRH), which signals the pituitary to release adrenocorticotropin hormone (ACTH), which in turn activates the production of adrenal steroids. In humans, the major steroid released from adrenal cortex is cortisol (corticosterone in rats), which completes a negative feedback loop by binding to brain glucocorticoid receptors and inhibiting the secretion of CRH.

HPA-axis activity is increased in depressed patients by a number of measures (Scott & Dinan, 1998; Modell, Yassouridis, Huber, & Holsboer, 1997; Rubin, Phillips, McCracken, & Sadow, 1996; Krishnan et al., 1991b). These include elevated baseline levels of plasma cortisol and ACTH, and the failure of dexamethasone, a synthetic glucocorticoid, to suppress cortisol production (Holsboer, Liebl, & Hofschuster, 1982). Current evidence suggests that hypercortisolism and dexamethasone non-suppression stem from faulty corticosteroid receptors in regulatory structures such as the hippocampus, which fails to relay the negative feedback signal from cortisol that normally constrains the stress response (Pariante & Miller, 2001). Consequently, the paraventricular nucleus of the hypothalamus (PVH) is thought to become hyperactive,

and postmortem studies of depressed patients have demonstrated increased numbers of peptide-containing neurons in the PVH—such as arginine vasopressin (AVP)immunoreactive neurons, oxytocin (OXT)-expressing neurons, and corticotropinreleasing hormone (CRH) neurons—as well as increased expression of CRH mRNA, which would permit more CRH synthesis in the PVH (Purba, Hoogendijk, Hofman, & Swaab, 1996; Raadsheer, Hoogendijk, Stam, Tilders, & Swaab, 1994). One would therefore expect PVH metabolism to be altered in an individual predisposed to depression.

Hippocampus. The hippocampus, with its high concentration of glucocorticoid receptors, likely mediates corticosterone negative feedback to the HPA axis (Sapolsky, Krey, & McEwen, 1986). Hippocampal volume reductions have been found in several magnetic resonance imaging (MRI) studies of patients with depression (Bremner et al., 2000; Krishnan et al., 1991a; Sapolsky, 2000; Shah, Ebmeier, Glabus, & Goodwin, 1998; Sheline, Wang, Gado, Csernansky, & Vannier, 1996); however, hippocampal function has rarely been directly assessed with functional neuroimaging. Only one study has reported reduced left hippocampal metabolism, which negatively correlated with depression severity (Saxena et al., 2001).

Amygdala. Abnormal elevations of resting regional cerebral blood flow or glucose metabolism in the amygdala have been reported in depression during both wakefulness (Drevets et al., 1992) and sleep (Ho et al., 1996; Nofzinger et al., 1999). Furthermore, metabolic rate in the amygdala correlated with depression severity (Drevets et al., 1992) or negative affect (Abercrombie et al., 1998) in depressed patients. However, amygdala changes have not been observed in most studies and may only be present in patients with comorbid anxiety disorders (Davidson et al., 2002).

Thalamus and habenula. Few studies have examined thalamic metabolism in depression, but, when it has been assessed, hypermetabolism has been found (Drevets et al., 1992; Morris, Smith, Cowen, Friston, & Dolan, 1999; Saxena et al., 2001). Of these three studies, two found that the thalamic hypermetabolism was localized to the dorsal midline (Drevets et al., 1992; Morris et al., 1999), and Morris et al. (1999) further localized the source to a region immediately superior and lateral to the posterior commissure. This region most likely corresponds to the habenula nucleus.

Studies of Post-Traumatic Stress Disorder

Compared to depression, far fewer neuroimaging studies have been conducted on PTSD, and the majority of these have employed an anxiogenic challenge paradigm (e.g., evoking traumatic memories). These studies frequently found increased activity in the premotor frontal region and decreased activity in the anterior cingulate and middle temporal gyri (Bremner et al., 1999b; Lanius et al., 2001; Liberzon et al., 1999; Rauch et al., 1996; Shin et al., 1999; Shin et al., 2001) although patients reporting dissociative symptoms showed increased activity in all these regions (Lanius et al., 2002). Some have also found amygdala activation (Rauch et al., 2000; Shin et al., 1997b; Shin et al., 1997a); however, most have not (e.g., Bremner et al., 1999b; Bremner et al., 1999a; Shin et al., 1999).

Only a few studies of resting state metabolism have been performed. Some were done on an exclusive substance abuse population and will not be discussed (Semple et al., 1993; Semple et al., 1996). Other resting state studies normalized their results to the cerebellum, which was assumed to have no relevance to PTSD (Lucey et al., 1997; Sachinvala, Kling, Suffin, Lake, & Cohen, 2000). However, because a subsequent study found this assumption to be invalid, these results will not be discussed either. This leaves only one resting state study (Bonne et al., 2003), which found that increased perfusion in

the following regions discriminated PTSD trauma survivors from healthy trauma survivors: the cerebellum, the left inferior lateral temporal lobe, and the left supramarginal and postcentral gyrus. Interestingly, the same study showed that reduced cortisol signaling predicted increased hippocampal and decreased frontal-cingulate activity in those vulnerable to PTSD. Structural MRI studies revealed reduced hippocampal volume in PTSD patients (Bremner et al., 1997b; Gurvits et al., 1996; Stein, Koverola, Hanna, Torchia, & McClarty, 1997; Villarreal et al., 2002). Interestingly, Gilbertson et al. (2002) found reduced hippocampal volume in PTSD patients' monozygotic twins who never experienced trauma, indicating that this may be a vulnerability marker.

Regions of Interest Suggested by Animal Studies

There are over a dozen animal models of depression (Willner, 1990), and reviewing the neuroscientific literature on all of them is beyond the scope of this project. Therefore, with the exception of one metabolic mapping study (which was relevant because of the methodology), our survey of the animal literature was limited to studies of brain regions which mediate the development and maintenance of learned helplessness. Collectively, the animal literature points to the hippocampus, septal nuclei, raphe nuclei, and habenula as being important players in helpless behavior.

Studies of Learned Helplessness

The hippocampus was the first brain region linked to the development of learned helplessness, which was blocked by fornix transection (Leshner & Segal, 1979). Many subsequent studies have also implicated hippocampal activation in the development of learned helplessness. For example, Petty and Sherman (1981) found that inescapable shock increased glutamate and decreased GABA release in the hippocampus. As

discussed in the first two chapters, dysfunctional glucocorticoid receptors specific to the hippocampal region appear to confer vulnerability to learned helplessness (Lachman et al., 1993; Papolos et al., 1993).

The related septal area was the second region shown to play a role in learned helplessness. Sherman and Petty (1980) found that learned helplessness was associated with decreased septal serotonin (5-HT) release and that 5-HT microinjected into the lateral septal nucleus (LS) reversed learned helpless behavior. Furthermore, rats which proved vulnerable to learned helplessness showed reduced c-fos expression in the LS (Steciuk, Kram, Kramer, & Petty, 1999), while rats which proved resistant to learned helplessness showed increased 5-HT metabolism in the LS (Ronan, Steciuk, Kramer, Kram, & Petty, 2000).

The other major region implicated in learned helplessness development is the dorsal raphe nucleus (DR). Enhanced 5-HT release from the DR appears to be a prerequisite for the development of learned helplessness (Amat et al., 2001; Grahn et al., 1999b; Grahn et al., 2002; Grahn, Watkins, & Maier, 2000; Grahn, Maswood, McQueen, Watkins, & Maier, 1999a; Greenwood et al., 2003; Hammack et al., 2003; Hammack, Pepin, DesMarteau, Watkins, & Maier, 2003; Hammack et al., 2002; Maier, Grahn, & Watkins, 1995; Maier, Kalman, & Grahn, 1994; Maier et al., 1993; Maswood, Barter, Watkins, & Maier, 1998; Sutton, Grahn, Wiertelak, Watkins, & Maier, 1997). While several neurotransmitters and neuropeptides (including norepinephrine, opiods, corticotropin-releasing factor, and urocortin) appear to mediate increased DR activity in response to inescapable stress (Grahn et al., 1999a; Grahn et al., 2002; Hammack et al., 2002; Hammack et al., 2003), the only specific brain region demonstrated to play a role in this is the habenula. Amat et al. (2001) found that lesioning the habenula prevented the

rise in DR 5-HT following inescapable shock and blocked the development of learned helplessness.

The amygdala may also play a role in some aspects of helpless behavior since lesions of the basolateral and central nuclei reduce immobility following inescapable shock. However, these lesions do not prevent the development of learned helplessness (Maier et al., 1993), but lesions of the related bed nucleus of stria terminalis (BNST) do prevent the development of learned helplessness (Hammack, Richey, Watkins, & Maier, 2004).

Metabolic Mapping of Three Models of Depression

Outside of our own work, only one other study has applied the metabolic brain mapping approach to study animal models of depression. Caldecott-Hazard, Mazziotta, and Phelps (1988) used $_{14}$ C-2-deoxyglucose (2-DG) autoradiography to examine the cerebral correlates of three depression models: amphetamine withdrawal, chronic stress, and α -methyl-para-tyrosine challenge (which reduces catecholamine activity). The only region that showed elevated metabolism across all three models was the lateral habenula, but several regions showed reduced metabolism in each model: the dorsal medial prefrontal cortex, anterior ventral nucleus of the thalamus (AV), and inferior colliculus. Forebrain global metabolic rates were also reduced in each of the models. In addition, reduced caudate metabolism was observed in two of the models.

OVERVIEW

The aim of this dissertation is three-fold: first, to behaviorally characterize the congenital helplessness model; second, to metabolically map the brains of naïve congenitally helpless adults and newborns; and third, to map the effects of the antidepressant fluoxetine on the brains of these rats.

Chapter 2 discusses the behavioral performance of congenitally helpless rats on tests of exploratory activity, anxiety, circadian activity, and interest in reward. Chapter 3 discusses the general methodology used in the subsequent metabolic mapping studies and includes background on quantitative cytochrome oxidase histochemistry and the statistical techniques used to analyze functional imaging data.

The remaining chapters report the results of the metabolic mapping studies. Chapter 4 discusses the brain map of naïve congenitally helpless adults, and Chapter 5 discusses the brain map of congenitally helpless newborns. Chapter 6 discusses changes in brain metabolism induced by chronic administration of fluoxetine. Finally, Chapter 7 summarizes and integrates the major findings of the dissertation project and discusses their theoretical predictions and implications.

Chapter 2: Behavioral Characterization of the Helpless Temperament

Outside of King et al. (2001) and Vollmayr et al. (2004), no one has examined baseline behavioral differences in the congenitally helpless strain. Such behavioral tests are important for establishing possible temperamental traits that predict vulnerability to stress in these animals, and for interpreting any brain differences they might show. We assessed the rats on a variety of behavioral parameters, including exploratory activity in the open field, anxiety in the light-dark test, circadian running wheel activity, and interest in sucrose reward. We decided to focus on adolescent, as opposed to adult rats, for a number of reasons. First and foremost, from a clinical standpoint, it would be useful to identify individuals at risk for psychopathology during childhood and adolescence prior to adult onset of disease, and temperamental markers present during this period of development in congenitally helpless rats may inform this question. Second, Vollmayr et al. (2004) have already verified open-field and sucrose consumption differences in adult congenitally helpless rats. Third, for this study, we wished to avoid the influence of estrous cycling on the behaviors of female rats. Finally, rats are more exploratory and playful during this period of development, which may be an ideal time for assessing novelty-seeking behaviors (Spear, 2000).

Experimental subjects were congenitally helpless males (n = 23) and females (n = 21) bred in our laboratory, and control subjects were Sprague-Dawley males (n = 12) and females (n = 12) obtained from a commercial animal supplier (Harlan). Congenitally helpless rats were separated from their mothers on postnatal day 21, and control rats were shipped to us at the same age. All subjects were group housed 3-5 to a cage ($45 \times 24 \times 21$ cm) in a temperature-controlled room ($22 \pm 1^{\circ}$ C). They were maintained on a 12/12 hour light cycle and provided continuous food and water.

Rats were handled daily for 5 min during postnatal days 22-28, and behavioral testing was conducted from postnatal day 29 to 48. Open-field and light-dark activity were assessed on postnatal days 29-31. Sucrose consumption was assessed prior to running wheel activity, but the exact start date varied due to limited testing equipment relative to number of subjects. Thus, some animals had sucrose testing on days 32-33 and running-wheel testing on days 34-35; others were delayed by intervals of 2 days, with the latest interval being sucrose testing on days 45-46 and running-wheel testing on days 47-48. Each testing period had equal numbers of each group and sex.

Note that neither these rats, nor their parents, were actually trained for learned helplessness because naïve subjects are needed in order to examine behavioral predispositions. Only their grandparents, obtained from the Central Institute for Mental Health (Mannheim, Germany), were verified as possessing the helplessness phenotype. However, it is typical for 88% of male and 94% of female offspring to express congenital helplessness (Edwards et al., 2000), and our larger sample size for these rats should compensate for the presence of a few outliers. The methodology and results of each test are discussed separately below.

OPEN-FIELD AND LIGHT-DARK ACTIVITY

The open field test is one of the most popular procedures in animal psychology (Prut & Belzung, 2003). Different versions are available, but the procedure generally consists of subjecting a rodent to an unknown environment from which escape is prevented by surrounding walls. In such a situation, rodents spontaneously prefer the periphery of the apparatus to activity in the central parts of the open field. This behavior is called thigmotaxis and is considered an index of anxiety since many anxiolytic drugs reduce it, increasing time spent in the central part of the field (Prut & Belzung, 2003). General locomotion and rearing activity appear to reflect level of arousal since stimulant

drugs increase locomotion and sedatives decrease rearing (Prut & Belzung, 2003). Based on the diagnostic criteria for depression, one might predict either increased or decreased open field activity in an animal model, since both psychomotor agitation and retardation have been linked to depression. The increased arousal observed in PTSD, however, would predict increased open field activity (American Psychiatric Association, 2000), but, because of their predisposition to become immobile when subjected to stress, one might expect congenitally helpless rats to express reduced locomotion in the open field. However, Vollmayr et al. (2004) found that they in fact show increased locomotion, but only during the first 5 minutes of the open field test. Thus, congenitally helpless rats do not show persistent hyperactivity or hypoactivity as part of their premorbid phenotype, but are temperamentally predisposed to increase locomotion in response to novelty.

The present experiment attempted to replicate this finding and extend the results to include measures of rearing and anxiety, which were not assessed by Vollmayr et al. (2004). In addition to thigmotaxis, on the second day of testing we used a variant of the open field called the light-dark test to assess anxiety. In this version, half of the field is covered by a dark box with a small opening, and the animal is allowed to move freely between the bright and dark compartments. This provides an important additional perspective on the animal's temperament since exploration of the uncovered area is voluntary. Simply put, it resembles a forced-choice test of personality type in humans: animals that show a strong preference for the dark field may be characterized as anxious or withdrawn, and animals that spend increased time in the light field have been characterized as impulsive or sensation-seeking (Kabbaj, 2004). Intuitively, one might expect that individuals predisposed to depression or PTSD would show increased withdrawal and anxiety, so we predicted that the congenitally helpless rats would show increased fearfulness on these behavioral measures. Finally, we repeated the open field test on the third day as a more stringent test of whether any group differences observed on the first day were due to novelty.

Method

An automated activity monitoring system from MED Associates (St. Albans, Vermont, USA) recorded various locomotion indices in a testing room illuminated at 100 lux. The 43.2 cm² chamber consisted of clear plastic sides 30.5 cm high and a white plexiglass floor. Parallel arrays of infrared motion detectors were located 2.5 cm and 10 cm above the chamber floor to record horizontal and vertical motion, respectively. The chambers were cleaned with a mild detergent between animals.

Three 10-minute sessions were conducted over 3 consecutive days. Each session began by placing the rat in the same corner of the field. On days 1 and 3 (referred to as the novel and familiar open-field sessions, respectively), the chamber was completely open. Measures included ambulatory distance, rearing (vertical counts and time), velocity, and time spent in the periphery vs. the center, defined as 38% of the area. On day 2 (referred to as the light-dark field session), a dark plexiglass insert covered half of the chamber, and a small hole permitted free access to either the open or covered compartments. The same activity parameters were recorded as in the open field, but, instead of time spent against the walls relative to time spent in the center, fearfulness was operationalized as time spent in the dark relative to the light. In addition, latency to first exit the dark chamber was recorded.

Results

With the exception of rearing duration and exit latency measures (see below), open-field data were analyzed with a $2 \times 2 \times 3 \times 10$ (Group \times Sex \times Session \times Time) repeated measures multivariate analysis of variance (MANOVA), with session (novel,

light-dark, or familiar open field) and time (in minutes) serving as repeated measures. All measures showed significant main effects of session and time: subjects always decreased activity over time within sessions, and, between sessions, they were most active in the light-dark open field, least active in the novel open field, and intermediately active in the familiar open field. However, these effects were of little interest, so the supporting data is not discussed. Sex effects were of interest; however, no significant sex differences or interactions were found for any of the analyzed variables, so the group means reported in the figures include both sexes. Due to the large number of effects and variables assessed, for sake of simplicity only group main effects and interactions showing statistical significance (p < .05) are reported.

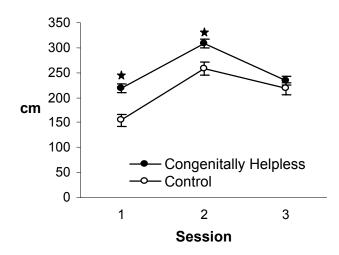
Ambulation

Repeated measures of ambulatory distance showed a significant two-way interaction (Group × Session), F(2, 66) = 3.9, p = .025. Figure 2.1A shows that within both the novel and light-dark open fields, helpless rats ambulated significantly more than controls, Fs(1, 67) = 18.9 and 9.98, ps = 0.00005 and .002, respectively. However, helpless and control rats were not significantly different in the familiar open field, F(1, 67) = 1.16, p = .29.

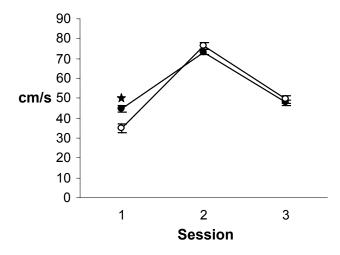
Concurrent measures of ambulatory velocity showed the same significant interaction (Group × Session), F(2,66) = 7.49, p = .001. Figure 2.1B shows that within the novel open field, helpless rats ambulated significantly faster than controls, F(1,67) = 12.7, p = .001. However, helpless and control rats were not significantly different in either the light-dark or familiar open fields, Fs(1, 67) = 2.37 and .436, ps = .128 and .511, respectively.

Figure 2.1: Ambulatory measures from activity chamber averaged across 10 minutes. Sessions 1, 2, and 3 refer to the novel open field, light-dark field, and familiar open field, respectively. $\star p < .01$





B. Mean Velocity Across Sessions



Rearing

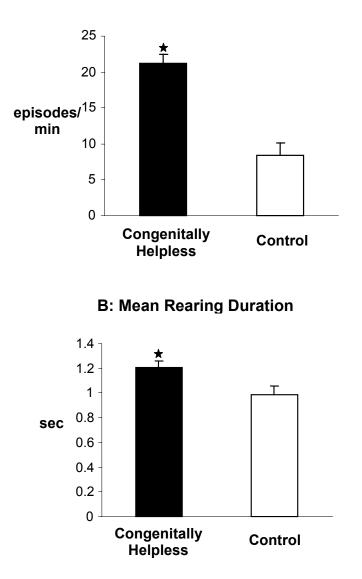
A significant three-way (Group × Session × Time) interaction was obtained for rearing counts because the two groups grew somewhat less different over time, both within and between sessions, F(18, 50) = 2.2, p = .02. However, this effect was of little interest because congenitally helpless rats continued to show significantly greater rearing than controls at every time point in every session, F(1,67) = 35.4, p = 0.0000001. Figure 2.2A shows the group means collapsed over time and session.

In addition to number of rearing episodes, the average duration of these episodes was assessed by dividing total time spent rearing by the total number of rearing episodes. Because several subjects did not rear at all during some time points, within-session repeated measures analysis could not be performed. Between-session repeated measures MANOVA showed only a main effect for group, F(1,67) = 6.89, p = .01. Figure 2.2B shows that congenitally helpless rats had longer rearing episodes than did controls.

Fearfulness

In the novel and familiar open fields, fearfulness was operationalized as the percentage of time spent against the walls (and not in the center), and, in the light-dark open field, the percentage of time spent under cover (and not in the open). While these two measures are technically different, they both reflect the relative preference of a defensive zone over a more vulnerable zone. Based on this conceptual similarity, we analyzed these fear indices as repeated measures. A significant two-way interaction (Group × Session) was obtained, F(2, 66) = 8.423, p = .001. Figure 2.3A indicates that the control group showed a substantial reduction in fearfulness following the first session, whereas congenitally helpless rats remained about the same. Thus, the groups showed a trend toward convergence across sessions. Indeed, simple-effects tests showed that, while congenitally helpless rats appeared less fearful across all three sessions, the difference

Figure 2.2: Rearing measures from activity chamber averaged across three 10 minute sessions. $\star p < .01$



A: Mean Rearing Frequency

was only significant in the novel and light-dark open fields, but not in the familiar open field, Fs(1, 67) = 39.8, 7.14, and 2.95, ps = 0.00000003, .009, and .09, respectively.

One additional fearfulness measure unique to the light-dark open field was the latency to first exit the dark chamber (Figure 2.3B), which was analyzed with a univariate ANOVA. This measure also indicated less fearfulness in the congenitally helpless group, with control rats taking much longer to exit the dark chamber than helpless rats, F(1, 67) = 23.7, p = 0.000006.

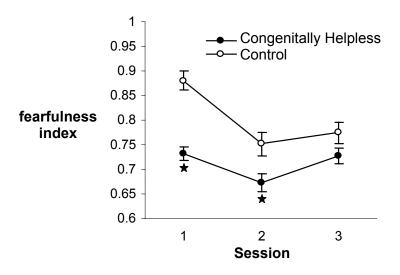
RUNNING WHEEL ACTIVITY

Monitoring voluntary activity in a running wheel for two days served as another test of general motor differences as well as a measure of circadian fluctuations in activity. Circadian rhythm has not been previously assessed in the congenitally helpless rats, but, since disruption of the sleep-wake cycle is an almost ubiquitous feature of anxiety and mood disorders (American Psychiatric Association, 2000), one might expect to observe sleep loss or a circadian phase shift in the congenitally helpless group.

Method

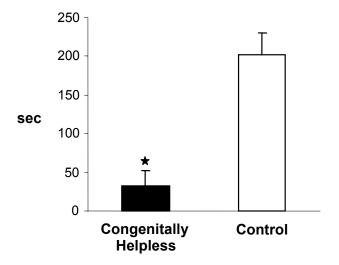
At the beginning of the dark cycle (1800 hours), rats were placed in the holding compartments of standard activity wheels. The door between the holding cage and wheel was open throughout the entire 48 hour session to allow continuous access to food and water. The numbers of revolutions were recorded at 600, 1000, 1400, and 1800 hours for the next 2 days. Thus, one reading was obtained for each 12 hour dark cycle, and 3 readings were obtained for each third of each light cycle.

Figure 2.3: Fearfulness measures from activity chamber. A: In sessions 1 (novel open field) and 3 (familiar open field), fear index is percentage of time spent against the walls (not in center). In session 2 (light-dark field), fear index is percentage of time spent in dark (not in light). B: In light-dark field, mean latency to first exit dark chamber after start of session. $\star p < .01$.



A. Fearfulness Across Sessions





Results

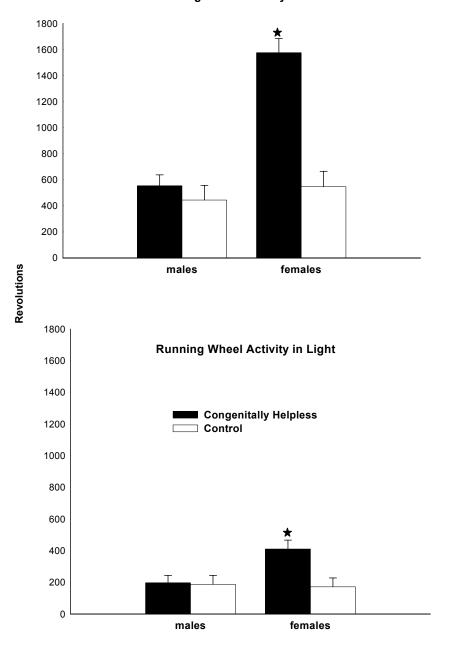
Absolute Light and Dark Activity

Data screening revealed 7 outliers (2 helpless males, 2 helpless females, 1 control male, and 2 control females), all associated with the same running wheel, which was subsequently found to have a malfunctioning counter that recorded approximately 2 revolutions for every actual revolution. These subjects were dropped from the analysis of absolute activity, which consisted of a $2 \times 2 \times 2 \times 2$ (Group × Sex × Session × Light vs. Dark) repeated measures MANOVA, with Session and Light vs. Dark serving as the repeated measures. A significant three-way (Group × Sex × Light vs. Dark) interaction was obtained, F(1, 60) = 5.28, p = 0.025. Figure 2.4 shows this interaction as 2 two-way (Group × Sex) interactions, one for the dark cycle and one for the light cycle. The three-way interaction indicates that helpless and control groups diverged more in females than males, and more in the dark than in the light. Simple-effects tests of the interaction showed that helpless males were not significantly different from control males in either the light or the dark, Fs(1, 60) = .517 and .001, ps = .47 and .97, respectively. In contrast, helpless females were significantly more active than control females in both the light and dark, Fs(1, 60) = 31.9 and 13.0, ps = 0.0000005 and 0.0006, respectively.

Relative Activity Across the Light Cycle

In order to assess circadian changes in activity independent of total activity, each subject's light-cycle activity (divided into three 4 hour intervals) was divided by its prior dark-cycle activity. Because this transformation normalized each subject for absolute differences in activity, the subjects excluded from the previous analysis were not outliers in this one. Therefore, all subjects were analyzed with a $2 \times 2 \times 2 \times 3$ (Group \times Sex \times Session \times Time) repeated measures MANOVA, with session (first or second 24-hour

Figure 2.4: Mean revolutions in running wheel as a function of group, sex, and phase of light-dark cycle. $\star p < .001$.



Running Wheel Activity in Dark

cycle) and time (early, middle, or late day) serving as repeated measures. A significant three-way (Sex × Session × Time) interaction was obtained, F(2, 66) = 5.02, p = .009. The three-way interaction indicates that, in both groups, females showed more light-relative-to-dark activity than males across the entire first light cycle, but only in the early third of the second light cycle (data not shown). A significant two-way (Group × Time) interaction was also obtained, F(2, 66) = 3.79, p = .028. Simple-effects tests of the interaction revealed that, for both sexes in both sessions, congenitally helpless rats showed significantly more light-relative-to-dark activity than controls only in the early phase of the light cycle, F(1, 67) = 9.23, p = .003, but not in the middle or late phase of the light cycle, Fs(1, 67) = .396 and .537, ps = .53 and .47, respectively. Figure 2.5 shows that this interaction is not merely a phase shift in circadian rhythm because helpless rats, like controls, began to increase activity nearing the end of the light cycle.

SUCROSE CONSUMPTION

Anhedonia, or decreased responsiveness to rewards, is a core symptom of major depression and PTSD (American Psychiatric Association, 2000). Responsiveness to reward is usually monitored by consumption of a weak sucrose solution, a test that was developed in conjunction with the chronic mild stress model of depression (Willner, Towell, Sampson, Sophokleous, & Muscat, 1987). Vollmayr et al. (2004) previously showed that, under a progressive ratio schedule of reinforcement (in which the amount of operant responding required to receive reward is steadily increased), congenitally helpless rats were less willing to work for a 7% sucrose reward, pressing the operant bar 4 times less frequently than controls. Furthermore, congenitally helpless rats gave up entirely if more than 7 presses were required, whereas control rats were willing to press the lever up to an average of 18 times. This study already establishes that congenitally

helpless rats show reduced reward-seeking motivation. Here, we tested if this motivational deficit would also manifest as a simple reduction in sucrose drinking time.

We used a modified version of a protocol developed by Willner et al. (1987), who tested consumption of a 1% sucrose solution following 14 hours of food and water deprivation. While chronic stress reduces consumption under these conditions, the extremely weak sucrose concentration and the presence of what should be significant thirst and hunger suggest a more fundamental disruption in sensory discrimination or homeostatic drive. A pure deficit in reward seeking should, in our opinion, express itself under baseline conditions and in the face of a more potent reward. We therefore used a sweeter 5% solution, which we tested under both baseline and deprived conditions. Also, instead of measuring change in solution volume as an index of consumption, we monitored instances of licking using an automated system, which tracked changes in drinking behavior over time.

Method

Baseline consumption of a 5% sucrose solution was assessed for 1 hour in four 30 $\times 25 \times 20$ cm operant chambers (Med Associates, St. Albans, VT) where rats had free access to a bottle containing the sucrose solution. The bottles were specially designed by Med Associates for use in these operant chambers, and a light beam passing in front of the tip of the bottle registered the time the rat spent drinking, divided into six 10 minute bins. Rats were then water deprived for 20 hours, and the same procedure was repeated. Thus, rats were tested in a low (non-deprived) motivational state and a high (deprived) motivational state.

Pilot data previously showed that the four operant bottles supplied by Med Associates consistently yielded different drinking times as a consequence of different flow-rates: each bottle required significantly more or less drinking time to obtain an equivalent volume. Therefore, bottle assignment was counterbalanced with respect to group and sex, and flow rate (inferred from the pilot data) was used as a covariate in the statistical analysis to reduce error.

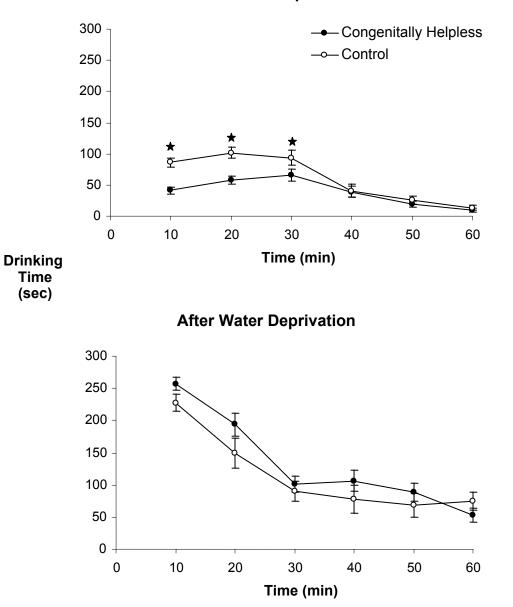
Results

Drinking time data were analyzed with a 2 × 2 × 2 × 6 (Group × Sex × Motivational State × Session Time) mixed analysis of covariance, with water bottle serving as a covariate and motivational state and session time serving as repeated measures. A significant three-way (Group × Motivational State × Session Time) interaction was obtained, F(5, 63) = 3.63, p = .006. Figure 2.6 shows this interaction as 2 two-way (Group × Session Time) interactions, one for each motivational state. The three-way interaction indicates that congenitally helpless rats spent less time drinking sucrose water than controls, but only during the first half of the baseline session, F(1, 67) = 22.5, p = 0.00004. In contrast, the groups did not differ in either the second half of the baseline session, F(1, 67) = .405, p = 1, or in the entire session following water deprivation, F(1, 67) = 2.11, p = .30.

GENERAL DISCUSSION

Congenitally helpless rats showed many behaviors consistent with diagnostic criteria for both depression and PTSD, even though they were adolescents and had never been subjected to traumatic stress. However, one potential confound is that, unlike the congenitally helpless rats, the control subjects were not bred in our laboratory but rather obtained from a commercial supplier. While it is possible that the stress of shipping could have impacted the behavior of these animals, Vollmayr et al. (2004) obtained similar results using congenitally non-helpless rats as a control. In addition, unpublished data from a separate experiment looking at the effects of maternal separation on the same

Figure 2.6: Mean time that congenitally helpless and control rats spent drinking a 5% sucrose solution, as a function of session time and level of motivation. $\star p < .01$.



Before Water Deprivation

behavioral outcome measures during the same developmental period indicates that the control group, which was born in our laboratory, was similarly different from congenitally helpless rats on all of the measures. Another factor which may have contributed to behavioral differences between congenitally helpless rats and controls is a possible difference in maternal care between congenitally helpless and control mothers. It is not known whether congenitally helpless mothers differ in their maternal care, so this remains a topic for further investigation.

Perhaps the most important feature of the depressive phenotype is reduced pursuit of reward. Vollmayr et al. (2004) previously established that adult congenitally helpless rats were less motivated to work for sucrose reward. We further found that, under baseline conditions, adolescent congenitally helpless rats maintained a low but steady level of sucrose consumption throughout the testing session, in contrast to the normally observed shift from high consumption at the beginning of the session to low consumption at the end. Thus, helpless rats were less reactive to sucrose reward than controls.

We also found that congenitally helpless rats were markedly more active during the first third of their light cycle than controls, who appeared to sleep for much of this period. While both sexes of congenitally helpless rats showed this same circadian pattern, females were far more active at all time points. Despite the fact that females are 2-3 times more vulnerable to depression and PTSD than males, this was the only reliable sex difference we observed in any of the behavioral tests, and its meaning is unclear. The overall absence of sex differences may be explained by the prepubescent age at which they were tested. Thus, one should not conclude that sex differences in the helplessness phenotype do not exist because differences could still emerge later in life.

In addition to finding reward insensitivity and sleep disruption in naïve congenitally helpless rats, we replicated Vollmayr et al.'s (2004) finding of hyperactivity

in the novel open field. Unlike Vollmayr et al., we found that this hyperactivity persisted for the duration of the novel session, but novelty-dependent hyperactivity was supported by the finding that hyperactivity was no longer present by the third exposure to the activity chamber.

Surprisingly, by every measure, the congenitally helpless rats were much more exploratory and less fearful than controls. Intuitively, increased exploration of a novel environment seems inconsistent with reduced reward sensitivity and the intense immobility that these rats show in response to shock. However, it is consistent with a body of work led by Mohamed Kabbaj, in collaboration with Huda Akil, examining rats which exhibit high rates of exploratory locomotion when exposed to novel environments (Kabbaj, Devine, Savage, & Akil, 2000; Kabbaj & Akil, 2001; Kabbaj, 2004). These rats, termed high responders (HR), are virtually identical to congenitally helpless rats in terms of their open-field and light-dark behavior. Moreover, they resemble congenitally helpless rats in expressing reduced glucocorticoid receptor mRNA in the hippocampus. In fact, impaired glucocorticoid signaling appears responsible for the novelty-induced hyperactivity since injection of RU38486, a glucocorticoid antagonist, into the hippocampus essentially turned low-responding (LR) rats into HR rats (Kabbaj et al., 2000).

The HR rat was originally proposed as a model of vulnerability to substance abuse since these rats exhibit higher rates of amphetamine- and cocaine-induced locomotor activity and self-administer these drugs at lower doses than LR rats (Hooks, Jones, Smith, Neill, & Justice, Jr., 1991; Piazza, Deminiere, Le Moal, & Simon, 1989). However, their behavioral and neural similarities to congenitally helpless rats may help to account for the comorbidity of substance abuse, PTSD, and depression, with impaired glucocorticoid signaling being a common denominator. This is supported by the finding that the same experimental manipulation which converts LR rats into HR rats hippocampal administration of RU38486—increases vulnerability to learned helplessness, causing normal rats to respond as if they were congenitally helpless (Papolos, Edwards, Marmur, Lachman, & Henn, 1993).

It still seems paradoxical, though, that increased exploratory behavior and reduced anxiety should co-exist with reduced reward sensitivity and increased helplessness vulnerability. However, a large body of evidence from family, personality, pharmacological, and behavioral conditioning studies suggest that these behaviors correspond to dimensions of temperament which vary independently: novelty seeking, reward dependence, and behavioral inhibition (Cloninger, 1987). Operationally defined, novelty seeking is excitation in response to novel stimuli leading to exploratory behavior; reward dependence is intense responding to signals of reward leading to approach behavior; and behavioral inhibition is intense responding to aversive stimuli leading to passive avoidance. Cloninger actually refers to this last dimension as "harm avoidance," but he meant a specific kind of harm avoidance: inhibition of a punished behavior. This raises an interesting issue regarding the congenitally helpless strain, which was selected for an impairment in active avoidance, or learning to initiate a novel response to avoid harm. But this does not necessarily mean that these rats are insensitive to aversive stimuli. It is possible that they are unimpaired or perhaps even superior at passive avoidance, or learning to inhibit a punished response to avoid harm—the kind of harm avoidance that Cloninger refers to. For this reason, it is perhaps more accurate and certainly less confusing to refer to this temperament dimension as "behavioral inhibition."

Our behavioral survey of the congenitally helpless rats suggests that they are high on novelty seeking and low on reward dependence, and they were essentially bred for high behavioral inhibition. Interestingly, two studies have found that Vietnam veterans with PTSD show an identical temperament profile when assessed with Cloninger's Tridimensional Personality Questionnaire (Richman & Frueh, 1996; Wang et al., 1997), and the authors conjecture that high novelty seeking, high behavioral inhibition, and low reward dependence confer trait vulnerability to PTSD. Moreover, all three dimensions correlated with severity of PTSD symptoms, with novelty seeking being the strongest correlate (Richman & Frueh, 1996). It seems counterintuitive that increased novelty seeking should predict vulnerability to an anxiety disorder, much as it seems counterintuitive that exploratory behavior should predict vulnerability to learned helplessness. This probably stems from the assumption that anxious individuals are stressed by unfamiliar settings, and that novelty-seeking individuals are not. However, this assumption is not supported by the facts.

In actuality, HR rats experience an abnormally intense activation of the HPA axis in response to their self-initiated exploratory activity (Kabbaj et al., 2000; Kabbaj & Akil, 2001). Similarly, Kendler, Karkowski, and Prescott (1999) found that individuals genetically predisposed to major depression paradoxically select themselves into highstress environments which put them at even greater risk of developing depression. Thus, it seems that animals and humans prone to stress-related psychopathology are likely to be novelty seekers, and novelty seeking is likely to trigger stress-related psychopathology.

From a folk psychology standpoint, this conclusion appears nonsensical. However, from a neuroendocrine standpoint the combination of high novelty seeking, low reward dependence, and vulnerability to helplessness, depression, and PTSD makes complete sense, since all these things have been linked to reduced glucocorticoid signaling. As already discussed, glucocorticoid antagonism increases novelty seeking and vulnerability to helplessness in rats (Kabbaj et al., 2000; Papolos et al., 1993). In addition, urinary cortisol levels negatively correlate with novelty-seeking scores on the Cloninger Tridimensional Personality Questionnaire (Wang et al., 1997). Reduced glucocorticoid function could also lead to low reward dependence since glucocorticoids appear to potentiate the reinforcing effects of many rewards, including food, sex, and drugs of abuse (Piazza & Le Moal, 1997), and a general anhedonic state is one of the major symptoms of pathologies, such as Addison disease, in which hypocortisolism is observed (Reiser & Reiser, 1995). Moreover, several human studies point to the role of reduced glucocorticoid signal in conferring vulnerability to PTSD and depression. Individuals who demonstrate low cortisol levels after an acute trauma, such as motor vehicle accident, are at higher risk of subsequently developing PTSD symptoms (Delahanty, Raimonde, & Spoonster, 2000). Cortisol levels are also negatively correlated with a number of risk factors for depression, such as neuroticism, introversion, and perceptions of uncontrollability; in contrast, cortisol levels are positively correlated with self esteem, hardiness, and affective stability-factors which confer resistance to depression (McCleery & Goodwin, 2001; Pruessner, Hellhammer, & Kirschbaum, 1999; Scarpa & Luscher, 2002; Zorrilla, DeRubeis, & Redei, 1995).

In conclusion, congenitally helpless rats show behaviors consistent with a temperament of high novelty seeking, high behavioral inhibition, and low reward dependence, which most closely resembles the temperament observed in combat veterans with PTSD. However, just as PTSD patients are markedly inclined to depression and substance abuse, the temperament of helpless rats may best be viewed as conferring risk to stress-related psychopathology in general. Paradoxically, part of this vulnerability may lie in a predisposition to seek out the very kinds of stressful situations which are likely to provoke helplessness and depression.

Chapter 3: Metabolic Brain Mapping Methods

Functional neuroimaging technologies, such as positron emission tomography (PET) and functional magnetic resonance imaging (fMRI), have enabled the in vivo investigation of the functional neuroanatomy of depression. Given the current wealth of functional imaging data from depressed patients, analogous techniques used in animal models could provide an excellent bridge to the human disorders they are purported to simulate. In particular, PET measurements of regional glucose metabolism are directly comparable to the autoradiographic deoxyglucose method developed in animals. However, the use of metabolic mapping in animal models of depression has been rare, and this provides the major impetus for my dissertation work.

Metabolic mapping provides an opportunity to compare an animal model with a human disorder using a similar in vivo brain measure, which may speak more to a model's validity than any apparently shared behavior. Furthermore, since current human brain-imaging technology cannot accurately localize signals to specific subcortical nuclei, metabolic brain mapping of animal models provides a means for inferring functional activity in these undetectable regions of humans: if a model shares a pattern of cortical activity with a human disorder, one may speculate that the animal's subcortical pattern of activity would be shared as well. Finally, and perhaps most important, metabolic mapping provides a circuit-level framework for integrating the plethora of cellularmolecular data that animal research has generated.

QUANTITATIVE CYTOCHROME OXIDASE HISTOCHEMISTRY

Background

Cytochrome oxidase histochemistry, like 2-deoxyglucose (2-DG) and fluorodeoxyglucose (FDG) autoradiography (Gonzalez-Lima, 1992), provides a functional marker for energy metabolism, and therefore neuronal activity (Wong-Riley, 1989; Wong-Riley, Nie, Hevner, & Liu, 1998). As the final step in the electron transport chain, the catalytic activity of the mitochondrial enzyme cytochrome oxidase is critical for glucose oxidization and the production of adenosine triphosphate (ATP). Furthermore, as neuronal activity increases or decreases (and the need for ATP increases or decreases), the number of mitochondria and the amount of cytochrome oxidase within mitochondria adjust accordingly to match the activity-dependent need for ATP (Wong-Riley, 1989; Wong-Riley et al., 1998). Thus, cytochrome oxidase activity within a brain region is tightly coupled to the neuronal activity of that region.

While many PET studies have used deoxyglucose radiotracers, cytochrome oxidase has never been measured in depressed humans. The question arises then, if the goal is to provide a common brain measure between the helplessness model and human depression, why use cytochrome oxidase histochemistry instead of deoxyglucose autoradiography? To be sure, there is not a one-to-one correspondence between a region's deoxyglucose uptake and its cytochrome oxidase activity (Gonzalez-Lima & Cada, 1998b), but both provide an index of oxidative metabolism. In addition, cytochrome oxidase has several advantages for mapping changes in neurobiological predispositions. Namely, cytochrome oxidase is unique in that it marks cumulative, long-term neuronal activity, while FDG uptake reflects state-dependent activity spanning approximately 45 minutes following tracer injection (Gonzalez-Lima & Cada, 1998b). In contrast, cytochrome oxidase in naïve animals from different genetic strains reflects

stabilized baseline differences (Gonzalez-Lima & Sadile, 2000; Papa, Sadile, Sergeant, Shumake, & Gonzalez-Lima, 1998). Thus, with naïve rats from the congenitally helpless strain, deoxyglucose would mark state-dependent differences, while cytochrome oxidase would mark trait-dependent differences in brain metabolism. Certainly traits (inherited, stable characteristics) and states (dynamic brain conditions) are related, and one would expect to see both overlap and distinctions among cytochrome oxidase and deoxyglucose measures. However, our initial goal has been to examine the neurobiological predisposition to develop helpless behavior—not the brain's evoked response to stress. Thus, quantitative cytochrome oxidase histochemistry provides a unique way to isolate this innate component by identifying the brain regions affected in the congenitally helpless rats.

Method

Following decapitation, brains were removed intact and frozen rapidly in isopentane. Using a cryostat (Reichert-Jung) at -20°C, brains were sectioned at 40 μ m and kept frozen at -40°C until they were processed using quantitative cytochrome oxidase histochemistry, as described previously (Gonzalez-Lima & Cada, 1994; Gonzalez-Lima & Garrosa, 1991). In brief, the cytochrome oxidase staining procedure involves a series of chemical exposures, the first of which (0.1 M phosphate buffer with 10% wt/vol sucrose and 0.5% vol/vol glutaraldehyde, pH 7.6, for 5 min) facilitates tissue adherence to the slides. The next series of exposures (four changes of 0.1 M phosphate buffer with 10% wt/vol sucrose, for 5 min each) removes red blood cells. Then the slides undergo metal intensification (0.05 M Tris buffer, pH 7.6, with 275 mg/l cobalt chloride, 10% wt/vol sucrose, and 0.5% vol/vol dimethylsulfoxide, for 10 min) to enhance staining contrast and reduce time spent in the following incubation procedure (350 mg diaminobenzidine tetrahydrochloride, 52.5 mg cytochrome c, 35 g sucrose, 14 mg

catalase, and 1.75 ml dimethylsulfoxide in 700 ml of oxygen-saturated 0.1 M phosphate buffer, at 37 °C for 1 hour). The reaction is stopped by fixing the tissue in buffered formalin (for 30 min at room temperature with 10% wt/vol sucrose and 4% vol/vol formalin). Finally, the slides are dehydrated in a series of ethanol baths (increasing from 30% to 100% vol/vol ethanol), cleared with xylene, and coverslipped with Permount.

To quantify enzymatic activity and to control for staining variability across different batches of cytochrome oxidase staining, sets of tissue-homogenate standards were included with each batch of slides. This tissue homogenate was obtained from 12 normal adult Sprague-Dawley male rats, whose brains were removed after decapitation, stored at 4°C (in sodium phosphate buffer, pH 7.6), and then homogenized at 4°C. The enzymatic activity of cytochrome oxidase in this homogenate was assayed using spectrophotometry, as described by Gonzalez-Lima and Cada (1994), and activity units were defined at pH 7 and 37°C, where 1 unit oxidizes 1 μ mol of reduced cytochrome c per min (μ mol/min/g tissue wet weight). Remaining tissue homogenate was frozen and stored at -40°C.

Immediately prior to each cytochrome oxidase staining procedure, the standard homogenate was sectioned at varying thickness (10, 20, 40, 60, and 80 µm), and these sections were thaw-mounted onto a slide and stained for cytochrome oxidase along with the experimental tissue. Activity values determined from the spectrophotometric procedure were assigned to each standard, and these values were correlated with corresponding optical density measurements of the cytochrome oxidase chromatic indicator, taken with an image-processing system as described by Gonzalez-Lima and Cada (1994). The resulting linear regression equations ($r^2 > .90$) were used to convert optical density readings from brain regions of interest into cytochrome oxidase activity values, which were used in all statistical analyses in this study. Using an image-processing system (JAVA, Jandel Scientific, Corte Madera, CA), optical density was sampled from the regions of interest. The size of the square-shaped sampling window was adjusted for each region so that it was as large as possible while still allowing four, non-overlapping readings to be taken bilaterally. For each region, optical density was sampled across three adjacent sections and averaged. These optical density values were then converted to cytochrome oxidase activity units, which were determined by spectrophotometry of cytochrome oxidase standards as described above. In addition to the regions of interest, whole-brain metabolism was assessed by taking the mean of the average activity for each section of the brain.

ANALYZING METABOLIC MAPPING DATA

Background

Two competing assumptions govern how imaging data is used to infer the involvement of brain regions in a behavioral function. The first assumption presupposes a modular brain, in which specific functions are localized to specific regions. The alternative assumption holds that functions are distributed across networks of interacting brain regions. Most likely, the reality lies somewhere in between. That is, brain regions are neither modular, nor fully interconnected. Rather, they are selectively connected, or "semiconnected" (McIntosh, 2000), and there are mathematical demonstrations that such semiconnected systems maximize complexity and computational power (Tononi, Sporns, & Edelman, 1994). Thus, a semiconnected or modular brain integrates much more information much more quickly than a fully interconnected or modular brain would.

The statistical analysis of such complex systems is rife with challenge. By far the most popular approach is to simply ignore the complexity and apply a univariate linear model for each region of interest, followed by a correction for multiple comparisons.

However, brain imaging data clearly has multivariate features, since activity measured at different regions covaries, and the standard Bonferroni correction overestimates the number of independent tests and may lead to the rejection of many real differences (McIntosh, Bookstein, Haxby, & Grady, 1996). Alternatively, one could assume the other simple extreme—that the brain is fully interconnected—and apply a multivariate linear model or treat all regions as repeated measures of a unitary system. However, multivariate models require that the number of subjects exceed the number of variables. This may be the case if one is conducting a narrow sample of regions, but usually one is conducting a brain-wide exploratory analysis. And including all ROIs in a single omnibus repeated-measures ANOVA does not result in a further reduction of error variance, despite the gain in degrees of freedom (McIntosh & Gonzalez-Lima, 1994).

This dilemma led McIntosh & Gonzalez-Lima (1994a) to a strategy that is essentially a compromise between the two extremes: conduct separate repeated measures ANOVAs on small subsets of brain areas, grouped on the basis of functional connectivity. This approach may have greater theoretical validity than the alternatives mentioned so far since it explicitly reflects the balance of segregation and integration present in brain organization. Still, the delineation of functional groups is somewhat arbitrary. Even though brain regions are not fully interconnected and it is clear that unconnected regions should be assigned to separate groups, brain connectivity is still sufficiently extensive to defy strict bounds. No matter which parceling system one chooses, there are bound to be a large number of regions that bridge two or more anatomical groups. For example, regions comprising frontal cortex could form a functional group, or these regions could be segregated and grouped with the subcortical structures with which they share the most connections. Once again, one can choose to ignore this troublesome complexity and proceed with one's favorite grouping. Or, if one were extremely zealous, one could analyze all possible anatomical groupings. Even so, potentially important covariance information about regional interactions remains neglected. Because metabolic mapping makes it possible to obtain functional data from many regions in a single brain, one would ideally like to consider the interactions among brain regions when inferring which ones are important to a particular behavioral function. Perhaps by examining the covariance of brain activity between different brain areas, one can infer which areas are important nodes and how they are functionally connected.

Thus, regional means and network covariances are complementary ways of looking at functional neuroimaging data. Both are necessary for a thorough understanding of metabolic mapping results.

Method

The general strategy I have chosen for analyzing the data sets reported in the next three chapters is as follows: First, I divide the regions into small functional subgroups. I then perform repeated measures ANOVA on each subgroup, followed by simple-effects tests as necessary. I also perform a step-wise discriminant analysis on each subgroup of regions, as used by McIntosh and Gonzalez-Lima (1994a), to identify regions which might differentiate the experimental groups based not on mean differences, but on interregional covariance differences. I then assess a smaller number of regions for functional connectivity using pair-wise correlations. The regions entering into this analysis are limited to those showing significant mean differences from the ANOVAs and those identified by the discriminant analyses. If there are numerous regions, I limit the regions to the most influential ones, arbitrarily defined as the top 10 regions in terms of absolute mean difference, the top 10 regions in terms of univariate effect sizes, and the top 10 regions in terms of standardized discriminant coefficients.

Regional Mean Differences

Each group of regions was analyzed with a separate repeated measures ANOVA, followed by simple-effects tests adjusted by a Bonferroni correction. (Correcting for the smaller number of regions within each ANOVA is a compromise between correcting for all possible comparisons and not correcting at all.) An alpha level of .05 was used to establish statistical significance.

Estimates of Regional Influence

In order to infer the relative contribution of each region to the helpless phenotype, three different statistics were calculated. The first is simply the magnitude of the mean difference between groups, expressed as the activity difference between groups divided by the control group's activity. The second estimate is the η_p^2 statistic, which is the proportion of the effect + error sum-of-squares (SS) variance that is attributable to the effect:

$$\eta_{\rm p}^2 = SS_{\rm effect} / (SS_{\rm effect} + SS_{\rm error})$$

In addition to these indices of univariate effects, there might also be a pattern of interregional relationships, reflected by the between-group covariances, that differentiates the experimental groups. Because a discriminant function is a weighted linear combination of variables, a stepwise discriminant analysis can reveal regions which are influential in combination, but which may not show individual mean differences (McIntosh & Gonzalez-Lima, 1994). Discriminant analyses were conducted using a forward step-wise procedure with the minimization of Wilk's Lambda (a measure of residual variance) as the criteria for deriving the discriminant function. A criterion of p < .05 was used for entry of a brain area into the discriminant function. The total number of potential areas that could be entered into an equation was limited to the total sample size minus two (n - 2), so all regions could not be assessed simultaneously. Therefore, regions

were arbitrarily regrouped and reanalyzed. Only brain areas that were able to consistently distinguish between the experimental groups are reported.

Functional Connectivity

Functional connectivity was assessed by calculating the Pearson correlation coefficient for each pair of regions. To ensure the reliability of correlations and to protect against the effects of outliers, all correlations were subjected to a modified jackknife procedure. In this modified procedure, an individual is removed from the data set, and correlations are computed on the data set with *n*-1 subjects. Significance tests are then done on the correlation using *n*-1 subjects. The individual is then replaced back into the data set, and another individual is removed. Correlations and significance tests are again computed on the data set with *n*-1 subjects. This procedure is repeated until all individuals have been taken out once. Correlations are considered to be reliably significant from zero if they remain significant (p < 0.05) throughout each iteration.

Chapter 4: The Untreated Congenitally Helpless Adult

Evidence from human neuroimaging and post-mortem studies of patients with depression or PTSD, as well as other animal models of depression, was used to develop a list of candidate brain systems to be explored.

Method

Subjects

Subjects were 10 congenitally helpless and 10 congenitally non-helpless males, age-matched and weighing 250-300 g. The rats were housed at the University of Maryland under constant humidity ($50 \pm 10\%$) and temperature ($23 \pm 1^{\circ}$ C), on a 12 hour light/dark cycle, and with food and water available continuously. Following decapitation of the subjects, brains were removed intact, frozen rapidly in isopentane, and shipped frozen to the University of Texas (Austin, TX). These naïve rats were neither subjected to inescapable shock, nor any other experimental manipulations. The only independent variable differentiating the two groups was their ancestry. All animal protocols were approved by the institutional care and use committee (IACUC) at the University of Maryland School of Pharmacy, and conform to all NIH and USDA guidelines.

Tissue Processing

Tissue processing proceeded according to the protocol for quantitative cytochrome oxidase histochemistry outlined in Chapter 3.

Delineating Regions of Interest

In addition to the regions of interest discussed in the first section of this chapter, readings were taken from some regions sharing prominent anatomical connectivity. ROIs were located using Paxinos and Watson's (1997) and Gonzalez-Lima and Cada's (1998a) atlases of the rat brain.

Unfortunately, a generally accepted nomenclature and boundaries for the prefrontal cortical areas in rats is still lacking (Uylings, Groenewegen, & Kolb, 2003). To avoid confusion, names and abbreviations from different anatomists are matched in Table 4.1, and the boundaries and terms used in our study are outlined in Figure 4.1. I will follow the terminology of Uylings and van Eden (1990), except that I refer to ACd and Fr2 collectively as dorsomedial PFC, and to the ACv as simply anterior cingulate. Dorsomedial PFC, prelimbic, and medial orbital cortex were imaged at Bregma 3.7 mm. Although not regions of interest, other cortical regions were also imaged at this level, including lateral frontal, agranular insular, and lateral orbital cortex. Infralimbic cortex was imaged at Bregma 2.2 mm, and anterior cingulate was imaged at Bregma 0.7 mm. Because frontal-cingulate cortex shows consistently greater cytochrome oxidase staining in the superficial layers compared to the deep layers, these two divisions were analyzed separately.

For the basal ganglia, cytochrome oxidase readings were averaged across rostral (Bregma -0.8 mm) and caudal (Bregma -3.8 mm) samples of the caudate-putamen. The globus pallidus and ventral pallidum were assessed at Bregma -0.3 mm, and the nucleus accumbens was measured at Bregma 0.7 mm.

The lateral septal nucleus (LS) and medial septal/vertical diagonal band nucleus (MSDB) were imaged at Bregma +0.2 mm.

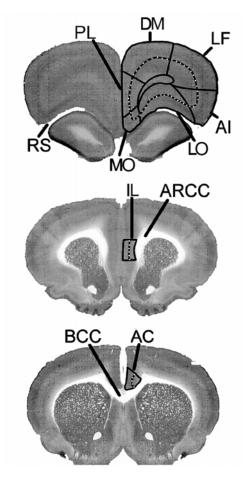
For the hippocampal complex, the dentate gyrus was imaged at Bregma -3.8 mm, and the subiculum was imaged at Bregma -5.8 mm. Fields CA1 and CA3 were assessed at both Bregma -3.8 and -5.8 mm.

 Table 4.1:
 Various terminologies of subdivisions of medial prefrontal cortex in the rat.

(Paxinos & Watson, 1998)	M2	Cg1	Cg2	PL	IL	MO
(Krettek & Price, 1977)	PrCm	ACd	ACv	PL	IL	MO
(Vogt & Peters, 1981)	4	24b	24a	32	25	14
(Zilles & Wree, 1985)	Fr2	Cg1	Cg2	Cg3	IL	MO
(Uylings & van Eden, 1990)	Fr2	ACd	AČv	ΡĹ	IL	MO

Note. M2 = secondary motor area; PrCm = medial precentral cortex; Fr2 = frontal cortex area 2; Cg1-Cg3 = cingulate cortex; ACd = dorsal anterior cingulate; ACv = ventral anterior cingulate; PL = prelimbic; IL = infralimbic)

Figure 4.1: Cytochrome oxidase-stained sections of frontal-cingulate regions of the rat brain. Segmented lines indicate division between superficial and deep layers of cortex. Abbreviations: AC, anterior cingulate; AI, agranular insular; ARCC, anterior radiation of the corpus callosum; BCC, body of the corpus callosum; DM, dorsomedial frontal; IL, infralimbic; LF, lateral frontal; LS, lateral orbital; MO, medial orbital; PL, prelimbic; RS, rhinal sulcus.



The basolateral and central amygdala were imaged at Bregma -2.3 mm. The anterior amygdala was imaged at Bregma -1.3 mm, and the medial amygdala was imaged at Bregma -2.3 mm. In addition, the bed nucleus of the stria terminalis (BNST), considered to be part of the "extended amygdala," was grouped with these structures and imaged at Bregma -0.3 mm.

The main hypothalamic area of interest was the paraventricular nucleus (PVH), which was imaged at Bregma -1.8 mm. In addition, various other hypothalamic areas implicated in the regulation of eating, sexual behavior, and circadian rhythm were imaged: the medial preoptic area (Bregma -0.3 mm), the lateral hypothalamic area and suprachiasmatic nucleus (Bregma -1.3 mm), and the ventral medial hypothalamic nucleus (Bregma -2.3 mm).

The following nuclei from the midline thalamic region were imaged: the reuniens (Bregma -1.8 mm), paraventricular, medial dorsal, and centromedial (Bregma -2.3 mm) nuclei. In addition, the anterior ventral nucleus (AV) was imaged at Bregma -1.8 mm.

The habenula (lateral and medial nuclei combined) was imaged at Bregma -3.8 mm. In addition, three brainstem regions which receive prominent projections from the habenula were imaged: the interpeduncular nucleus (IPN) and ventral tegmental area (VTA) at Bregma –6.3 mm, and the dorsal raphe (DR) at Bregma –8.0 mm.

RESULTS

Means Analysis

Means and standard errors for all regions of interest are reported in Tables 4.2 and 4.3.

Whole-Brain Metabolism

A univariate ANOVA showed no significant difference in whole-brain metabolism between groups, F(1,18) = .015, p = .91.

Frontal-Cingulate Cortex

Frontal and cingulate cortical regions were analyzed with a $2 \times 8 \times 2$ (Group × Region × Layers) repeated measures ANOVA. A significant three-way (Group × Region × Layers) interaction was obtained, F(7, 126) = 2.63, p = .014. Simple-effects tests of the interaction showed significant decreases in helpless rats in the superficial layers of dorsomedial PFC, F(1, 18) = 5.0, p = 0.038, medial orbital, F(1, 18) = 5.3, p = 0.034, and anterior cingulate, F(1, 18) = 5.3 p = 0.03, whereas a significant increase was found in the superficial layers of infralimbic cortex, F = 8.46, p = 0.009. No significant differences were observed in prelimbic cortex or in the inner layers of any region.

Basal Ganglia

Basal ganglia regions were analyzed with a $2 \times 2 \times 2$ (Group × Striatum-Pallidum × Dorsal-Ventral) repeated measures ANOVA, so that striatal regions (CP and Acb) could be contrasted with pallidal regions (GP and VP) and dorsal regions (CP and GP) could be contrasted with ventral regions (Acb and VP). However, only a significant main effect of group was obtained, F(1,18) = 24.9, p = 0.0001, indicating that metabolism throughout the basal ganglia was reduced in the helpless group (256 vs. 296 µmol/min/g).

Septal Region

The septal region was analyzed with a 2 × 2 (Group × Region) repeated measures ANOVA. Only a significant main effect of group was obtained, F(1,18) = 16.5, p = .001, indicating that metabolism was similarly reduced in both the LS and MSVDB of helpless rats (285 vs. 324 µmol/min/g).

Cortical Region	Helples	S S	Non-helpless		
	Superficial Layers	Deep Layers	Superficial Layers	Deep Layers	
Lateral frontal	298 ± 15	254 ± 11	323 ± 19	269 ± 18	
Dorsomedial PFC	$266 \pm 9*$	249 ± 10	315 ± 20	278 ± 14	
Prelimbic	202 ± 12	209 ± 10	229 ± 11	232 ± 13	
Medial orbital	212 ± 7*	197 ± 6	242 ± 11	224 ± 12	
Lateral orbital	248 ± 7	217 ± 8	277 ± 13	233 ± 13	
Anterior insular	257 ± 8	239 ± 7	284 ± 13	256 ± 15	
Infralimbic	$244 \pm 13*$	224 ± 10	198 ± 9	224 ± 11	
Anterior cingulate	$256 \pm 6*$	200 ± 5	291 ± 14	213 ± 9	

Table 4.2:	Mean \pm S.E. of baseline cytochrome oxidase activity (µmol/min/g tissue) for
	frontal-cingulate cortex in adult males. $*p < .05$.

Region	Helpless	Non-helpless
Basal ganglia		-
Caudate putamen (CP)	$219 \pm 9*$	261 ± 10
Nucleus accumbens (Acb)	313 ± 8*	364 ± 7
Ventral pallidum (VP)	$284 \pm 14*$	329 ± 3
Globus pallidus (GP)	$192 \pm 7*$	212 ± 5
Septal nuclei		
Lateral septum (LS)	$308 \pm 9*$	349 ± 8
Medial septum/Vertical diagonal band (MSVDB)	$248 \pm 10*$	290 ± 10
Hippocampal complex		
CA1 field, dorsal	259 ± 7	254 ± 10
CA1 field, ventral	310 ± 8*	257 ± 8
CA3 field, dorsal	254 ± 7	252 ± 7
CA3 field, ventral	321 ± 16*	278 ± 6
Dentate gyrus (DG)	345 ± 25	330 ± 10
Subiculum (Sub)	399 ± 14*	300 ± 18
Amygdaloid nuclei		
Bed nucleus stria terminalis (BNST)	$229 \pm 8*$	262 ± 8
Basolateral amygdala (BlA)	$359 \pm 9*$	420 ± 11
Central amygdala (CeA)	$333 \pm 9*$	398 ± 7
Anterior amygdala (AA)	202 ± 9	199 ± 12
Medial amygdala (MeA)	322 ± 15	341 ± 7
Hypothalamus		
Paraventricular nucleus (PVH)	$262 \pm 22*$	145 ± 6
Lateral nucleus (LH)	205 ± 8	206 ± 10
Medial preoptic area (MPA)	232 ± 20	250 ± 15
Suprachiasmatic nucleus (SCH)	232 ± 17	241 ± 11
Ventromedial nucleus (VMH)	245 ± 20	252 ± 29
Limbic thalamus		
Anteroventral nucleus (AV)	286 ± 32	210 ± 9
Centromedial nucleus (CM)	295 ± 23	254 ± 17
Medial dorsal nucleus (MD)	299 ± 20	266 ± 10
Paraventricular nucleus (Pv)	247 ± 26	237 ± 15
Reuniens nucleus (Reu)	251 ± 27	175 ± 11
Habenula and Brainstem Efferents		
Habenula	$338 \pm 46*$	198 ± 14
Interpeduncular nucleus (IPN)	$461 \pm 10^{*}$	368 ± 20
Ventral tegmental area (VTA)	$158 \pm 14*$	218 ± 16
Dorsal raphe (DR)	178 ± 4	194 ± 5

Table 4.3: Mean \pm S.E. of baseline cytochrome oxidase activity (µmol/min/g) for subcortical regions in adult males. *p < .05.

Hippocampus

The hippocampal region was analyzed with a $2 \times 2 \times 3$ (Group × Anterior-Posterior × Region) repeated measures ANOVA. A significant two-way (Group × Anterior-Posterior) interaction was obtained, F(1, 18) = 9.27, p = .007. Simple-effects tests of the interaction showed that mean cytochrome oxidase activity in the congenitally helpless strain was significantly increased throughout the posterior hippocampus, F(1, 18) = 26.8, p = 0.00006, but not in the anterior hippocampus, F(1, 18) = .442, p = .52.

Amygdala

The amygdala region was analyzed with a 2 × 5 (Group × Region) repeated measures ANOVA. A significant two-way (Group × Region) interaction was obtained, F(4, 72) = 4.61, p = .002. Simple-effects tests of the interaction showed that mean cytochrome oxidase activity in the congenitally helpless strain was significantly reduced in the BNST, basolateral, and central amygdala, Fs(1,18) = 8.42, 18.3, 30.9, ps = .01, 0.0005, 0.00003, respectively; but not in the medial or anterior amygdala, Fs(1, 18) = 1.4, .086, ps = .25, .77, respectively.

Hypothalamus

Hypothalamic nuclei were analyzed with a 2 × 6 (Group × Region) repeated measures ANOVA. A significant two-way (Group × Region) interaction was obtained, F(4, 72) = 4.61, p = .002. Simple-effects tests of the interaction showed that mean cytochrome oxidase activity in the congenitally helpless strain was significantly increased in the PVH, F(1, 18) = 28.2, p = .0.00005. No other regions were significantly different.

Midline and Anterior Dorsal Thalamic Regions

Thalamic nuclei were analyzed with a 2 × 5 (Group × Region) repeated measures ANOVA. No significant effects were obtained, either for the group main effect, F(1, 18) = 2.48, p = .14, or for the interaction (Group × Region), F(1, 18) = 1.46, p = .24.

Habenula and Efferent Brainstem Nuclei

The habenula and its efferent brainstem targets were analyzed with a 2 × 4 (Group × Region) repeated measures ANOVA. A significant two-way (Group × Region) interaction was obtained, F(3, 54) = 10.2, p = 0.00002. Simple-effects tests of the interaction showed that mean cytochrome oxidase activity in the congenitally helpless strain was significantly increased in the habenula and IPN, Fs(1, 18) = 9.31 and 18.0, ps = .007 and .0005, respectively; and significantly decreased in the VTA, F(1, 18) = 8.33, p = .01. The dorsal raphe was not significantly different between groups, F(1, 18) = 2.74, p = .12.

Estimates of Regional Influence

Estimates of regional influence are given in Tables 4.4 and 4.5. Percent mean differences and partial-eta-squared (η_p^2) estimates give very different assessments for some regions. For example, the habenula and VTA are ranked high in terms of difference magnitude but low in terms of η_p^2 because the larger group difference for these regions was accompanied by a disproportionate increase in error, or unexplained variance. Vice versa, the basal ganglia and amygdala regions showed relatively small mean differences but very large η_p^2 because group membership accounted for a far greater portion of their variance. For other regions, the two statistics are more consistent. For example, both suggest that the PVH, IPN, and ventral hippocampus are among the most influential regions, whereas frontal-cingulate cortex and BNST are among the least.

Table 4.4:	Top 10 regions in terms of effect size, ranked according to relative
	magnitude of difference (left) or partial eta-squared (right).

Region	% Dif.	Region	${oldsymbol{\eta}_{\mathrm{p}}}^2$
1. PVH	.80	1. Central amygdala	.63
2. Habenula	.68	2. PVH	.62
3. VTA	28	3. Ventral hippocampus	.60
4. IPN	.25	4. Basal ganglia	.58
5. Ventral hippocampus	.20	5. Basolateral amygdala	.50
6. ACd	19	5. IPN	.50
7. Central amygdala	16	7. Septal nuclei	.48
8. Basolateral amygdala	15	8. ACd	.36
8. Infralimbic cortex	.15	9. Habenula	.34
10. BNST	14	9. Infralimbic cortex	.34

Table 4.5:Standardized discriminant coefficients for regions selected by a stepwise
discriminant analysis.

Region	Standardized Discriminant Coefficients
1. Ventral CA1	.89
2. VTA	82
3. Central amygdala	74
4. Habenula	.57

In addition to these univariate estimates of effect size, a multivariate stepwise discriminant analysis selected four regions on the basis of their ability to discriminate significantly between the two groups: the CA1 field of the ventral hippocampus, the ventral tegmental area, the habenula, and the central amygdala. Note that all of these regions also showed significant mean differences.

In summary, the PVH and habenula showed the largest mean differences between groups. The central amygdala and PVH showed the largest effect sizes, and ventral CA1 and the VTA showed the largest discriminant coefficients. These appear to be the regions most affected by the selective breeding for helplessness susceptibility.

Analysis of Functional Connectivity

Pearson product-moment correlations were performed for the regions showing the ten largest mean differences and ten largest effects sizes. The basal ganglia regions (CP, Acb, GP, VP) were considered as a collective, as were the septal nuclei (MS and LS) and posterior hippocampal regions (CA1, CA3, and subiculum). This is because, in addition to showing similar mean differences, regions within each collective were highly correlated with each other in both helpless and non- helpless groups (data not shown). However, no other significant correlations were found in either group (p > .05 following jackknife, data not shown).

DISCUSSION

Using cytochrome oxidase as a marker for metabolic capacity, the results indicate that congenitally helpless rats, as compared to congenitally non-helpless rats, show reduced metabolism in dorsomedial prefrontal cortex, infralimbic cortex, basal ganglia, septal nuclei, basolateral and central amygdala, and ventral tegmental area. In contrast, elevated metabolism was found in the paraventricular hypothalamus, habenula, interpeduncular nucleus, and ventral hippocampus. In interpreting these findings, it is important to keep in mind the following issues. First, it should be emphasized that the assayed animals never experienced the learned helplessness training paradigm, nor any other stressful challenges. Therefore, the results reported here reflect endogenous differences in brain metabolism. While these differences likely influence the neurobehavioral response to stress, they should not be interpreted as reflecting the stress response itself.

A second issue concerns the fact that this is a comparison between two genetically altered strains, not a comparison between an altered strain and a randomly bred strain. Although biochemical assays have shown congenitally non-helpless rats to be similar to randomly bred Sprague-Dawley rats (Henn & Edwards, 1994), the possibility exists that any given cytochrome oxidase difference between strains may be due as much to deviation from normal of the non-helpless rats as the helpless rats. However, we have cytochrome oxidase data available from normal control rats from other studies in this lab (Cada, Gonzalez-Lima, Rose, & Bennett, 1995; Gonzalez-Lima & Cada, 1998b), and they are similar to values in non-helpless rats, with the exception of the PVH and habenula. In these regions, normal rats appear to have activity intermediate to that of the congenitally helpless and non-helpless rats. Thus, a mutual divergence in activity may account for the unusually large groups differences observed in the PVH and habenula. Still, the absence of a random control group does not permit a conclusive answer to the question of which strain is driving which regional difference, and it is best to interpret the findings in light of a resistance-vulnerability continuum. That is, from the perspective of the congenitally helpless rats, strain differences reflect increased vulnerability to helplessness (even if the increased vulnerability is technically due to an absence of resistance).

Prefrontal Cortex (PFC)

Congenitally helpless rats showed changes in frontal-cingulate regions comparable to those that have demonstrated differences in human depression, and the direction of these changes are the same as those seen in human studies. Located ventral to the genu of the corpus callosum, subgenual anterior cingulate has shown both metabolic decrements (Drevets et al., 1997) and mild hyperactivity (Mayberg et al., 1999) in depression. This apparent discrepancy may be resolved by the finding that depressed patients show a marked reduction in glial cells within the subgenual region (Ongur, Drevets, & Price, 1998), giving a false impression of hypometabolism when measured with low-resolution PET. Computer simulations that correct PET data for this volume effect conclude that metabolic activity in the remaining subgenual tissue is actually increased in these patients (Drevets, 2000). Moreover, the subgenual cingulate was the only region that showed a correlation between activity and overall depressive symptoms in both unipolar and bipolar patients (Dunn et al., 2002), and increased subgenual metabolism is a well-replicated correlate of normal human sadness (Liotti et al., 2000). Infralimbic cortex, the homologous region in the rat, was likewise hypermetabolic in the congenitally helpless rat, indicating that excessive activity in this region may be a susceptibility trait that exists prior to the onset of depression.

Subgenual/infralimbic cortex appears to be important for modifying the activity of the autonomic nervous system (Bacon & Smith, 1993; Frysztak & Neafsey, 1991). Humans with lesions that include the subgenual cingulate show abnormal autonomic responses to emotionally provocative stimuli and are unable to experience emotion related to concepts that ordinarily evoke emotion (Damasio, Tranel, & Damasio, 1990). Likewise, rats with infralimbic lesions show significantly reduced freezing and little or no ultrasonic vocalizations when presented with a conditioned stimulus predicting shock (Frysztak & Neafsey, 1991). If hyperactivity of this region reflects the converse of the lesion effects, then increased subgenual/infralimbic metabolism may confer increased emotionality.

Other PFC regions where dysfunction may impair the ability to modulate emotional responses in mood disorders include the dorsomedial PFC (area 9), dorsolateral PFC (areas 9/46), and the area of anterior cingulate immediately dorsal to the corpus callosum (area 24). Abnormally decreased blood flow and metabolism in these areas have been the most extensively replicated findings in depression neuroimaging (Baxter, Jr. et al., 1989a; Bench et al., 1992; Biver et al., 1994; Ebert et al., 1991; Mayberg et al., 1997), and these metabolic deficits appear to relate to the negative thoughts (Dunn et al., 2002) and subtle impairments of attention and memory (Bench et al., 1993) that accompany major depression episodes. The congenitally helpless rats also showed metabolic suppression in homologous dorsomedial PFC. Since helpless rats also show learning deficits related to impaired attention (Lee & Maier, 1988; Minor et al., 1984), diminished functioning of their dorsomedial PFC may likewise be to blame.

The pattern of reduced metabolism in dorsomedial frontal and elevated metabolism in infralimbic cortex generally agrees with the human literature. It is also consistent with a circuit model put forth by Helen Mayberg (1997), who suggests there is a ventral-dorsal, limbic-cortical opposition in effect in depression, as well as normal sadness. Her hypothesis is based on a series of neuroimaging studies showing that cognitive tasks activate the dorsal PFC and anterior cingulate while suppressing more ventral, limbic structures, and that, vice versa, emotional tasks activate more ventral limbic structures while suppressing dorsal regions (Mayberg, 1997). Interestingly, similar reciprocal relationships in brain metabolism appear to exist in rodents. Mice that have learned to suppress an emotional response show increased metabolism in dorsomedial

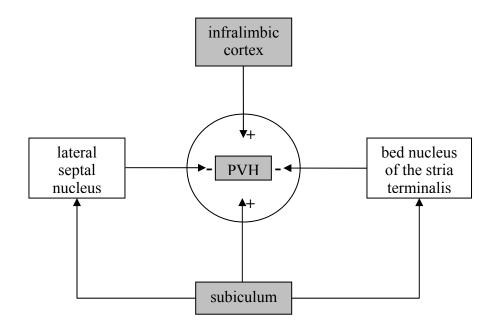
PFC which is strongly correlated with decreased metabolism in multiple subcortical areas related to emotional expression (Barrett, Shumake, Jones, & Gonzalez-Lima, 2003). The implication is that depressed humans and helpless rats may experience a brain state in which negative emotions interfere with new learning and new learning is unable to change negative emotions.

Regions Regulating the Hypothalamic-Pituitary-Adrenal (HPA) Axis

The PVH of the congenitally helpless rat showed a large elevation of cytochrome oxidase activity, more than five times the difference observed in frontal and cingulate cortex. Indeed, it is the largest cytochrome oxidase difference we have ever detected in any study. (Originally, magnocellular and parvocellular divisions of the PVH were imaged separately, but, because both showed the same magnitude of difference, the PVH data is reported and discussed as a single region.) Since stress activates the PVH (Palkovits, 2000) and since chronic stress can lead to depression, one might expect the PVH to become hyperactive in stress-induced depression. What is interesting about the congenitally helpless rats is that they show PVH hypermetabolism in the absence of any explicit stress, indicating that PVH hyperactivity can also be triggered genetically. This finding suggests a novel theoretical mechanism linking the contributions of heredity and stress in the etiology of depression: if sustained hyperactivity of the PVH leads to depression, then individuals with higher baseline levels of PVH activity would simply be closer to this threshold.

Although the CRH-producing neurons of the PVH receive few direct forebrain afferents, several forebrain regions influence PVH activity via local interneurons scattered in the area immediately surrounding the PVH. These include glutamate projections from the ventral subiculum and infralimbic cortex and GABA projections from the lateral septal nucleus (LS), bed nucleus of the stria terminalis (BNST), medial amygdaloid nucleus, paraventricular thalamic nucleus, suprachiasmatic nucleus, and ventromedial hypothalamic nucleus (Canteras & Swanson, 1992; Moga, Weis, & Moore, 1995; Risold & Swanson, 1997; Watts, Swanson, & Sanchez-Watts, 1987). Of these, infralimbic cortex and the ventral subiculum showed metabolic increases in the congenitally helpless rat, and the LS and BNST showed metabolic decreases. This metabolic pattern complements the transmitter pattern: the structures sending excitatory projections to the PVH are overactive, and the structures sending inhibitory projections are underactive (Fig. 4.2). However, the intervening neurons in the PVH-surround are both GABAergic and glutamatergic, so incoming inhibitory and excitatory signals could potentially be reversed by the GABA interneurons. Thus, increased activity of infralimbic cortex and hippocampus coupled with decreased activity of LS and BNST could either be a cause of PVH hyperactivity or an adaptive attempt to constrain a hyperactive PVH.

Even though congenitally helpless rats were not subjected to shock or any other experimental stress, their dissociative septo-hippocampal pattern fits with data obtained from learned helpless rats that have undergone inescapable shock. First, fornix transection completely blocks the development of learned helplessness (Leshner & Segal, 1979), suggesting that hippocampal output facilitates helpless behavior. Moreover, the hippocampi of learned helpless rats show increased glutamate activity and decreased GABA activity (Petty & Sherman, 1981). This is consistent with our finding of hippocampal hypermetabolism, which may be related to functional changes in the processing of corticosterone feedback; chronic antagonism of hippocampal glucocorticoid receptors increases vulnerability to helplessness (Papolos et al., 1993), and these receptors appear dysfunctional in congenitally helpless rats (Lachman et al., 1993). Hippocampal hypermetabolism is consistent with some human neuroimaging data, which has shown elevated hippocampal metabolism in PTSD patients with low cortisol levels Figure 4.2: Regions projecting to interneurons surrounding the PVH. Shaded regions are hypermetabolic, and unshaded regions are hypometabolic. + = excitatory glutamate influence; - = inhibitory GABA influence.



(Bonne et al., 2003) and reduced hippocampal metabolism following six weeks of antidepressant treatment (Mayberg et al., 2000).

Our finding of reduced septal metabolism also fits with data from learned helplessness experiments. Reduced c-fos activity was observed in the lateral septum of rats that became helpless following inescapable shock (Steciuk et al., 1999). Interestingly, the same study found that resistant rats, which received the same inescapable shock but did not become helpless, showed no change in septal activity. This indicates that the septal effect was due, at least in part, to phenotypic differences, perhaps similar to those expressed by the congenitally helpless rats. It also raises the possibility that a baseline septal difference may exist prior to stress exposure in rats that go on to become learned helpless.

Reduced septal metabolism may also reflect an impairment in reward motivation. The medial and lateral septal nuclei, the nucleus of the diagonal band, and the BNST all support electrical self-stimulation (Cazala, Galey, & Durkin, 1988; Lithgow & Barr, 1984). Furthermore, stimulation of the medial forebrain bundle increases cytochrome oxidase activity in the LS (Bielajew, 1991), lateral hypothalamic self-stimulation increases c-fos activity in the BNST (Arvanitogiannis, Flores, Pfaus, & Shizgal, 1996), and stimulation of the ventral tegmental area increases glucose uptake in both the LS and BNST (Esposito et al., 1984). In addition, several different antidepressant treatments including drugs, sleep deprivation, and electroshock—were all found to increase neuronal firing rates in the LS (Contreras, Alcala-Herrera, & Marvan, 1989).

Evidence also suggests that the LS and BNST play a role in regulating the PVH, which receives significant projections from both regions (Silverman, Hoffman, & Zimmerman, 1981). Since these projections are thought to be predominantly GABAergic and inhibitory (Swanson & Risold, 2000), decreased activity in the LS and BNST could

disinhibit PVH activity, contributing to its hyperactivity in congenitally helpless rats. This idea is further supported by evidence showing that adrenalectomy reduces BNST input to the PVH, particularly to the corticotropin-releasing hormone neurons (Mulders, Meek, Hafmans, & Cools, 1997), and that the hippocampus may mediate adrenal feedback via its projections to the LS and BNST (Cullinan, Herman, & Watson, 1993). In conclusion, the dissociation between hippocampal and septal activity in congenitally helpless rats may reflect a disruption in negative feedback to the PVH, which may be a key predisposing factor for the development of helpless behavior.

Habenula, VTA, and Related Regions

The results suggest that the habenula (reported and discussed as a single region because both medial and lateral nuclei showed equivalent differences) may be an especially important region affected in helpless rats, yet it has been overlooked by many depression researchers, in spite of its major role in regulating monoamine transmission (Christoph, Leonzio, & Wilcox, 1986; Ho et al., 1996; Lisoprawski, Herve, Blanc, Glowinski, & Tassin, 1980; Matsuda & Fujimura, 1992; Nishikawa, Fage, & Scatton, 1986; Speciale, Neckers, & Wyatt, 1980; Wang & Aghajanian, 1977). In fact, animal studies have implicated the habenula in a variety of processes disrupted in depression, including the sleep-wake cycle (Haun, Eckenrode, & Murray, 1992; Valjakka et al., 1998), reward mechanisms (Boyd & Celso, 1970; Gallistel, Gomita, Yadin, & Campbell, 1985; Sutherland & Nakajima, 1981), antinociception (Benabid & Jeaugey, 1989; Fuchs & Cox, 1993), and behavioral inhibition (Lee & Huang, 1988). In addition, numerous reports have linked the habenula with mating behavior, avoidance learning, maze learning, feeding behavior, and hormonal response to stress (Sandyk, 1991; Sutherland, 1982). Three other models of depression-amphetamine withdrawal, chronic stress, and α -methyl-para-tyrosine challenge—were shown to increase habenula metabolism

(Caldecott-Hazard, Mazziotta, & Phelps, 1988), and it appears that selective breeding for helplessness has produced the same result. At least one human neuroimaging study has found a strong correlation between habenula activity and depression severity (Morris et al., 1999). Perhaps most striking, though, is the finding that habenula ablation completely blocks the development of learned helplessness (Amat et al., 2001).

The IPN, which receives a major projection from the medial habenula (Contestabile & Flumerfelt, 1981), may also play an important role in helplessness susceptibility. Like the habenula, it is metabolically hyperactive in congenitally helpless rats, and input from the habenula likely contributes to IPN hypermetabolism. In addition, the IPN receives more acetylcholine input than any other region in the mammalian brain (Woolf & Butcher, 1985). This is interesting in light of evidence implicating excessive cholinergic tone in the etiology of depression (Charles et al., 1994; Dilsaver & Coffman, 1989; Dube, 1993; Janowsky, el Yousef, Davis, & Sekerke, 1972; Janowsky, Risch, & Gillin, 1983; Steingard et al., 2000). Cholinergic agonists increase the drive for REM sleep, and excessive REM sleep is a reliable marker of endogenous depression (Riemann, Berger, & Voderholzer, 2001). Furthermore, the FSL rat, which was selectively bred for sensitivity to cholinergic agonists, shows many features of depression, including the characteristic REM sleep abnormality (Yadid et al., 2000), which may be related to IPN function. In particular, cholinergic innervation of the IPN is necessary for maintaining REM sleep episodes (Haun et al., 1992), and deafferentation of the IPN reduces time spent in REM sleep by 79%, without affecting non-REM sleep (Valjakka et al., 1998). A reasonable prediction is that the converse-cholinergic stimulation of the IPN-would increase the drive toward REM sleep. Thus, IPN hypermetabolism might reflect a localized increase of cholinergic activity and a corresponding alteration in REM sleep.

Assessing electroencephalographic parameters of the sleep cycle in congenitally helpless rats remains an important experiment yet to be done.

The finding of habenula hypermetabolism is also consistent with the finding of diminished VTA activity, which may reflect defective dopamine innervation. Numerous studies have shown that habenula metabolism is sensitive to changes in dopamine transmission and that the relationship is consistently an inverse one. Dopamine agonists and stimulation of the medial forebrain bundle strongly inhibit metabolism in the habenula (Trugman, James, & Wooten, 1991; Weissman, Marquis, Moreton, & London, 1989; Wilkerson & London, 1989), while dopamine antagonists strongly promote it (Gomita & Gallistel, 1982; Pizzolato, Soncrant, & Rapoport, 1984). Interestingly, a recent human fMRI study revealed analogous habenula responses to positive and negative feedback in a motion prediction test; blood flow to the habenula decreased following successful trials and increased following failures (Ullsperger & Von Cramon, 2003). The authors relate this finding to animal studies showing increased dopamine activity following reward and decreased dopamine activity following omission of expected reward (Schultz, 2000; Schultz & Dickinson, 2000; Schultz, 2002), and they speculate that the habenula may inhibit dopamine activity in response to failure. However, the causality between VTA activity and habenula activity could be reciprocal. The VTA and nucleus accumbens have been shown to project to the habenula (Felton, Linton, Rosenblatt, & Morell, 1999), and these projections may be responsible for dopamine-mediated changes in habenula activity. However, the habenula also sends projections to the VTA and can indeed inhibit the firing of dopamine neurons (Christoph et al., 1986; Lisoprawski et al., 1980; Matsuda & Fujimura, 1992; Nishikawa et al., 1986).

The major targets of VTA projections also showed metabolic differences in this study, with almost equivalent metabolic decreases in the striatum and basal and central regions of the amygdala. Moreover, destruction of VTA dopamine neurons leads to an identical pattern of activity as that found here: increased metabolism in the habenula and reduced metabolism in the striatum and amygdala (Kozlowski & Marshall, 1980). Decreased metabolism in nucleus accumbens, caudate-putamen, ventral pallidum, and globus pallidus is consistent with evidence implicating basal ganglia dysfunction in depression. There appears to be a connection between major depression and other neurological disorders affecting the basal ganglia (Rogers et al., 1998). For example, symptoms of major depression often accompany loss of forebrain dopamine in Parkinson's disease (Mayeux, Stern, Rosen, & Leventhal, 1981; Stern & Langston, 1985), and depression frequently precedes the onset of the cardinal motor signs of Parkinson's disease (Austin & Mitchell, 1996). Many human neuroimaging studies have also shown reduced blood flow or metabolism in the basal ganglia of depressed patients (Videbech, 2000).

The finding of decreased basolateral and central amygdala metabolism is also consistent with reduced VTA activity. Although we anticipated the amygdala would be hypermetabolic based on human neuroimaging studies, these findings appear to be restricted to a subtype of depression known as "familial pure depressive disease" and also non-psychotic bipolar depression (Drevets, 2000). However, in patients meeting the criteria for major depressive disorder, right amygdala metabolism predicted negative affect, but neither left nor right amygdala metabolism predicted depression severity or differentiated depressives from controls (Abercrombie et al., 1998). In addition, decreased metabolism has been observed in anterior paralimbic areas of depressed patients, around and encompassing some of the amygdala (Kimbrell et al., 2002).

Although the role of the amygdala in fear and anxiety has dominated the literature, there is also a substantial literature linking the amygdala with reward-related processing (Everitt et al., 1999). Indeed, in rats, rewarding intracranial self-stimulation and administration of dopamine agonists increase glucose metabolism in the amygdala (Porrino et al., 1984), while aversive, arousing stimulation of the midbrain reticular formation decreases amygdala metabolism (Gonzalez-Lima & Scheich, 1985). Likewise, human studies have shown that amphetamine and cocaine administration increase blood flow to the amygdala (Devous, Sr., Trivedi, & Rush, 2001; Breiter & Rosen, 1999), and cocaine withdrawal, which mimics many symptoms of major depression, was accompanied by amygdala hypometabolism in rats (Hammer, Jr., Pires, Markou, & Koob, 1993). The projection fields of the VTA are also consistent with the hypothesis that amygdala hypometabolism may reflect reduced dopamine innervation in the helpless rats: the VTA sends substantial projections to the basal and central amygdala, which showed significant differences, but not to the anterior and medial amygdala, which did not (Fallon & Ciofi, 1992). Furthermore, depressed humans show decreased expression of dopamine transporters in the basal and central amygdala and increased expression of D2/D3 receptors in the basal, lateral, and central amygdala, consistent with a loss of dopamine innervation (Klimek, Schenck, Han, Stockmeier, & Ordway, 2002).

The habenula is also poised to play a major role in regulating serotonin neurons of the DRN since habenula afferents account for the largest excitatory amino acid input to the DRN (Kalen, Karlson, & Wiklund, 1985; Kalen, Pritzel, Nieoullon, & Wiklund, 1986). However, the effects of this input are complicated by the fact that the habenula excites both serotonin neurons and local GABA neurons, which in turn inhibit the serotonin neurons (Ferraro, Montalbano, Sardo, & LaGrutta, 1997). The net effect seems to be an excitation of serotonin neurons with low-frequency habenula stimulation, but an inhibition with high-frequency stimulation (Ferraro et al., 1997). While dorsal raphe activation appears to be necessary for the development of learned helplessness, its baseline activity appears relatively normal in congenital helplessness. It is still possible that the amygdala, DRN, or other regions could become activated if the rats were exposed to inescapable shock or more severe stress.

Conclusion

The current findings suggest that the genetic predisposition to helpless behavior is more linked to the habenula and VTA, which showed the second largest increase and decrease, respectively, and which were identified by a stepwise discriminant analysis as reliably distinguishing congenitally helpless rats from non-helpless rats. The PVH, central amygdala, and ventral hippocampus also appear to be important. These regions appear to be critical nodes in a broader circuit underlying helplessness susceptibility and, as such, may be particularly effective targets for therapeutic intervention.

Chapter 5: The Congenitally Helpless Newborn

In the previous chapter, congenitally helpless rats were found to possess activity patterns in prefrontal (PFC) and cingulate cortex similar to those detected in human imaging studies of depression. The paraventricular hypothalamic nuclei (PVH) showed an 80% increase in metabolism, and the septal-hippocampal regions regulating the PVH appeared dissociated, with hypermetabolic hippocampal regions and hypometabolic septal regions. The habenula and interpeduncular nucleus (IPN) also appeared hyperactive, while metabolism was decreased in the ventral tegmental area (VTA), basal ganglia, and amygdala.

By using the metabolic marker cytochrome oxidase, which reflects brain activity averaged over a long period of time (Wong-Riley, 1989), we reasoned that any phasic stress an animal might experience in day-to-day life (e.g., moving to a new cage) would have minimal impact on cytochrome oxidase levels. Thus, innate pathology could theoretically be viewed apart from stress pathology. However, this is likely an overidealized scenario for several reasons. First, research has shown that quality of maternal care affects the neural circuitry governing stress reactivity, and maternal behavior itself appears to be learned (Meaney, 2001). It is not known whether congenitally helpless mothers are deficient in maternal care, but, if so, this could further contribute to the pattern of brain activity seen in adult helpless animals. Second, what we might regard as mild, phasic stress might not be perceived as such by an animal that becomes helpless after a single electric shock. If the helpless phenotype involves high reactivity to stress, then even "mild" stress such as cage changing may lead to profound brain changes. Finally, even if postnatal stress were not a factor, many brain differences seen in congenitally helpless adults may reflect compensatory developmental processes; a few regions showing pathology may, over time, confer abnormal activity to many interconnected structures.

We attempted to address these issues by examining congenitally helpless brains collected at birth. The neonatal brain provides an opportunity to examine pathology before functional connections are well established, and this may shed light on which brain regions are at the core of the activity pattern in adults. In addition, examining newborn brains is a simple way to avoid the potential confound of postnatal stress and establish which brain changes, if any, are purely congenital—that is, determined by genetics and/or prenatal environment.

METHODS

Subjects

Subjects were congenitally helpless (n = 16) and non-helpless (n = 20) newborn rats bred in our laboratory from 4 breeding pairs (2 of each strain) obtained from the Central Institute for Mental Health (Mannheim, Germany), courtesy of Prof. Fritz Henn. In order to avoid excessive inbreeding, first and second order relatives were excluded from mating. Breeding pairs were composed of first or second cousins.

When not mating, rats were individually housed in polycarbonate cages $(45 \times 24 \times 21 \text{ cm})$ in a temperature-controlled room $(22 \pm 1^{\circ}\text{C})$. They were maintained on a 12/12 hour light cycle and provided continuous food and water. Males were introduced into the female cages daily at the beginning of the dark cycle, and sexual behavior was monitored for 1 hour. If a female proved receptive, the pair was left together until the beginning of the next dark cycle when they were separated. Otherwise, males were returned to their home cages at the close of the hour.

Females were weighed three times a week in order to monitor pregnancy progress, and they were moved to clean cages twice per week. Otherwise, they were left undisturbed during the pregnancy period. With the mating period considered Day 0, females rats who gave birth did so invariably during the light cycle on Day 22. On the expected delivery date, the pregnant rat was checked once per hour, and pup brains were harvested within one hour of birth. Sex of pup was determined by measuring anogenital distance. There were 7 male and 9 female helpless pups from 2 litters (2 males and 1 female from one breeding pair, 5 males and 8 females from the other), and 10 male and 10 female non-helpless pups from 3 litters (3 males and 5 females from one breeding pair and 2 males and 5 females from the first litter and 5 males culled from a second litter of 7 males and 6 females from the other breeding pair). These methods were approved by the Institutional Animal Care and Use Committee.

Tissue Processing

Following decapitation, brains were removed intact and frozen rapidly in isopentane. Using a cryostat (Reichert-Jung) at -20°C, brains were sectioned at 40 μ m and kept frozen at -40°C until they were processed using quantitative cytochrome oxidase histochemistry, as described in Chapter 3.

Delineating Regions of Interest

Regions of interest were limited to regions showing significant effects in the adult. The Paxinos, Törk, Tecott, and Valentino (1991) Atlas of the Developing Rat Brain was used to delineate regions of the newborn brain. In the adult helpless brain, significant differences in cortical regions were limited to the outer layers. Therefore, we divided the cortical regions of the newborn into equal outer and inner halves, which were sampled separately.

Dorsomedial PFC was sampled across the dorsomedial quadrant of atlas level P0-3, which combined the medial half of frontal cortex and the dorsal half of cingulate cortex. Medial orbital (MO) cortex was defined as the medial half of the orbital region, which was sampled at the same level.

Infralimbic cortex (IL) was sampled at atlas level P0-5, adjacent to the portion of the forceps minor corpus callosum where the medial side runs roughly parallel to the midline. Readings were taken from the dorsal most part of this region, which should correspond to the IL. However, the boundary with dorsal peduncular cortex was extremely close and undefined, so it is possible that some readings overlapped with this more ventral region.

The anterior cingulate (AC) was sampled at atlas level P0-10, where it is bounded by white matter and an imaginary perpendicular line bisecting the cingulum. Also imaged at this level were the nucleus accumbens core and shell, sampled just lateral and medial, respectively, to the anterior commissure.

The septal area was imaged at atlas level P0-11. Here, the lateral septal nucleus (LS) is bounded dorsally by the corpus callosum and laterally and ventrally by the septal neuroepithelium (spt). Readings were taken adjacent to these structures to insure no overlap with the medial septal nucleus (MS). The dorsal limit of the MS was defined by an imaginary horizontal line through the most lateral point of the LS, and the ventral limit was defined by an imaginary horizontal line through the most ventral point of the spt. MS readings were taken adjacent to the midline to insure no overlap with the LS. Vertical diagonal band (DB) readings were also taken adjacent to the midline, immediately ventral to the MS, and extending to the base of the brain.

Basal ganglia regions and the bed nucleus of the stria terminalis (BNST) were sampled at atlas level P0-16, which provides the best anatomical landmarks for delineating these regions. The BNST was sampled within an area delimited dorsally by the caudate putamen neuroepithelium, medially by the anterior commissure, laterally by the internal capsule epithelium (ic), and ventrally by an imaginary horizontal line extending through the ventral most point of the ic. This same horizontal line was also used to roughly divide the globus pallidus (GP) and ventral pallidum (VP), with readings taken lateral and ventral, respectively, to the ic. The boundaries of the caudate putamen (CPu) were visually obvious.

The basolateral (BLA) and central (CeA) amygdaloid nuclei were imaged at atlas level P0-23, where the external capsule (ec) and stria terminalis (st) could serve as landmarks, with readings for the BLA taken immediately ventromedial to the ec and readings for the CeA taken immediately dorsolateral to the st. The PVH was also imaged at this level. This nucleus was well delineated by the cytochrome oxidase stain although magnocellular and parvocellular divisions could not be discriminated. The medial (MHb) and lateral (LHb) habenula, imaged at atlas level P0-29, were also readily discerned from the cytochrome oxidase stain itself.

Hippocampal regions (CA1, CA3, and subiculum) were imaged at atlas level P0-37. (It was only at this more posterior level that hippocampal differences were found in the adult.) In the neonate hippocampus, the pyramidal cell bodies show the darkest cytochrome oxidase staining (Di Rocco, Kageyama, & Wong-Riley, 1989), appearing as a thin line which, at this level, defines the dorsal-ventral span of the hippocampus proper. If this line is trisected, dividing the hippocampus proper into three parts, then CA1 readings can be taken within the lateral half of the most dorsal part and CA3 readings from the medial half of the most ventral part. Subicular (sub) readings can be taken adjacent to either the dorsal or ventral boundaries of CA1 and CA3, respectively. Midbrain regions were imaged at atlas level P0-44. The interpeduncular nuclei (IPN) stained darkly against background, while readings for the VTA were taken between the IPN and medial lemniscus. Dorsal raphe nuclei (DR) readings were taken at atlas level P0-49, adjacent to the midline and immediately ventral to the aqueduct.

RESULTS

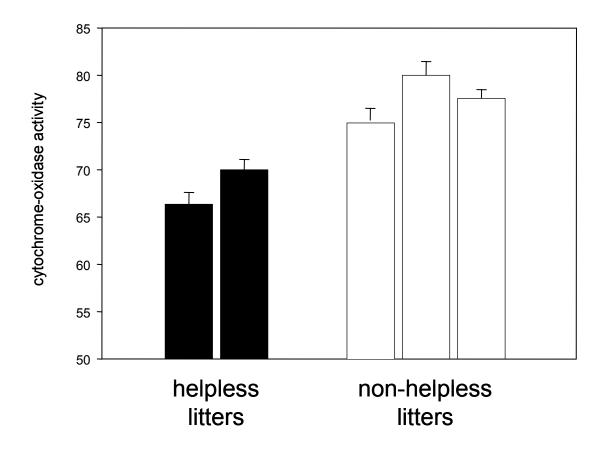
Means Analysis

Due to the small size of the newborn brains, sections which were randomly lost or damaged during sectioning had a greater impact on our ability to match anatomical levels across subjects. This resulted in several missing values, more or less randomly distributed across all subjects and regions. Because of this, the usual method of repeated measures ANOVAs could not be performed as this would have resulted in the list-wise exclusion of too many subjects. Therefore, all mean group data were analyzed using separate twofactor (Group × Sex) univariate ANOVAs for each region. This also meant that the compromise of applying the Bonferroni correction to each repeated measures ANOVA could not be used. Instead, corrections for multiple comparisons were not performed, but a more stringent alpha level (p < .01) was used to assess statistical significance.

There were no significant sex differences for any of the variables assessed, nor did sex interact with group to influence any of these measurements (data not shown). Birth weight was not significantly different between groups, F(1, 30) < 1, suggesting there were no gross differences in fetal development. There was, however, an overall difference in brain metabolism between groups, F(1, 30) = 41.2, p = 0.0000004, with congenitally helpless rats showing an 11% reduction in whole-brain cytochrome oxidase activity (Figure 5.1). Unlike adult congenitally helpless rats, which show a balance of severe hypermetabolism in a few regions and mild hypometabolism in several regions,

Figure 5.1: Whole-brain cytochrome oxidase activity values broken down by litter. Each bar is a separate litter.

Whole-Brain Activity Broken Down by Litter



congenitally helpless newborns showed unidirectional decreases in brain metabolism. Separate ANOVAs for each brain region revealed, though, that some regions were more hypometabolic than others, as displayed in Table 5.1. Estimates of regional influence, shown in Table 5.2, further revealed that the regions most affected in adult congenitally helpless rats—PVH, hippocampal system, and habenula—were the regions showing the largest effects in the newborn. Following these were the lateral septal nuclei and several medial PFC regions.

Functional Connectivity Analysis

Because the ANOVAs indicated no sex interactions, male and female subjects were pooled for the covariance analysis. Since a discriminant analysis could not be used to narrow the focus of the analysis (because of too many missing values), Pearson correlations were computed for all measured regions, with missing values dropped on a pair-wise basis. For ease of interpretation, several simplifications were made in reporting these correlations in Tables 5.3 and 5.4 and Figure 5.1:

- Regions showing no significant correlations in either group were omitted. Only basal ganglia regions (caudate-putamen, globus pallidus, and ventral pallidum) met this criterion.
- Cortical regions were not broken down into inner and outer layers because, in each group, layers within a region were highly correlated with each other, *r*(13-15) > .87, and did not differ in their significant correlations with other regions. Thus, correlations with cortical regions were averaged across layers.
- 3) The same averaging was done for other regions highly correlated with each other and not differentially correlated with other regions if they could also be regarded as functional units based on anatomical proximity. Thus, the core and shell of the accumbens, r(15) = .71 for helpless group, r(16) = .92 for non-helpless, were

Region	n	Helpless	n	Non-helpless	р
Amygdala, basolateral nucleus	13	77 ± 5	18	80 ± 4	.564
Amygdala, central nucleus	13	81 ± 5	18	87 ± 4	.354
Anterior cingulate, outer cortex	15	79 ± 3	16	92 ± 4	.014
Bed nucleus of the stria terminalis	13	65 ± 2	17	73 ± 2	.010
Caudate putamen	12	73 ± 2	14	72 ± 4	.859
Globus pallidus	14	70 ± 2	18	72 ± 2	.511
Habenula	13	$89 \pm 3*$	18	107 ± 3	.000
Hippocampus, CA1	12	$75 \pm 3*$	16	91 ± 2	.000
Hippocampus, CA3	12	$86 \pm 2*$	16	96 ± 2	.001
Infralimbic, outer cortex	14	$66 \pm 2*$	16	75 ± 2	.002
Interpeduncular nucleus	12	90 ± 4	15	101 ± 5	.099
Lateral septal nucleus	14	$67 \pm 2*$	18	78 ± 2	.001
Medial orbital, outer cortex	14	$63 \pm 2*$	14	71 ± 2	.005
Medial septal diagonal band	14	67 ± 3	18	74 ± 2	.119
Nucleus accumbens	15	50 ± 4	16	59 ± 4	.084
Paraventricular hypothalamus	13	$62 \pm 3^{*}$	19	81 ± 3	.000
Prefrontal, dorsomedial outer cortex	13	64 ± 2	14	70 ± 2	.048
Subiculum	12	$82 \pm 2*$	16	95 ± 2	.000
Ventral pallidum	14	67 ± 2	18	74 ± 2	.045
Ventral tegmental area	12	67 ± 4	15	77 ± 3	.031

Table 5.1:Mean \pm S.E. cytochrome oxidase activity (µmol/min/g tissue) inbrains of newborn helpless and non-helpless rats. * p < .01</td>

Table 5.2:Top 10 regions in terms of effect size for congenitally helpless newborns,
ranked according to relative magnitude of difference (left) or partial eta-
squared (right).

Region	% Dif.	Region	${oldsymbol{\eta}_{\mathrm{p}}}^2$
1. PVH	23	1. PVH	.63
2. Habenula	18	2. Ventral subiculum	.62
2. Ventral CA1	18	3. Ventral CA1	.60
4. Ventral subiculum	14	4. Habenula	.58
4. Lateral septal nucleus	14	5. Ventral CA3	.50
6. Infralimbic	12	5. Lateral septal nucleus	.50
7. Medial orbital	11	7. Infralimbic	.48
7. BNST	11	8. Medial orbital	.36
9. VTA	10	9. BNST	.34
9. Ventral CA3	10	9. Anterior cingulate	.34

		MO	IL	AC	Acb	LS	MSDB	BNST	Amyg	Hab	PVH	IPN
IL	R	.34										
	N	13										
AC	R	26	01									
110	Ν	11	14									
Acb	R	38	.15	.71								
1100	Ν	13	13	14								
LS	R	21	.52	.42	.68							
LO	Ν	14	16	15	15							
MSDB	R	38	.48	.47	.76	.79						
MISDD	Ν	14	16	15	15	18						
BNST	R	09	.60	.11	.28	.60	.68					
DINGI	Ν	14	15	14	15	17	17					
Amua	R	04	.17	.10	.08	.16	.46	.71				
Amyg	Ν	13	15	16	16	17	17	16				
Hab	R	26	.34	20	.26	.68	.51	.42	.29			
Пао	Ν	13	15	16	16	17	17	16	18			
PVH	R	23	.02	.23	.37	.64	.46	.36	.07	.58		
гүп	Ν	14	16	16	16	18	18	17	18	18		
IPN	R	.03	.57	.16	.53	.70	.67	.71	.45	.72	.43	
IPN	Ν	12	13	14	14	14	14	13	15	15	15	
DR	R	.46	.76	.25	.49	.78	.60	.69	.40	.36	.29	.71
DK	Ν	10	10	12	13	12	12	12	13	13	13	12

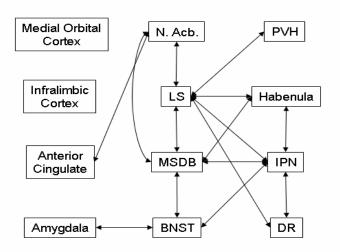
Table 5.3:Correlations of cytochrome oxidase activity, non-helpless newborns.
Significant correlations (p < .05) appear in bold.

Abbreviations: MO = medial orbital cortex, IL = infralimbic cortex, AC = anterior cingulate, Acb = accumbens nucleus, LS = lateral septal area, MSDB = medial septal/diagonal band nuclei, BNST = bed nucleus of the stria terminalis, Amyg = amygdala, Hab = habenula, PVH = paraventricular hypothalamus, IPN = interpeduncular nucleus, DR = dorsal raphe

		MO	IL	AC	Acb	LS	MSDB	BNST	Amyg	Hab	PVH	IPN
IL	r	.68										
	Ν	13										
ACv	r	.28	.30									
	Ν	14	14									
Acb	r	.11	01	.69								
	Ν	14	14	15								
LS	r	.49	.42	.26	.13							
	Ν	13	13	14	14							
MSDB	r	.64	.54	.24	.23	.61						
	Ν	13	13	14	14	14						
BNST	r	.72	.79	.26	02	.55	.79					
	Ν	12	12	13	13	13	13					
Amyg	r	.16	.24	.73	.71	.60	.40	.44				
	Ν	12	12	13	13	13	13	12				
Hab	r	15	29	20	48	19	15	25	13			
	Ν	12	12	13	13	13	13	12	12			
PVH	r	.40	.03	07	.06	.21	.22	.18	.08	.82		
	Ν	12	12	13	13	13	13	12	13	12		
IPN	r	.24	41	04	10	.18	.30	.04	.15	.45	.53	
	Ν	11	11	12	12	11	11	10	11	11	11	
DR	R	06	62	38	.22	32	.04	08	14	30	.07	.62
	Ν	10	10	11	11	11	11	10	11	10	11	9

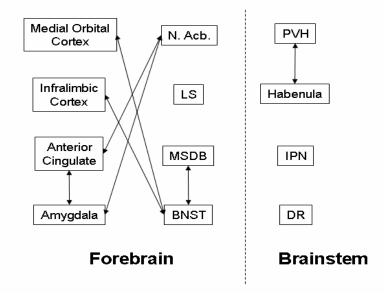
Table 5.4: Correlations of cytochrome oxidase activity, helpless newborns. Significant correlations (p < .05) appear in bold.

Abbreviations: MO = medial orbital cortex, IL = infralimbic cortex, AC = anterior cingulate, Acb = accumbens nucleus, LS = lateral septal area, MSDB = medial septal/diagonal band nuclei, BNST = bed nucleus of the stria terminalis, Amyg = amygdala, Hab = habenula, PVH = paraventricular hypothalamus, IPN = interpeduncular nucleus, DR = dorsal raphe. Figure 5.2: A diagram of the significant correlations among brain regions in helpless vs. non-helpless rats. All correlations are positive. The dashed line in the helpless diagram emphasizes the functional decoupling between forebrain and brainstem regions in this group.



NON-HELPLESS

HELPLESS



unified as the nucleus accumbens (Acb); the basolateral and central nuclei of the amygdala, r(13-18) > .90 for both groups, were unified as the amygdala (Amyg); the medial septal and diagonal band nuclei, r(14) = .84 for helpless group, r(18) = .71 for non-helpless, were unified as the MSDB.

- 4) Isolated correlations (when two regions were correlated exclusively with each other) were also omitted from the tables and figure. These consisted of, in the helpless group, coupling between CA1 and subiculum, r(12) = .76, and between the VTA and PAG, r(12) = .85, and, in the non-helpless group, coupling between CA1 and CA3, r(16) = .79.
- 5) Dorsomedial PFC was omitted from the tables because, in both groups, it was correlated only with medial orbital cortex and the correlations were similarly high: r(13) = .90 for helpless group, r(13) = .80 for non-helpless.

Following these simplifications, Tables 5.3 and 5.4 and Figure 5.1 show that nonhelpless rats had twice as many significant correlations as helpless rats, and, with the exception of coupling between the anterior cingulate and nucleus accumbens, and between the MSDB and BNST, the regions showing significant correlations were completely different across groups. Notably, the non-helpless newborns showed extensive correlations both within the basal forebrain (Acb, LS, MSDB, BNST) and between the basal forebrain and much of the rest of the brain (amygdala, PVH, habenula, IPN, and DR). These correlations were strikingly absent in the helpless newborns: instead, more correlations appeared between the BNST and medial orbital and infralimbic cortex, between the amygdala and anterior cingulate and nucleus accumbens, and between the habenula and the PVH.

In particular, the LS and IPN appeared to be network hubs in the non-helpless brain; together they accounted for two-thirds of the significant correlations in this group. In marked contrast, the LS and IPN showed no significant correlations with any regions in the helpless group.

DISCUSSION

Despite decreased whole-brain metabolism in congenitally helpless newborns, only a third of the regions examined showed a statistically reliable difference, and, more important, the regions most hyperactive in the adult showed the largest effect sizes in the newborn. In particular, the PVH showed the largest group difference in both adults and newborns. This means in effect that the brain regions most underactive at birth go on to become the most hyperactive in adulthood. What could account for this radical shift? One consideration is that congenitally helpless rats show abnormal reversals of HPA-axis responsiveness across development. Relative to controls, adrenocorticotropin hormone (ACTH) response to cold stress is hypoactive at postnatal day 7 but hyperactive at postnatal day 14 (King et al., 1993), and ACTH responsiveness in adulthood may be either blunted or hyper depending on the type and timing of stress encountered during early life (King et al., 1993; King & Edwards, 1999). Thus, a developmental difference in stress responsiveness could account for the metabolic reversal.

Another important consideration is that, in the barely sensate newborn, cytochrome oxidase may predominantly reflect morphogenetic intrauterine differences. This is because most brain pathways still being laid down at birth have yet to be functionally activated, such that cytochrome oxidase activity may simply reflect the genesis and maintenance of synapses, independent of the sensorimotor activation of those synapses (Di Rocco et al., 1989). In this light, reduced cytochrome oxidase in the PVH may reflect a loss of connectivity with other regions.

The current covariance analysis of functional connectivity corroborates this story. Before discussing this, it is important to qualify what functional connectivity means in the context of cytochrome oxidase data. With most functional imaging techniques, which assess blood flow or metabolism in the here and now, brain regions couple and decouple on a dynamic basis according to stimulus or task, forming different networks to accomplish different functions (Horwitz, 1994). However, because cytochrome oxidase is a historical marker, reflecting past metabolic demands instead of current ones, its functional couplings are relatively static. Transient dynamic couplings will tend to average out over time, leaving correlations that reflect stable anatomical relationships between regions. Thus, cytochrome oxidase correlations should be interpreted as reflecting behavioral predispositions conferred by anatomical wiring (Sakata, Coomber, Gonzalez-Lima, & Crews, 2000). This is expected to be even more true in the case of neonates since metabolic activity at this phase of development is primarily concerned with the establishment of anatomical connections, rather than the recruitment of specific connections for specific behaviors (Di Rocco et al., 1989).

In the congenitally helpless newborns, the correlations suggest that the PVH is cut off from forebrain regulation, along with every other brainstem region (LHb, MHb, IPN, DRN). Of all the forebrain inputs to the PVH, perhaps the most important comes from the subiculum, which projects not only to the zone surrounding the PVH, but also to other regulatory regions, such as the LS and BNST (Herman et al., 2003). The subiculum is thus well-positioned to translate the activation of glucocorticoid receptors, which the hippocampus has in abundance (McEwen, Weiss, & Schwartz, 1968), into feedback inhibition of the PVH, but the subiculum may also inhibit the PVH response to certain stressors even in the absence of glucocorticoid signaling (Herman, Dolgas, & Carlson, 1998).

Neither strain of newborns showed hippocampal coupling with the PVH or with any other regions. This is likely because of the relative immaturity at birth of the hippocampus, which undergoes extensive postnatal development and begins to show functional coupling with other regions only a few days prior to weaning (Nair & Gonzalez-Lima, 1999). However, next to the PVH, the subiculum and CA1 field showed the largest effect sizes in the newborns, perhaps indicating their future importance in determining the helpless phenotype. Again, hypometabolism in the immature hippocampus suggests that it is developing fewer axons and/or dendrites to link it with other regions (Di Rocco et al., 1989).

Assuming that primary causal pathology is more likely to be present at birth and secondary consequential adaptation is more likely to occur during postnatal development, the results may help to resolve some competing causal scenarios suggested by the adult brain map. For example, reduced VTA metabolism accompanied increased habenula metabolism in congenitally helpless adults. Since the habenula and VTA share mutually inhibitory connections, it was possible that habenula hyperactivity had decreased VTA activity, or that VTA hypoactivity had increased habenula activity. (See Chapter 4.) The present results suggest the former scenario since the habenula showed a more significant difference in newborns. Thus, deficits in psychomotor and reward-related circuitry may stem from the habenula, which showed the second largest mean difference in both neonates and adults. In addition, the habenula appears decoupled in helpless newborns from the IPN, its primary efferent target through which it influences various brainstem nuclei (Contestabile & Flumerfelt, 1981). Perhaps the most important of these brainstem targets is the DR (Wang & Aghajanian, 1977), which is also decoupled from the IPN in helpless newborns. Interestingly, helpless newborns showed novel coupling between the habenula and PVH that was not present in non-helpless newborns. We have found no evidence in the literature for projections which might directly link these two regions, nor were they functionally coupled to any of the sampled regions which could serve as intermediaries. Intraventricular corticotropin-releasing hormone (CRH) administration was found to increase c-fos expression in the habenula (Imaki, Shibasaki, Hotta, & Demura, 1993), but it is unclear how this functional link is established since the habenula does not express CRH receptors (Van Pett et al., 2000). It does appear, however, that the habenula and PVH operate as a functional unit in congenitally helpless rats, since both regions were severely hypermetabolic in the adult.

Among those regions which remain hypometabolic in the congenitally helpless adult, the LS showed the largest effect size in the newborn. It also showed the greatest difference in functional coupling, being connected to the nucleus accumbens, MSDB, and every single brainstem nucleus in non-helpless newborns while being completely disconnected in helpless newborns. The LS contributes a significant number of GABA projections to the PVH (Herman et al., 2003), and the relevance of LS dysfunction to depression vulnerability is supported by the effects of septal lesions, which increase susceptibility to mild stress (Brown, Uhlir, Seggie, Schally, & Kastin, 1974; Seggie, Uhlir, & Brown, 1974; Uhlir, Seggie, & Brown, 1974).

Another causal question stems from the relationship between PVH hyperactivity and frontal-cingulate metabolism: does a hyperactive HPA axis alter PFC function, or do PFC deficits promote a hyperactive HPA axis? The current results argue against the former scenario since a PFC abnormality exists prior to the onset of PVH hyperactivity. However, the smaller effect sizes for PFC regions suggest that, relative to the hippocampus, habenula, and LS, the PFC plays more of a supporting role in determining the helpless phenotype. Interestingly, helpless newborns showed increased functional connectivity between medial PFC regions and the BNST, and between the amygdala, anterior cingulate, and nucleus accumbens. It is unclear what role these extra functional connections play in the helpless predisposition, but a human PET study showed that positive coupling between the amygdala and anterior cingulate in posttraumatic stress disorder (PTSD) was one of the few functional connections that differentiated PTSD patients from healthy trauma survivors (Gilboa et al., 2004).

In conclusion, the major findings were that the PVH was the most altered brain region in congenitally helpless newborns and that it, along with all other brainstem regions, was functionally decoupled from the forebrain. These results suggest that the essence of the helplessness vulnerability does not reside in an endogenously hyperactive HPA axis per se, but rather in the limbic networks which regulate it. This implies that when a congenitally helpless animal encounters a stressful situation, no matter how mild, regulatory regions such as the hippocampus, LS, and medial PFC may be unable to constrain the response of the PVH. Because congenitally helpless animals also lack functional glucocorticoid receptors (Lachman et al., 1993), which ordinarily might exert some local negative feedback, the PVH, once activated, would be unable to turn itself off. This could, over time, lead to the chronic PVH hypermetabolism seen in congenitally helpless adults.

Chapter 6: Response to Treatment

All current antidepressant medications require several weeks of administration before clinical efficacy is observed (Hickie, Scott, & Davenport, 1999; Nurnberg, Thompson, & Hensley, 1999). This response delay follows myriad neurophysiological adaptations to enhanced aminergic transmission, including autoreceptor desensitization, multiple increases and decreases of postsynaptic receptor sites, altered second-messenger pathways, and enhanced neurotrophism (Blier & de Montigny, 1999; Duman, Malberg, & Thome, 1999; Hyman & Nestler, 1996). However, the brain regions mediating these events are not well characterized.

The study reported in this chapter examined changes in regional brain cytochrome oxidase activity associated with 2 weeks of fluoxetine treatment in adult congenitally helpless males. Adult males were chosen as subjects so that their brain maps could be compared to those of the untreated adult males reported in Chapter 4. Fluoxetine was chosen because it has become a first-line drug for the treatment of affective disorders and it is the only drug of its class—the selective serotonin reuptake inhibitors (SSRIs)—whose effects have been metabolically mapped in the rat brain following chronic administration. Freo, Ori, Dam, Merico, and Pizzolato (2000) examined resting glucose metabolism in over fifty regions following two weeks of daily fluoxetine administration. They found changes in only two regions: the lateral habenula and CA3 field of the hippocampus both showed significant decreases.

Although the human imaging literature on antidepressant treatment effects is highly variable, studies using SSRIs have most often found increases in prefrontal and anterior cingulate cortex and decreases in the hippocampus and in ventromedial frontal, subgenual cingulate, and anterior insular cortex (Brody et al., 1999; Buchsbaum et al., 1997; Mayberg et al., 1999). For purposes of comparison with our study, we have focused on one exceptional study by Mayberg et al. (2000), who examined glucose metabolism in depressed males both one week and six weeks after initiating fluoxetine treatment. The authors found some changes that were common at both time points, some that were opposite, and some that were only present after depression remission. Changes present at one week which persisted at six weeks included increased metabolism in the rostral brainstem, premotor cortex, and inferior parietal cortex, and decreased metabolism in the caudate, medial thalamus, insular cortex, and cerebellum. Initial increases were observed in the hippocampus, medial temporal cortex, putamen, and pallidum, but net decreases were observed in these same regions at six weeks. The opposite was seen in the posterior cingulate, where an initial decrease was followed by a final increase. Changes in prefrontal and cingulate cortex were only observed after depression remission. These changes constituted a normalization of prefrontal cortex and dorsal anterior cingulate metabolism, but also a decrease of subgenual cingulate metabolism to below normal.

METHODS

Subjects

Twenty congenitally helpless males were selected from the cohort of subjects used in the behavioral characterization study reported in Chapter 2. These rats were randomly assigned for either fluoxetine treatment (n = 10) or vehicle control (n = 10). Drug administration began when the subjects weighed 450-550 g, approximately 3 months after the end of behavioral testing which took place between postnatal days 29 and 48. Rats were individually housed at this time in 45 × 24 × 21 cm cages in a temperature-controlled room ($22 \pm 1^{\circ}$ C). They were maintained on a 12/12 hour light cycle and provided continuous food and water.

Drug Administration

Fluoxetine hydrochloride (Sigma) was dissolved in a 25% dimethylsulfoxide (DMSO)-75% saline vehicle (2.5 mg/ml). For 2 weeks, rats received daily i.p. injections of either 5 mg/kg fluoxetine/vehicle or vehicle. Injections were administered during the last hour of the light cycle.

Forced-Swim Test

On the last day of drug administration, rats were placed in a glass cylinder (46 cm high \times 20 cm in diameter) containing a 30-cm water column (24 ± 1 °C). Two swimming sessions were conducted: an initial 15-min pretest, followed by a 5-min test 24 h later. Test sessions were video-taped for scoring. A time-sampling technique was employed to score several behaviors during a single viewing. This previously described method has proven reliable and valid for detecting the effects of different antidepressant drugs (Detke, Rickels, & Lucki, 1995). At the end of each 5-s period during the 5 min test, the scorer rated the rat's behavior as belonging to one of the following behavioral categories: (1) immobility—floating passively in the water and only making slight movements to keep the head above the water line (Porsolt, Le Pichon, & Jalfre, 1977); (2) swimming—making active swimming motions, more than necessary to merely keep the head above water; or (3) climbing—making active movements with forepaws in and out of the water, usually directed against the walls. Two raters blind to the treatment conditions scored all of the behaviors. Scores for each behavior were expressed as percentages of time.

Tissue Processing

Immediately after the conclusion of the forced-swim test, rats were taken to a separate room and decapitated. Two brains were damaged during decapitation, so the *n*s for the brain analysis were 9 for each group. Brain extraction and processing proceeded

according to the protocol for quantitative cytochrome oxidase histochemistry outlined in Chapter 3.

Delineating Regions of Interest

Regions of interest were limited to regions showing significant effects in the previous studies, except for the dorsal raphe (DR). Although the DR did not show significant mean differences in either congenitally helpless adults or newborns, it was of interest because fluoxetine, an SSRI, might be expected to alter its serotonergic output. Regions were imaged according to the specifications given in Chapter 4.

Statistical Methods

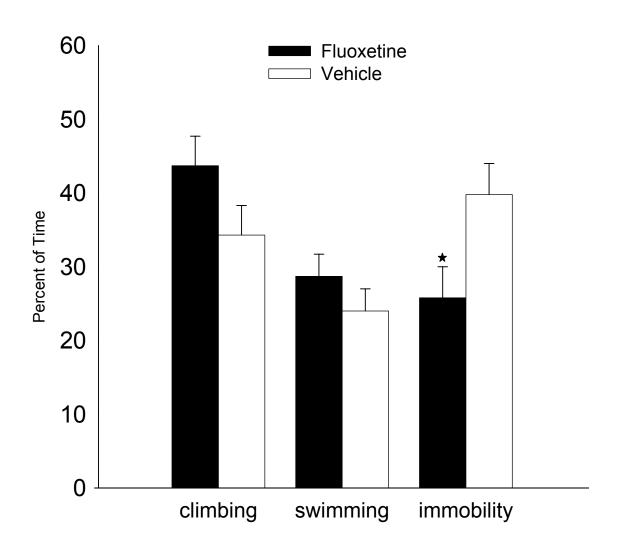
Repeated measures ANOVAs were performed according to the same groupings given in Chapter 4. A multivariate discriminant analysis and pair-wise correlations were also performed according to the criteria outlined in Chapter 3.

RESULTS

Forced-Swim Test

Due to animal welfare concerns, we agreed to terminate the swim test if at any time a subject remained completely submerged for 5 consecutive seconds. Unfortunately, this meant that during the initial swim session, only 11 out of 20 subjects were able to swim the entire 15 minutes. Of the remaining subjects, the test was terminated between 5.5 and 14.5 minutes (mean = 11.5 min). Three of these subjects were from the fluoxetine-treated group (mean = 11.8 min), and six were from the vehicle-control group (mean = 10.9 min). This, of course, introduces a possible bias into the interpretation of the results, in that some rats experienced a shorter duration of swim stress than others. Thus, rats experiencing less stress might show reduced immobility in the final test, regardless of group assignment. However, this appears not to have been the case since a

Figure 6.1: Effect of two weeks of administration of fluoxetine or vehicle on immobility, climbing, and swimming in the forced swim test in adult male congenitally helpless rats. $\star p < .05$



linear regression indicated that, if anything, rats with earlier sink times on day 1 tended to show more, not less, immobility on day 2, r(9) = -.65, p = .06. Therefore, all subjects were included in the univariate ANOVAs of the forced-swim scores. These tests indicated that fluoxetine-treated rats showed significantly less immobility than vehiclecontrol rats, F(1,18) = 5.5, p = .03, while swimming and climbing measures were not significantly different between groups, Fs(1, 18) = 1.2 and 2.7, ps = .29 and .12, respectively (Figure 6.1).

Regional Effects

Means and standard errors for all regions of interest are reported in Table 6.1. A univariate ANOVA of whole-brain activity found no significant difference between groups, F(1, 16) = .091, p = .70. Separate repeated measures ANOVAs were performed for each group of regions (frontal-cingulate cortex, basal ganglia, septal nuclei, ventral hippocampus, PVH, and the habenula and its efferent targets). The only significant omnibus finding was a two-way (Group × Region) interaction involving the habenula and VTA, F(3, 48) = 5.3, p = 0.003. Simple-effects tests showed that mean cytochrome oxidase activity in the fluoxetine-treated group was significantly decreased in the habenula, F(1, 16) = 10.4, p = .005, and significantly increased in the VTA, F(1, 16) = 9.1, p = .008.

The habenula and VTA had almost equivalent univariate effect sizes, $\eta_p^2 = .39$ and .36, respectively. Moreover, the mean differences in activity differed by only a couple of points, although this translated into a larger percent change for the VTA since it stains much lighter than the habenula (15% vs. 8%). Likewise, a multivariate stepwise discriminant analysis selected only the VTA and habenula, which had equal and opposite standardized discriminant coefficients. However, despite the opposite changes in the

Table 6.1:Effects of fluoxetine on cytochrome oxidase activity (μ mol/min/g tissue).Data are mean \pm S.E. *p < .05.</td>

Region	Fluoxetine	Vehicle
Amygdala, basolateral nucleus	244 ± 9	249 ± 7
Amygdala, central nucleus	226 ± 13	230 ± 9
Anterior cingulate, outer cortex	278 ± 15	293 ± 10
Bed nucleus of the stria terminalis	228 ± 9	231 ± 3
Caudate putamen	315 ± 10	335 ± 8
Globus pallidus	160 ± 7	157 ± 4
Habenula	$282 \pm 5*$	308 ± 6
Hippocampus, CA1	206 ± 8	224 ± 3
Hippocampus, CA3	235 ± 2	252 ± 5
Infralimbic, outer cortex	229 ± 5	241 ± 7
Interpeduncular nucleus	392 ± 7	383 ± 10
Lateral septal nucleus	286 ± 13	310 ± 10
Dorsal raphe nucleus	212 ± 8	212 ± 9
Medial septal diagonal band	213 ± 12	223 ± 2
Nucleus accumbens	260 ± 9	271 ± 9
Paraventricular hypothalamus	156 ± 7	173 ± 5
Prefrontal, dorsomedial outer cortex	290 ± 6	289 ± 6
Subiculum	251 ± 9	254 ± 8
Ventral pallidum	202 ± 8	202 ± 4
Ventral tegmental area	$172 \pm 5*$	149 ± 6

VTA and habenula, the regions were not functionally coupled in either the treated or untreated groups, rs(9) = .22 and .13, respectively.

DISCUSSION

There are two principal findings of this study. The first is that 2 weeks of fluoxetine treatment reduced immobility in the forced-swim test. The second is that, out of several regions which showed cytochrome oxidase differences in congenitally helpless adults and newborns, only the habenula and VTA responded significantly to fluoxetine treatment, and both changes were in the direction of metabolic normalization—a decrease in the habenula and an increase in the VTA.

Forced-Swim Test

Two weeks of fluoxetine administration (5 mg/kg/day) reduced immobility in the forced swim test, a finding consistent with studies in normal rats (Vazquez-Palacios, Bonilla-Jaime, & Velazquez-Moctezuma, 2004). However, the results were unusual in a number of ways. First, fluoxetine and other SSRIs typically reduce immobility by increasing swimming without affecting climbing (Detke et al., 1995; Vazquez-Palacios et al., 2004), whereas antidepressants targeting the norepinephrine system do just the opposite (Detke et al., 1995). The congenitally helpless rats in our experiment did not show a significant difference in either swimming or climbing, indicating an additive effect of both behaviors in reducing immobility. More important, climbing is typically the least prevalent behavior in both treated and untreated groups (Detke et al., 1995; Vazquez-Palacios et al., 2004), whereas in our study it was the most prevalent. The reason for this aberration is unclear, but it does not appear to be caused by the congenital helplessness phenotype since the only other study assessing forced swim behavior in this strain found the typical pattern in response to an experimental 5-HT_{2A} receptor

antagonist: increased swimming was solely responsible for decreased immobility, while climbing remained the least frequent behavior (Patel et al., 2004). Even the congenitally non-helpless rats in Patel et al.'s experiment climbed far less than our untreated congenitally helpless rats.

One possible explanation for the discrepancy is that we erroneously identified as climbing many behaviors which another investigator would have considered swimming. While we relied on published descriptions of these swim behaviors for our scoring, climbing is a dramatic behavior, involving thrashing and splashing which should be hard to mistake for swimming. Another possible explanation is that our rats were larger than those typically used (450-550 g vs. 250-300 g). Given the narrowness of the cylinder, the larger rats were forced into a mostly vertical posture which restricted their horizontalrotational range of motion. This may have led to an artifactual increase in climbing and decrease in swimming. However, Patel et al. (2004) used congenitally helpless rats of approximately the same size (400-500 g), which apparently had no bearing on their swim-test results. One final possibility is that the extensive handling and behavioral testing conducted when these rats were 3-6 weeks old may have constituted a form of early environmental enrichment, which may have had a long-lasting beneficial impact on these animals. However, the increased propensity for these animals to sink during the initial 15 minutes of swimming would seem to indicate vulnerability to this test, not resistance. But, again, Patel et al. (2004) did not report any abnormalities during the initial forced-swim training with their congenitally helpless animals. In conclusion, all of these explanations are possible, but none of them are obvious. The experiment would need to be repeated under many conditions (different scorers of behavior, presence vs. absence of behavioral testing in early life, varying ages and weights, or smaller vs. larger cylinders) in order to resolve this issue.

Regional Effects

Cytochrome oxidase changes identified the habenula and VTA as the only regions responding to 2 weeks of fluoxetine treatment. These results are fairly consistent with the 2-DG study of Freo et al. (2000), who found baseline changes in only the CA3 pyramidal layer and in the habenula in response to the same duration of treatment. (They did not assess the VTA.) Like Freo et al., we found habenula metabolism to be significantly reduced in treated rats. One human imaging study has also implicated the habenula in the therapeutic response to antidepressant treatment. Using a group of depressed patients who had recovered under antidepressant medication, Morris et al. (1999) demonstrated that habenula metabolism increased in response to tryptophan depletion, but only in subjects who experienced a relapse of depressive symptoms. This suggests that antidepressant effects dependent on serotonin are related to an inhibition of habenula activity. These are the only two studies to my knowledge which have examined the habenula in connection with antidepressant response. Thus, the current finding of a fluoxetine-induced reduction of habenula metabolism adds to an extremely small but consistent literature.

The finding of increased VTA metabolism concurrent with decreased habenula metabolism adds to a large number of metabolic mapping studies which have consistently found opposite changes in these two regions (see pages 70-71), but increased VTA activity appears inconsistent with several studies showing that fluoxetine acutely antagonizes the dopamine system and suppresses the firing rates of VTA neurons (Esposito, 1996; Prisco & Esposito, 1995). However, after 3 weeks of chronic fluoxetine administration, acute administration of fluoxetine no longer affected the firing rates of VTA neurons (Prisco & Esposito, 1995), suggesting a compensatory response had occurred. Increased metabolic capacity of excitatory synapses innervating the VTA might account for its resistance to fluoxetine inhibition after chronic administration.

Alternatively, fluoxetine may have a normalizing effect on VTA metabolism in congenitally helpless rats which might not be seen in normal rats.

While our results are consistent with those obtained from normal rats (Freo et al., 2000), we had hypothesized that we might see more widespread changes in line with those observed in human imaging studies of depression treatment, such as normalization of metabolic deficits in prefrontal cortex. This was not the case. There are several possible explanations for out negative findings, the most simple being that the rat brain may not respond to antidepressants in the same way the human brain does, at least not in terms of regional metabolism. Alternatively, we may have observed more widespread changes if we had administered fluoxetine at a higher dose or for a longer time. For example, Mayberg et al. (2000) did not observe normalization of cortical metabolism until after 6 weeks of treatment. Although still more widespread than our observations, changes that Mayberg et al. (2000) observed after 1 week did include decreased metabolism in the medial thalamus and increased metabolism in the brainstem. The habenula and VTA, respectively, might be involved in these effects, in which case the directions of change are consistent with our findings. Yet another possibility is that, despite a modest response to fluoxetine in the forced-swim test, congenitally helpless rats might be resistant to fluoxetine treatment or might respond better to another class of drug. Regardless, the results suggest that the habenula and VTA are highly sensitive to chronic fluoxetine treatment and are among the first regions to adapt to it. These regions are then well-positioned to affect the function of many other systems.

Chapter 7: Conclusion

FROM THE NEWBORN BRAIN MAP TO THE ADULT BRAIN MAP

In comparing the brain maps of newborn and adult congenitally helpless rats, the effects may be categorized as follows: 1) brain regions affected at birth and in adulthood, 2) brain regions affected in adulthood but not at birth, and 3) functional couplings present at birth but not in adulthood.

Brain Regions Affected in Both Adults and Newborns

Among those brain regions showing significant differences in both adults and newborns, all of the regions (except for the IPN) which showed significant hypermetabolism in adulthood showed significant differences at birth; however, the changes were in the opposite direction. In contrast, almost none of the regions which showed hypometabolism in adulthood were significantly different at birth, the exceptions being the LS and medial orbital cortex.

Regions Showing Opposite Effects

The PVH, habenula, ventral hippocampus, and infralimbic cortex were hypometabolic at birth but hypermetabolic in adulthood, but, interestingly, the relative magnitudes of the differences showed a strikingly similar pattern. Namely, the PVH and habenula showed, respectively, the greatest and second greatest mean difference in both adult and newborn congenitally helpless rats. This means in effect that the regions most hypometabolic at birth become the most hypermetabolic in adulthood.

Possible explanations for this finding were discussed extensively in Chapter 5. In summary, the most likely interpretation stems from developmental differences in the cytochrome oxidase stain. During the prenatal period, cytochrome oxidase is functionally coupled to the growth and development of synapses, but during the postnatal period, there is a shift to functional coupling with synaptic activity. This difference between *connectivity* on the one hand and *activity* on the other was judged to underlie the shift from hypometabolism to hypermetabolism. In essence, impoverished connectivity of a region could causally lead to hyperactivity of that same region, particularly if there were a selective loss of inhibitory inputs. For example, if the PVH received reduced regulatory input from forebrain regions (as the correlational study in the newborns seems to suggest), then it would have a propensity to overreact to environmental stimuli which normally recruit it. This would likely be reflected by reduced regional cytochrome oxidase activity at birth and increased activity following postnatal experience. However, while our brain mapping results are consistent with this explanation, they do not prove it. A more direct follow-up experiment would be to inject a retrograde tracer into the zone surrounding the PVH; if there were reduced labeling in the forebrain and hippocampus of congenitally helpless rats, the reduced connectivity hypothesis would be supported.

Regions Showing Common Effects

The LS and medial orbital cortex showed reduced metabolism in both adults and newborns. However, this does not mean that they are hypometabolic for the same reasons. As with the regions showing opposite effects, reduced cytochrome oxidase activity in the newborn likely reflects reduced connectivity with other regions. The interregional correlations in the newborn suggest this is especially true for the LS, which was the most extensively interconnected region in the non-helpless newborn but was completely disconnected in the helpless newborn. Indeed, the metabolic deficit was even larger in the newborn than in the adult for this region, the only region to show such an effect. As for why these regions remain hypometabolic in adulthood, there are two possibilities. One is that there is a modular malfunction in these regions, independent of their connections with other regions, and certainly the work of Petty and colleagues points to the LS as a key player in the development of helplessness (Ronan et al., 2000; Sherman & Petty, 1984; Steciuk et al., 1999). The other possibility is that adult hypoactivity follows from newborn hypoconnectivity. This would be similar to the scenario suggested for the regions showing opposite effects, except in this case a relative loss of excitatory influence to the LS could result in hypometabolism at both developmental time points.

Brain Regions Affected in Adults but not in Newborns

In attempting to account for why some regions affected in adults were not significantly affected in newborns, one must first acknowledge that several of these regions came close to achieving significance (p < .10) in the newborn, such as the VTA, BNST, IPN, nucleus accumbens, and ventral pallidum. It would therefore be premature to make too much of the "absence" of these effects although some regions, namely the amygdala and basal ganglia, were much more equivocal between newborn groups.

There are two general explanations for the apparent involvement of additional regions in the congenitally helpless adult. One is "spreading activation": that is, due to abnormalities in some structures at birth, additional regions may alter their activity during development as a consequence of receiving more or less input from these regions. The possible scenarios here are almost infinite and impossible to determine from the current data, but, as an example, decreased inhibitory input to the habenula (reflected by neonatal hypometabolism) could translate into increased habenula output to the IPN and VTA. Because the habenula appears to have a net excitatory influence over the IPN and inhibitory influence over the VTA (see Chapter 4, Discussion), this could translate into the IPN hypermetabolism and VTA hypometabolism seen in the adult. Reduced VTA output to the basolateral and central nuclei of the amygdala could then cause hypometabolism in these regions. Indeed, as discussed in Chapter 4, the basolateral and

central nuclei (which receive VTA projections) showed changes in the adult, whereas the other amygdala nuclei (which do not receive VTA projections) did not. However, one would need to perform additional metabolic mapping experiments at numerous developmental time points to test this spreading-activation hypothesis.

The other general explanation for the appearance of new effects in the adult may be postnatal experience, including possible differences in maternal care and peer interaction. These two explanations—spreading activation and postnatal experience—are not mutually exclusive and may in fact be synergistic. For example, brain regions altered at birth may predispose the animals to react to certain experiences and engage in certain behaviors in a way which feeds back on the brain and alters its function. Indeed, this was suggested to be the case for explaining PVH and habenula hypermetabolism, which is hypothesized to result from an interaction between reduced regulatory input from other brain regions and some minimal form of environmental stress. Similar diathesis-stress interactions may produce additional changes in other brain regions not primarily altered at birth.

Functional Couplings Present in Newborns but not in Adults

Unlike both the helpless and non-helpless newborns, which showed extensive interregional correlations, neither the helpless nor non-helpless adults showed any significant interregional correlations. There are several explanations for this, one of which may simply be that there was an insufficient number of subjects to detect significant correlations in the adults. (Because male and female subjects were pooled, the sample size was about twice as large for the newborns.) A more likely explanation may again have to do with the functional meaning of cytochrome oxidase staining in the newborn, which, as discussed previously, is more indicative of the growth of anatomical connections. Thus, regions forming synapses with each other may be more likely to show correlations in their cytochrome oxidase activity. However, once cytochrome oxidase activity becomes coupled to synaptic activations, transient dynamic couplings between regions, which might be revealed by correlations in deoxyglucose uptake, might go undetected by correlations in cytochrome oxidase, which reflect a much longer time span.

FROM BRAIN MAPS TO BEHAVIOR

Congenitally helpless rats, by design, are highly susceptible to develop learned helplessness. In addition, the behavioral characterization experiments in Chapter 2 showed two principle findings: increased novelty seeking but decreased reward seeking. How do the brain maps relate to these behavioral predispositions? As discussed in Chapter 2, one biological marker that has been linked to both vulnerability to learned helplessness and increased novelty seeking is impaired glucocorticoid signaling in the hippocampus (Kabbaj et al., 2000; Papolos et al., 1993). Glucocorticoid receptor activation has been shown to suppress the excitability of hippocampal neurons (de Kloet, 2004), to reduce blood flow to the hippocampus (Endo, Nishimura, Kobayashi, & Kimura, 1997), and to decrease glucose uptake by hippocampal cells (Horner, Packan, & Sapolsky, 1990). Vice versa, adrenalectomy was shown to increase blood flow to the hippocampus (Endo, Nishimura, & Kimura, 1994). These lines of evidence are consistent with the increase in cytochrome oxidase activity found in the ventral hippocampus of adult congenitally helpless rats and their increased predisposition to novelty seeking and learned helplessness. Markedly increased expression of CRH mRNA was also found in the PVH of high novelty-seeking rats (Kabbaj et al., 2000), which suggests that the PVH hypermetabolism in adult congenitally helpless rats may also be related to their novelty-seeking temperament (and may also stem from reduced glucocorticoid signal). The other brain region most consistently linked with exploratory behavior, especially rearing, is the septal area. However, the literature suggests that septal activation increases rearing and septal lesions decrease rearing (Monmaur, Sharif, & M'Harzi, 1997b; Lee et al., 1988; Myhrer, 1989a; Poplawsky & Isaacson, 1983b), which is not consistent with the finding of decreased septal metabolism in both congenitally helpless adults and newborns. However, decreased activity of the LS is consistent with increased vulnerability to helplessness (Ronan et al., 2000; Steciuk et al., 1999), as is its apparent functional decoupling from other brain regions, as observed in the newborn helpless rats.

Hypometabolism of the basal ganglia would seem to predict psychomotor retardation; indeed, this is how the finding in humans has generally been interpreted. However, the congenitally helpless rats do not show any evidence of psychomotor retardation, and, if anything, appear more active than non-helpless rats. Hypometabolism in the ventral striatum and pallidum, however, is consistent with the reduced reward-seeking behavior (in the form of sucrose consumption) of congenitally helpless rats, as is hypometabolism in the VTA and the basolateral and central amygdala (see Chapter 4, discussion). In addition, reduced amygdala metabolism may be related to the role of this region in fear and anxiety, which congenitally helpless rats appear to show less of, at least in the open-field and light-dark test (see Chapter 2). And, as already discussed repeatedly, habenula hypermetabolism may be a state marker of depression (Caldecott-Hazzard et al., 1988) and habenula activation appears necessary for the development of learned helplessness (Amat et al., 2001).

In the end, though, it is premature at this point to relate specific regional cytochrome oxidase changes to specific behaviors. Ideally, one would like to show that differences in habenula metabolism, for example, correlate with some of the behavioral measures shown to distinguish helpless from non-helpless rats. However, attempts to demonstrate this with the vehicle control subjects used in the fluoxetine study were

unsuccessful. This could be due to any number of reasons, including non-linear effects or insufficient sample size. Future studies aimed at correlating individual differences in regional cytochrome oxidase differences with individual behavioral differences in a large normative sample should shed more light on this question.

THE HABENULA AS A THERAPEUTIC TARGET

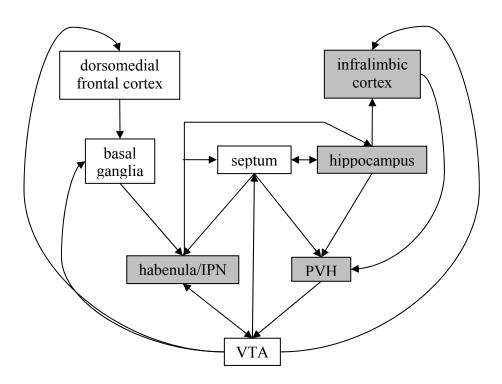
Caldecott-Hazard et al. (1988) showed that habenula hypermetabolism was the common denominator across three animal models of depression, and this finding has now been extended to a rat strain that models the predisposition to depression and PTSD. Moreover, the habenula was the only region showing a significant decrease in metabolism following 2 weeks of antidepressant treatment. As shown in the circuit diagramed in Fig. 8.1, the habenula \rightarrow IPN pathway appears to serve as an interface between frontostriatal and septohippocampal systems and the brainstem monoaminergic nuclei, of which the most important for helplessness susceptibility may be the VTA. The habenula \rightarrow IPN is linked to PVH regulatory regions via a major input to the habenula from the entire septal area, including LS and BNST (Felton et al., 1999), and via a major output from the IPN to the hippocampus (Montone, Fass, & Hamill, 1988). The IPN further receives cholinergic input from the medial septal nucleus and reciprocates a serotonergic projection to the basal forebrain and ventral hippocampus (Groenewegen & Steinbusch, 1984; Montone et al., 1988).

The chemical neuroanatomy of the medial and lateral habenula is rich with a considerable number of different neurotransmitters and neuropeptides, including somatostatin, substance P, neurotensin, serotonin, agmatine, parathyroid hormone, neuromedin, Leu-enkephalin, dopamine, acetylcholine, and urocortin III (Freed et al., 1995; Geisler, Andres, & Veh, 2003; Hervieu & Emson, 1998; Honzawa, Sudoh, Minamino, Tohyama, & Matsuo, 1987; Jennes, Stumpf, & Kalivas, 1982; Li, Vaughan,

Sawchenko, & Vale, 2002; Otake et al., 1998; Schafer, Eiden, & Weihe, 1998; Shinoda, Inagaki, Shiosaka, Kohno, & Tohyama, 1984; Shinoda & Tohyama, 1987; Wang, Palkovits, Rusnak, Mezey, & Usdin, 2000). In addition, there are at least two receptor types which are relatively specific to the habenula. First, the medial habenula is especially rich in several types of nicotinic receptors, including one with a selective affinity for epibatidine that appears uniquely expressed there and in the fasciculus retroflexus and IPN (Plenge, Mellerup, & Wortwein, 2002). Even more promising is the finding of an orphan G-protein-coupled receptor (GPCR-2037), the mRNA for which was exclusively expressed throughout both the medial and lateral habenula (Berthold, Collin, Sejlitz, Meister, & Lind, 2003). It is considered an "orphan" because a ligand for it has not yet been discovered. Further research elucidating the structure and function of this receptor could eventually provide a way of selectively targeting the habenula with a systemically administered drug, which could be of immense therapeutic utility.

CONCLUSIONS

This body of research has provided several unique insights. One is that noveltyseeking behaviors may be an early temperamental indicator of vulnerability to stressrelated psychopathology, and that novelty seeking, helplessness susceptibility, and reduced adrenal signaling may go hand in hand. Another is the suggestion that animals characterized by these behavioral and endocrine features have extensively altered brains even at birth, particularly in terms of how stress-responsive regions appear functionally disconnected. Finally, the habenula may be a nexus through which antidepressants may influence these dysfunctional neural networks, and its unique pharmacology offers great promise as a therapeutic target for treating emotional disorders in the future. Figure 7.1: Model of major circuitry disruption in adult congenitally helpless rats. Shaded regions are hypermetabolic, and unshaded regions are hypometabolic.



- Abercrombie, H. C., Schaefer, S. M., Larson, C. L., Oakes, T. R., Lindgren, K. A., Holden, J. E. et al. (1998). Metabolic rate in the right amygdala predicts negative affect in depressed patients. *Neuroreport*, 9, 3301-3307.
- Amaral, D. G. & Kurz, J. (1985). An analysis of the origins of the cholinergic and noncholinergic septal projections to the hippocampal formation of the rat. *The Journal of Comparative Neurology*, 240, 37-59.
- Amat, J., Sparks, P. D., Matus-Amat, P., Griggs, J., Watkins, L. R., & Maier, S. F. (2001). The role of the habenular complex in the elevation of dorsal raphe nucleus serotonin and the changes in the behavioral responses produced by uncontrollable stress. *Brain Research*, 917, 118-126.
- American Psychiatric Association (2000). Diagnostic and Statistical Manual of Mental Disorders. (Fourth Edition, Text Revision ed.) Washington, DC: American Psychiatric Press.
- Arvanitogiannis, A., Flores, C., Pfaus, J. G., & Shizgal, P. (1996). Increased ipsilateral expression of Fos following lateral hypothalamic self-stimulation. *Brain Research*, 720, 148-154.
- Austin, M. P., Dougall, N., Ross, M., Murray, C., O'Carroll, R. E., Moffoot, A. et al. (1992). Single photon emission tomography with 99mTc-exametazime in major depression and the pattern of brain activity underlying the psychotic/neurotic continuum. J Affect.Disord., 26, 31-43.
- Austin, M. P. & Mitchell, P. (1996). Melancholia as a neurological disorder. In G.Parker
 & D. Hadzi-Pavlovic (Eds.), *Melancholia: A disorder of movement and mood* (pp. 223-236). Cambridge: Cambridge University Press.
- Bacon, S. J. & Smith, A. D. (1993). A monosynaptic pathway from an identified vasomotor centre in the medial prefrontal cortex to an autonomic area in the thoracic spinal cord. *Neuroscience*, *54*, 719-728.
- Baker, D. G., West, S. A., Nicholson, W. E., Ekhator, N. N., Kasckow, J. W., Hill, K. K. et al. (1999). Serial CSF corticotropin-releasing hormone levels and adrenocortical activity in combat veterans with posttraumatic stress disorder. *The American Journal of Psychiatry*, 156, 585-588.
- Barrett, D., Shumake, J., Jones, D., & Gonzalez-Lima, F. (2003). Metabolic mapping of mouse brain activity after extinction of a conditioned emotional response. *The Journal of Neuroscience*, 23, 5740-5749.

- Baxter, L. R., Jr., Schwartz, J. M., Phelps, M. E., Mazziotta, J. C., Guze, B. H., Selin, C. E. et al. (1989a). Reduction of prefrontal cortex glucose metabolism common to three types of depression. *Archives of General Psychiatry*, 46, 243-250.
- Baxter, L. R., Jr., Schwartz, J. M., Phelps, M. E., Mazziotta, J. C., Guze, B. H., Selin, C. E. et al. (1989b). Reduction of prefrontal cortex glucose metabolism common to three types of depression. *Archives of General Psychiatry*, 46, 243-250.
- Benabid, A. L. & Jeaugey, L. (1989). Cells of the rat lateral habenula respond to high-threshold somatosensory inputs. *Neuroscience Letters*, *96*, 289-294.
- Bench, C. J., Friston, K. J., Brown, R. G., Frackowiak, R. S., & Dolan, R. J. (1993). Regional cerebral blood flow in depression measured by positron emission tomography: the relationship with clinical dimensions. *Psychological Medicine*, 23, 579-590.
- Bench, C. J., Friston, K. J., Brown, R. G., Scott, L. C., Frackowiak, R. S., & Dolan, R. J. (1992). The anatomy of melancholia--focal abnormalities of cerebral blood flow in major depression. *Psychological Medicine*, 22, 607-615.
- Berthold, M., Collin, M., Sejlitz, T., Meister, B., & Lind, P. (2003). Cloning of a novel orphan G protein-coupled receptor (GPCR-2037): in situ hybridization reveals high mRNA expression in rat brain restricted to neurons of the habenular complex. *Brain Research.Molecular Brain Research*, *120*, 22-29.
- Bielajew, C. H. (1991). Distribution of cytochrome oxidase in response to rewarding brain stimulation: effect of different pulse durations. *Brain Research Bulletin, 26,* 379-384.
- Biver, F., Goldman, S., Delvenne, V., Luxen, A., De, M., V, Hubain, P. et al. (1994). Frontal and parietal metabolic disturbances in unipolar depression. *Biological Psychiatry*, 36, 381-388.
- Blier, P. & de Montigny, C. (1999). Serotonin and drug-induced therapeutic responses in major depression, obsessive-compulsive and panic disorders. *Neuropsychopharmacology, 21,* 91S-98S.
- Bonne, O., Gilboa, A., Louzoun, Y., Brandes, D., Yona, I., Lester, H. et al. (2003). Resting regional cerebral perfusion in recent posttraumatic stress disorder. *Biological Psychiatry*, 54, 1077-1086.
- Boyd, E. S. & Celso, M. B. (1970). Effect of some brain lesions on septal intracranial self-stimulation in the rat. *The American Journal of Physiology*, *219*, 734-741.

- Breiter, H. C. & Rosen, B. R. (1999). Functional magnetic resonance imaging of brain reward circuitry in the human. Annals of the New York Academy of Sciences, 877, 523-547.
- Bremner, J. D., Innis, R. B., Salomon, R. M., Staib, L. H., Ng, C. K., Miller, H. L. et al. (1997a). Positron emission tomography measurement of cerebral metabolic correlates of tryptophan depletion-induced depressive relapse. *Archives of General Psychiatry*, 54, 364-374.
- Bremner, J. D., Narayan, M., Anderson, E. R., Staib, L. H., Miller, H. L., & Charney, D. S. (2000). Hippocampal volume reduction in major depression. *American Journal of Psychiatry*, 157, 115-118.
- Bremner, J. D., Narayan, M., Staib, L. H., Southwick, S. M., McGlashan, T., & Charney, D. S. (1999a). Neural correlates of memories of childhood sexual abuse in women with and without posttraumatic stress disorder. *American Journal of Psychiatry*, 156, 1787-1795.
- Bremner, J. D., Randall, P., Vermetten, E., Staib, L., Bronen, R. A., Mazure, C. et al. (1997b). Magnetic resonance imaging-based measurement of hippocampal volume in posttraumatic stress disorder related to childhood physical and sexual abuse--a preliminary report. *Biological Psychiatry*, 41, 23-32.
- Bremner, J. D., Staib, L. H., Kaloupek, D., Southwick, S. M., Soufer, R., & Charney, D. S. (1999b). Neural correlates of exposure to traumatic pictures and sound in Vietnam combat veterans with and without posttraumatic stress disorder: a positron emission tomography study. *Biological Psychiatry*, 45, 806-816.
- Brody, A. L., Saxena, S., Silverman, D. H., Alborzian, S., Fairbanks, L. A., Phelps, M. E. et al. (1999). Brain metabolic changes in major depressive disorder from pre- to post-treatment with paroxetine. *Psychiatry Research*, 91, 127-139.
- Brown, G. M., Uhlir, I. V., Seggie, J., Schally, A. V., & Kastin, A. J. (1974). Effect of septal lesions on plasma levels of MSH, corticosterone, GH and prolactin before and after exposure to novel environment: role of MSH in the septal syndrome. *Endocrinology*, 94, 593-597.
- Buchsbaum, M. S., Wu, J., DeLisi, L. E., Holcomb, H., Kessler, R., Johnson, J. et al. (1986). Frontal cortex and basal ganglia metabolic rates assessed by positron emission tomography with [18F]2-deoxyglucose in affective illness. J Affect.Disord., 10, 137-152.
- Buchsbaum, M. S., Wu, J., Siegel, B. V., Hackett, E., Trenary, M., Abel, L. et al. (1997). Effect of sertraline on regional metabolic rate in patients with affective disorder. *Biological Psychiatry*, 41, 15-22.

- Cada, A., Gonzalez-Lima, F., Rose, G. M., & Bennett, M. C. (1995). Regional brain effects of sodium azide treatment on cytochrome oxidase activity: a quantitative histochemical study. *Metabolic Brain Disease*, *10*, 303-320.
- Caldecott-Hazard, S., Mazziotta, J., & Phelps, M. (1988). Cerebral correlates of depressed behavior in rats, visualized using 14C- 2-deoxyglucose autoradiography. *The Journal of Neuroscience*, *8*, 1951-1961.
- Calfa, G., Kademian, S., Ceschin, D., Vega, G., Rabinovich, G. A., & Volosin, M. (2003). Characterization and functional significance of glucocorticoid receptors in patients with major depression: modulation by antidepressant treatment. *Psychoneuroendocrinology, 28,* 687-701.
- Canteras, N. S. & Swanson, L. W. (1992). Projections of the ventral subiculum to the amygdala, septum, and hypothalamus: a PHAL anterograde tract-tracing study in the rat. *The Journal of Comparative Neurology, 324,* 180-194.
- Cazala, P., Galey, D., & Durkin, T. (1988). Electrical self-stimulation in the medial and lateral septum as compared to the lateral hypothalamus: differential intervention of reward and learning processes? *Physiology & Behavior, 44,* 53-59.
- Charles, H. C., Lazeyras, F., Krishnan, K. R., Boyko, O. B., Payne, M., & Moore, D. (1994). Brain choline in depression: in vivo detection of potential pharmacodynamic effects of antidepressant therapy using hydrogen localized spectroscopy. *Progress in Neuro-psychopharmacology & Biological Psychiatry*, 18, 1121-1127.
- Chen, A. C., Shirayama, Y., Shin, K. H., Neve, R. L., & Duman, R. S. (2001). Expression of the cAMP response element binding protein (CREB) in hippocampus produces an antidepressant effect. *Biological Psychiatry*, *49*, 753-762.
- Christoph, G. R., Leonzio, R. J., & Wilcox, K. S. (1986). Stimulation of the lateral habenula inhibits dopamine-containing neurons in the substantia nigra and ventral tegmental area of the rat. *The Journal of Neuroscience*, *6*, 613-619.
- Cloninger, C. R. (1987). A systematic method for clinical description and classification of personality variants. A proposal. *Archives of General Psychiatry*, 44, 573-588.
- Coffey, C. E., Wilkinson, W. E., Weiner, R. D., Parashos, I. A., Djang, W. T., Webb, M. C. et al. (1993). Quantitative cerebral anatomy in depression. A controlled magnetic resonance imaging study. *Archives of General Psychiatry*, 50, 7-16.
- Contestabile, A. & Flumerfelt, B. A. (1981). Afferent connections of the interpeduncular nucleus and the topographic organization of the habenulo-interpeduncular pathway: an HRP study in the rat. *The Journal of Comparative Neurology, 196,* 253-270.

- Contreras, C. M., Alcala-Herrera, V., & Marvan, M. L. (1989). Action of antidepressants on the septal nuclei of the rat. *Physiology & Behavior, 46,* 793-798.
- Cullinan, W. E., Herman, J. P., & Watson, S. J. (1993). Ventral subicular interaction with the hypothalamic paraventricular nucleus: evidence for a relay in the bed nucleus of the stria terminalis. *The Journal of Comparative Neurology, 332*, 1-20.
- Damasio, A. R., Tranel, D., & Damasio, H. (1990). Individuals with sociopathic behavior caused by frontal damage fail to respond autonomically to social stimuli. *Behavioural Brain Research*, 41, 81-94.
- Davidson, R. J., Lewis, D. A., Alloy, L. B., Amaral, D. G., Bush, G., Cohen, J. D. et al. (2002). Neural and behavioral substrates of mood and mood regulation. *Biological Psychiatry*, 52, 478-502.
- de Kloet, E. R. (2004). Hormones and the stressed brain. Annals of the New York Academy of Sciences, 1018, 1-15.
- Delahanty, D. L., Raimonde, A. J., & Spoonster, E. (2000). Initial posttraumatic urinary cortisol levels predict subsequent PTSD symptoms in motor vehicle accident victims. *Biological Psychiatry*, 48, 940-947.
- Detke, M. J., Rickels, M., & Lucki, I. (1995). Active behaviors in the rat forced swimming test differentially produced by serotonergic and noradrenergic antidepressants. *Psychopharmacology*, 121, 66-72.
- Devous, M. D., Sr., Trivedi, M. H., & Rush, A. J. (2001). Regional cerebral blood flow response to oral amphetamine challenge in healthy volunteers. *Journal of Nuclear Medicine*, 42, 535-542.
- Di Rocco, R. J., Kageyama, G. H., & Wong-Riley, M. T. (1989). The relationship between CNS metabolism and cytoarchitecture: a review of 14C-deoxyglucose studies with correlation to cytochrome oxidase histochemistry. *Computerized Medical Imaging and Graphics*, 13, 81-92.
- Dilsaver, S. C. & Coffman, J. A. (1989). Cholinergic hypothesis of depression: a reappraisal. *Journal of Clinical Psychopharmacology*, *9*, 173-179.
- Drevets, W. C. (2000). Neuroimaging studies of mood disorders. *Biological Psychiatry*, 48, 813-829.
- Drevets, W. C., Price, J. L., Simpson, J. R., Jr., Todd, R. D., Reich, T., Vannier, M. et al. (1997). Subgenual prefrontal cortex abnormalities in mood disorders. *Nature*, *386*, 824-827.

- Drevets, W. C., Videen, T. O., Price, J. L., Preskorn, S. H., Carmichael, S. T., & Raichle, M. E. (1992). A functional anatomical study of unipolar depression. *The Journal* of Neuroscience, 12, 3628-3641.
- Dube, S. (1993). Cholinergic supersensitivity in affective disorders. In J.J.Mann & D. J. Kupfer (Eds.), *Biology of depressive disorders, Part A: a systems perspective* (pp. 51-78). New York: Plenum.
- Dudar, J. D. (1977). The role of the septal nuclei in the release of acetyl-choline from the rabbit cerebral cortex and dorsal hippocampus and the effect of atropine. *Brain Research*, 129, 237-246.
- Dudar, J. D., Whishaw, I. Q., & Szerb, J. C. (1979). Release of acetylcholine from the hippocampus of freely moving rats during sensory stimulation and running. *Neuropharmacology*, 18, 673-678.
- Duman, R. S., Malberg, J., & Thome, J. (1999). Neural plasticity to stress and antidepressant treatment. *Biological Psychiatry*, 46, 1181-1191.
- Dunn, R. T., Kimbrell, T. A., Ketter, T. A., Frye, M. A., Willis, M. W., Luckenbaugh, D. A. et al. (2002). Principal components of the Beck Depression Inventory and regional cerebral metabolism in unipolar and bipolar depression. *Biological Psychiatry*, 51, 387-399.
- Ebert, D., Feistel, H., & Barocka, A. (1991). Effects of sleep deprivation on the limbic system and the frontal lobes in affective disorders: a study with Tc-99m-HMPAO SPECT. *Psychiatry Research, 40,* 247-251.
- Edwards, E., King, J. A., & Fray, J. (2000). Hypertension and insulin resistant models have divergent propensities to learned helpless behavior in rodents. *American Journal of Hypertension: Journal of the American Society of Hypertension, 13,* 659-665.
- Edwards, E., King, J. A., & Fray, J. C. (1999). Increased basal activity of the HPA axis and renin-angiotensin system in congenital learned helpless rats exposed to stress early in development. *Int.J Dev.Neurosci.*, 17, 805-812.
- Endo, Y., Nishimura, J., & Kimura, F. (1994). Adrenalectomy increases local cerebral blood flow in the rat hippocampus. *Pflugers Arch.*, 426, 183-188.
- Endo, Y., Nishimura, J. I., Kobayashi, S., & Kimura, F. (1997). Long-term glucocorticoid treatments decrease local cerebral blood flow in the rat hippocampus, in association with histological damage. *Neuroscience*, *79*, 745-752.
- Esposito, E. (1996). An indirect action for fluoxetine on the dopamine neurotransmitter system. *Trends Pharmacol.Sci.*, 17, 400-402.

- Esposito, R. U., Porrino, L. J., Seeger, T. F., Crane, A. M., Everist, H. D., & Pert, A. (1984). Changes in local cerebral glucose utilization during rewarding brain stimulation. *Proceedings of the National Academy of Sciences of the United States* of America, 81, 635-639.
- Everitt, B. J., Parkinson, J. A., Olmstead, M. C., Arroyo, M., Robledo, P., & Robbins, T. W. (1999). Associative processes in addiction and reward. The role of amygdalaventral striatal subsystems. *Annals of the New York Academy of Sciences*, 877, 412-438.
- Fallon, J. H. & Ciofi, P. (1992). Distribution of monoamines within the amygdala. In J.P.Aggleton (Ed.), *The Amygdala: Neurobiological Aspects of Emotion, Memory,* and Mental Dysfunction (pp. 97-114). New York: Wiley-Liss.
- Fava, M. & Kendler, K. S. (2000). Major depressive disorder. Neuron, 28, 335-341.
- Felton, T. M., Linton, L., Rosenblatt, J. S., & Morell, J. I. (1999). First and second order maternal behavior related afferents of the lateral habenula. *Neuroreport*, 10, 883-887.
- Ferraro, G., Montalbano, M. E., Sardo, P., & LaGrutta, V. (1997). Lateral habenula and hippocampus: a complex interaction raphe cells- mediated. *Journal of Neural Transmission*, 104, 615-631.
- Flicker, C. & Geyer, M. A. (1982). Behavior during hippocampal microinfusions. II. Muscarinic locomotor activation. *Brain Research*, 257, 105-127.
- Foa, E. B., Zinbarg, R., & Rothbaum, B. O. (1992). Uncontrollability and unpredictability in post-traumatic stress disorder: an animal model. *Psychological Bulletin*, 112, 218-238.
- Fredriksen, K., Rhodes, J., Reddy, R., & Way, N. (2004). Sleepless in Chicago: tracking the effects of adolescent sleep loss during the middle school years. *Child Dev.*, 75, 84-95.
- Freed, C., Revay, R., Vaughan, R. A., Kriek, E., Grant, S., Uhl, G. R. et al. (1995). Dopamine transporter immunoreactivity in rat brain. *J Comp Neurol.*, 359, 340-349.
- Freo, U., Ori, C., Dam, M., Merico, A., & Pizzolato, G. (2000). Effects of acute and chronic treatment with fluoxetine on regional glucose cerebral metabolism in rats: implications for clinical therapies. *Brain Research*, *854*, 35-41.
- Friston, K. J. (1994). Functional and effective connectivity in neuroimaging: a synthesis. *Human Brain Mapping, 2,* 56-78.

- Frysztak, R. J. & Neafsey, E. J. (1991). The effect of medial frontal cortex lesions on respiration, "freezing," and ultrasonic vocalizations during conditioned emotional responses in rats. *Cerebral Cortex*, 1, 418-425.
- Fuchs, P. & Cox, V. C. (1993). Habenula lesions attenuate lateral hypothalamic analgesia in the formalin test. *Neuroreport, 4*, 121-124.
- Gallistel, C. R., Gomita, Y., Yadin, E., & Campbell, K. A. (1985). Forebrain origins and terminations of the medial forebrain bundle metabolically activated by rewarding stimulation or by reward-blocking doses of pimozide. *The Journal of Neuroscience*, *5*, 1246-1261.
- Geisler, S., Andres, K. H., & Veh, R. W. (2003). Morphologic and cytochemical criteria for the identification and delineation of individual subnuclei within the lateral habenular complex of the rat. *J Comp Neurol.*, *458*, 78-97.
- Gilbertson, M. W., Shenton, M. E., Ciszewski, A., Kasai, K., Lasko, N. B., Orr, S. P. et al. (2002). Smaller hippocampal volume predicts pathologic vulnerability to psychological trauma. *Nat.Neurosci.*, *5*, 1242-1247.
- Gilboa, A., Shalev, A. Y., Laor, L., Lester, H., Louzoun, Y., Chisin, R. et al. (2004). Functional connectivity of the prefrontal cortex and the amygdala in posttraumatic stress disorder. *Biological Psychiatry*, *55*, 263-272.
- Gomita, Y. & Gallistel, C. R. (1982). Effects of reinforcement-blocking doses of pimozide on neural systems driven by rewarding stimulation of the MFB: a 14C-2-deoxyglucose analysis. *Pharmacology, biochemistry, and Behavior, 17,* 841-845.
- Gonzalez-Lima, F. (1992). Brain imaging of auditory learning functions in rats: Studies with fluorodeoxyglucose autorediography and cytochrome oxidase histochemistry. In F.Gonzalez-Lima, Th. Finkenstädt, & H. Scheich (Eds.), Advances in metabolic mapping techniques for brain imaging of behavioral and learning functions (pp. 39-110). Dordrecht, The Netherlands: Kluwer Academic Publishers.
- Gonzalez-Lima, F. & Cada, A. (1994). Cytochrome oxidase activity in the auditory system of the mouse: a qualitative and quantitative histochemical study. *Neuroscience*, 63, 559-578.
- Gonzalez-Lima, F. & Cada, A. (1998a). Cytochrome oxidase atlas of rat brain. In F.Gonzalez-Lima (Ed.), *Cytochrome oxidase in neuronal metabolism and Alzheimer's disease* (pp. 263-280). New York: Plenum Press.

- Gonzalez-Lima, F. & Cada, A. (1998b). Quantitative histochemistry of cytochrome oxidase activity. In F.Gonzalez-Lima (Ed.), Cytochrome oxidase in neuronal metabolism and Alzheimer's disease (pp. 55-90). New York: Plenum Press.
- Gonzalez-Lima, F. & Garrosa, M. (1991). Quantitative histochemistry of cytochrome oxidase in rat brain. *Neuroscience Letters*, 123, 251-253.
- Gonzalez-Lima, F. & Sadile, A. G. (2000). Network operations revealed by brain metabolic mapping in a genetic model of hyperactivity and attention deficit: the naples high- and low- excitability rats. *Neuroscience and Biobehavioral Reviews*, 24, 157-160.
- Gonzalez-Lima, F. & Scheich, H. (1985). Ascending reticular activating system in the rat: a 2-deoxyglucose study. *Brain Research*, 344, 70-88.
- Grahn, R. E., Hammack, S. E., Will, M. J., O'Connor, K. A., Deak, T., Sparks, P. D. et al. (2002). Blockade of alpha1 adrenoreceptors in the dorsal raphe nucleus prevents enhanced conditioned fear and impaired escape performance following uncontrollable stressor exposure in rats. *Behavioural Brain Research*, 134, 387-392.
- Grahn, R. E., Maswood, S., McQueen, M. B., Watkins, L. R., & Maier, S. F. (1999a). Opioid-dependent effects of inescapable shock on escape behavior and conditioned fear responding are mediated by the dorsal raphe nucleus. *Behavioural Brain Research*, 99, 153-167.
- Grahn, R. E., Watkins, L. R., & Maier, S. F. (2000). Impaired escape performance and enhanced conditioned fear in rats following exposure to an uncontrollable stressor are mediated by glutamate and nitric oxide in the dorsal raphe nucleus. *Behavioural Brain Research*, 112, 33-41.
- Grahn, R. E., Will, M. J., Hammack, S. E., Maswood, S., McQueen, M. B., Watkins, L. R. et al. (1999b). Activation of serotonin-immunoreactive cells in the dorsal raphe nucleus in rats exposed to an uncontrollable stressor. *Brain Research*, 826, 35-43.
- Greenwood, B. N., Foley, T. E., Day, H. E., Campisi, J., Hammack, S. H., Campeau, S. et al. (2003). Freewheel running prevents learned helplessness/behavioral depression: role of dorsal raphe serotonergic neurons. *The Journal of Neuroscience*, 23, 2889-2898.
- Groenewegen, H. J. & Steinbusch, H. W. (1984). Serotonergic and non-serotonergic projections from the interpeduncular nucleus to the ventral hippocampus in the rat. *Neuroscience Letters*, *51*, 19-24.

- Gross-Isseroff, R., Dillon, K. A., Israeli, M., & Biegon, A. (1990). Regionally selective increases in mu opioid receptor density in the brains of suicide victims. *Brain Research*, *530*, 312-316.
- Gurvits, T. V., Shenton, M. E., Hokama, H., Ohta, H., Lasko, N. B., Gilbertson, M. W. et al. (1996). Magnetic resonance imaging study of hippocampal volume in chronic, combat-related posttraumatic stress disorder. *Biological Psychiatry*, 40, 1091-1099.
- Hammack, S. E., Pepin, J. L., DesMarteau, J. S., Watkins, L. R., & Maier, S. F. (2003). Low doses of corticotropin-releasing hormone injected into the dorsal raphe nucleus block the behavioral consequences of uncontrollable stress. *Behavioural Brain Research*, 147, 55-64.
- Hammack, S. E., Richey, K. J., Schmid, M. J., LoPresti, M. L., Watkins, L. R., & Maier, S. F. (2002). The role of corticotropin-releasing hormone in the dorsal raphe nucleus in mediating the behavioral consequences of uncontrollable stress. *The Journal of Neuroscience*, 22, 1020-1026.
- Hammack, S. E., Richey, K. J., Watkins, L. R., & Maier, S. F. (2004). Chemical lesion of the bed nucleus of the stria terminalis blocks the behavioral consequences of uncontrollable stress. *Behavioral Neuroscience*, 118, 443-448.
- Hammack, S. E., Schmid, M. J., LoPresti, M. L., Der-Avakian, A., Pellymounter, M. A., Foster, A. C. et al. (2003). Corticotropin releasing hormone type 2 receptors in the dorsal raphe nucleus mediate the behavioral consequences of uncontrollable stress. *The Journal of Neuroscience, 23*, 1019-1025.
- Hammer, R. P., Jr., Pires, W. S., Markou, A., & Koob, G. F. (1993). Withdrawal following cocaine self-administration decreases regional cerebral metabolic rate in critical brain reward regions. *Synapse*, 14, 73-80.
- Haun, F., Eckenrode, T. C., & Murray, M. (1992). Habenula and thalamus cell transplants restore normal sleep behaviors disrupted by denervation of the interpeduncular nucleus. *The Journal of Neuroscience*, *12*, 3282-3290.
- Henn, F. A. & Edwards, E. (1994). Animal models in the study of genetic factors in human psychopathology. In D.F.Papolos & H. M. Lachman (Eds.), Genetic studies in affective disorders: Overview of basic methods, current directions, and critical research issues (pp. 177-192). New York: John Wiley and Sons.
- Henn, F. A., Johnson, J., Edwards, E., & Anderson, D. (1985). Melancholia in rodents: neurobiology and pharmacology. *Psychopharmacology Bulletin, 21*, 443-446.

- Herman, J. P., Dolgas, C. M., & Carlson, S. L. (1998). Ventral subiculum regulates hypothalamo-pituitary-adrenocortical and behavioural responses to cognitive stressors. *Neuroscience*, *86*, 449-459.
- Herman, J. P., Figueiredo, H., Mueller, N. K., Ulrich-Lai, Y., Ostrander, M. M., Choi, D. C. et al. (2003). Central mechanisms of stress integration: hierarchical circuitry controlling hypothalamo-pituitary-adrenocortical responsiveness. *Frontiers in Neuroendocrinology*, 24, 151-180.
- Hervieu, G. & Emson, P. C. (1998). The localization of somatostatin receptor 1 (sst1) immunoreactivity in the rat brain using an N-terminal specific antibody. *Neuroscience*, *85*, 1263-1284.
- Hickie, I. B., Scott, E. M., & Davenport, T. A. (1999). Are antidepressants all the same? Surveying the opinions of Australian psychiatrists. *Aust.N.Z.J Psychiatry*, 33, 642-649.
- Ho, A. P., Gillin, J. C., Buchsbaum, M. S., Wu, J. C., Abel, L., & Bunney, W. E., Jr. (1996). Brain glucose metabolism during non-rapid eye movement sleep in major depression. A positron emission tomography study. *Archives of General Psychiatry*, 53, 645-652.
- Holsboer, F., Liebl, R., & Hofschuster, E. (1982). Repeated dexamethasone suppression test during depressive illness. Normalisation of test result compared with clinical improvement. *Journal of Affective Disorders*, 4, 93-101.
- Honzawa, M., Sudoh, T., Minamino, N., Tohyama, M., & Matsuo, H. (1987). Topographic localization of neuromedin U-like structures in the rat brain: an immunohistochemical study. *Neuroscience*, 23, 1103-1122.
- Hooks, M. S., Jones, G. H., Smith, A. D., Neill, D. B., & Justice, J. B., Jr. (1991). Response to novelty predicts the locomotor and nucleus accumbens dopamine response to cocaine. *Synapse*, 9, 121-128.
- Horner, H. C., Packan, D. R., & Sapolsky, R. M. (1990). Glucocorticoids inhibit glucose transport in cultured hippocampal neurons and glia. *Neuroendocrinology*, 52, 57-64.
- Horwitz, B. (1994). Data analysis paradigms for metabolic-flow data: combining neural modeling and functional neuroimaging. *Human Brain Mapping*, *2*, 112-122.
- Hyman, S. E. & Nestler, E. J. (1996). Initiation and adaptation: a paradigm for understanding psychotropic drug action. *American Journal of Psychiatry*, 153, 151-162.

- Ifft, J. D. (1972). An autoradiographic study of the time of final division of neurons in rat hypothalamic nuclei. *The Journal of Comparative Neurology*, *144*, 193-204.
- Imaki, T., Shibasaki, T., Hotta, M., & Demura, H. (1993). Intracerebroventricular administration of corticotropin-releasing factor induces c-fos mRNA expression in brain regions related to stress responses: comparison with pattern of c-fos mRNA induction after stress. *Brain Research*, *616*, 114-125.
- Janowsky, D. S., el Yousef, M. K., Davis, J. M., & Sekerke, H. J. (1972). A cholinergicadrenergic hypothesis of mania and depression. *Lancet*, *2*, 632-635.
- Janowsky, D. S., Risch, S. C., & Gillin, J. C. (1983). Adrenergic-cholinergic balance and the treatment of affective disorders. *Progress in Neuro-psychopharmacology & Biological Psychiatry*, 7, 297-307.
- Jennes, L., Stumpf, W. E., & Kalivas, P. W. (1982). Neurotensin: topographical distribution in rat brain by immunohistochemistry. *J Comp Neurol.*, 210, 211-224.
- Kabbaj, M. (2004). Neurobiological bases of individual differences in emotional and stress responsiveness: high responders-low responders model. *Arch.Neurol.*, *61*, 1009-1012.
- Kabbaj, M. & Akil, H. (2001). Individual differences in novelty-seeking behavior in rats: a c-fos study. *Neuroscience*, *106*, 535-545.
- Kabbaj, M., Devine, D. P., Savage, V. R., & Akil, H. (2000). Neurobiological correlates of individual differences in novelty-seeking behavior in the rat: differential expression of stress-related molecules. *J Neurosci.*, 20, 6983-6988.
- Kalen, P., Karlson, M., & Wiklund, L. (1985). Possible excitatory amino acid afferents to nucleus raphe dorsalis of the rat investigated with retrograde wheat germ agglutinin and D- [3H]aspartate tracing. *Brain Research*, 360, 285-297.
- Kalen, P., Pritzel, M., Nieoullon, A., & Wiklund, L. (1986). Further evidence for excitatory amino acid transmission in the lateral habenular projection to the rostral raphe nuclei: lesion-induced decrease of high affinity glutamate uptake. *Neuroscience Letters*, 68, 35-40.
- Kendler, K. S., Karkowski, L. M., & Prescott, C. A. (1999). Causal relationship between stressful life events and the onset of major depression. *The American Journal of Psychiatry*, 156, 837-841.
- Kendler, K. S., Kessler, R. C., Walters, E. E., MacLean, C., Neale, M. C., Heath, A. C. et al. (1995). Stressful life events, genetic liability, and onset of an episode of major depression in women. *American Journal of Psychiatry*, 152, 833-842.

- Kessler, R. C., Sonnega, A., Bromet, E., Hughes, M., & Nelson, C. B. (1995). Posttraumatic stress disorder in the National Comorbidity Survey. Archives of General Psychiatry, 52, 1048-1060.
- Kimbrell, T. A., Ketter, T. A., George, M. S., Little, J. T., Benson, B. E., Willis, M. W. et al. (2002). Regional cerebral glucose utilization in patients with a range of severities of unipolar depression. *Biological Psychiatry*, 51, 237-252.
- King, J. A., Abend, S., & Edwards, E. (2001). Genetic predisposition and the development of posttraumatic stress disorder in an animal model. *Biological Psychiatry*, 50, 231-237.
- King, J. A., Campbell, D., & Edwards, E. (1993). Differential development of the stress response in congenital learned helplessness. *International Journal of Developmental Neuroscience*, 11, 435-442.
- King, J. A. & Edwards, E. (1999). Early stress and genetic influences on hypothalamicpituitary-adrenal axis functioning in adulthood. *Hormones and Behavior*, 36, 79-85.
- Klimek, V., Schenck, J., Han, H., Stockmeier, C., & Ordway, G. (2002). Dopaminergic abnormalities in amygdaloid nuclei in major depression: a postmortem study. *Biological Psychiatry*, *52*, 740.
- Kohen, R., Neumaier, J. F., Hamblin, M. W., & Edwards, E. (2003). Congenitally learned helpless rats show abnormalities in intracellular signaling. *Biological Psychiatry*, 53, 520-529.
- Kozlowski, M. R. & Marshall, J. F. (1980). Plasticity of [14C]2-deoxy-D-glucose incorporation into neostriatum and related structures in response to dopamine neuron damage and apomorphine replacement. *Brain Research*, 197, 167-183.
- Krettek, J. E. & Price, J. L. (1977). The cortical projections of the mediodorsal nucleus and adjacent thalamic nuclei in the rat. *The Journal of Comparative Neurology*, *171*, 157-191.
- Krishnan, K. R., Doraiswamy, P. M., Figiel, G. S., Husain, M. M., Shah, S. A., Na, C. et al. (1991a). Hippocampal abnormalities in depression. *J Neuropsychiatry Clin.Neurosci.*, 3, 387-391.
- Krishnan, K. R., Doraiswamy, P. M., Lurie, S. N., Figiel, G. S., Husain, M. M., Boyko, O. B. et al. (1991b). Pituitary size in depression. *The Journal of Clinical Endocrinology and Metabolism*, 72, 256-259.
- Krishnan, K. R., McDonald, W. M., Escalona, P. R., Doraiswamy, P. M., Na, C., Husain, M. M. et al. (1992). Magnetic resonance imaging of the caudate nuclei in

depression. Preliminary observations. Archives of General Psychiatry, 49, 553-557.

- Lacerda, A. L., Nicoletti, M. A., Brambilla, P., Sassi, R. B., Mallinger, A. G., Frank, E. et al. (2003). Anatomical MRI study of basal ganglia in major depressive disorder. *Psychiatry Research*, 124, 129-140.
- Lachman, H. M., Papolos, D. F., Boyle, A., Sheftel, G., Juthani, M., Edwards, E. et al. (1993). Alterations in glucocorticoid inducible RNAs in the limbic system of learned helpless rats. *Brain Research*, 609, 110-116.
- Lachman, H. M., Papolos, D. F., Weiner, E. D., Ramazankhana, R., Hartnick, C., Edwards, E. et al. (1992). Hippocampal neuropeptide Y mRNA is reduced in a strain of learned helpless resistant rats. *Brain Research.Molecular Brain Research*, 14, 94-100.
- Lanius, R. A., Williamson, P. C., Boksman, K., Densmore, M., Gupta, M., Neufeld, R. W. et al. (2002). Brain activation during script-driven imagery induced dissociative responses in PTSD: a functional magnetic resonance imaging investigation. *Biological Psychiatry*, 52, 305-311.
- Lanius, R. A., Williamson, P. C., Densmore, M., Boksman, K., Gupta, M. A., Neufeld, R. W. et al. (2001). Neural correlates of traumatic memories in posttraumatic stress disorder: a functional MRI investigation. *American Journal of Psychiatry*, 158, 1920-1922.
- Lee, E. H. & Huang, S. L. (1988). Role of lateral habenula in the regulation of exploratory behavior and its relationship to stress in rats. *Behavioural Brain Research*, 30, 265-271.
- Lee, E. H., Lin, Y. P., & Yin, T. H. (1988). Effects of lateral and medial septal lesions on various activity and reactivity measures in rats. *Physiology & Behavior*, 42, 97-102.
- Lee, R. K. & Maier, S. F. (1988). Inescapable shock and attention to internal versus external cues in a water discrimination escape task. *Journal of Experimental Psychology.Animal Behavior Processes*, 14, 302-310.
- Leshner, A. I. & Segal, M. (1979). Fornix transection blocks "learned helplessness" in rats. *Behavioral and Neural Biology*, 26, 497-501.
- Li, C., Vaughan, J., Sawchenko, P. E., & Vale, W. W. (2002). Urocortin IIIimmunoreactive projections in rat brain: partial overlap with sites of type 2 corticotrophin-releasing factor receptor expression. *J Neurosci.*, 22, 991-1001.

- Liberzon, I., Taylor, S. F., Amdur, R., Jung, T. D., Chamberlain, K. R., Minoshima, S. et al. (1999). Brain activation in PTSD in response to trauma-related stimuli. *Biological Psychiatry*, 45, 817-826.
- Liotti, M., Mayberg, H. S., Brannan, S. K., McGinnis, S., Jerabek, P., & Fox, P. T. (2000). Differential limbic--cortical correlates of sadness and anxiety in healthy subjects: implications for affective disorders. *Biological Psychiatry*, *48*, 30-42.
- Lisoprawski, A., Herve, D., Blanc, G., Glowinski, J., & Tassin, J. P. (1980). Selective activation of the mesocortico-frontal dopaminergic neurons induced by lesion of the habenula in the rat. *Brain Research*, *183*, 229-234.
- Lithgow, T. & Barr, G. A. (1984). Self-stimulation in 7- and 10-day-old rats. *Behavioral Neuroscience*, *98*, 479-486.
- Lucey, J. V., Costa, D. C., Adshead, G., Deahl, M., Busatto, G., Gacinovic, S. et al. (1997). Brain blood flow in anxiety disorders. OCD, panic disorder with agoraphobia, and post-traumatic stress disorder on 99mTcHMPAO single photon emission tomography (SPET). *Br.J Psychiatry*, 171, 346-350.
- Maier, S. F. (1984). Learned helplessness and animal models of depression. *Progress in Neuro-psychopharmacology & Biological Psychiatry*, 8, 435-446.
- Maier, S. F., Grahn, R. E., Kalman, B. A., Sutton, L. C., Wiertelak, E. P., & Watkins, L. R. (1993). The role of the amygdala and dorsal raphe nucleus in mediating the behavioral consequences of inescapable shock. *Behavioral Neuroscience*, 107, 377-388.
- Maier, S. F., Grahn, R. E., & Watkins, L. R. (1995). 8-OH-DPAT microinjected in the region of the dorsal raphe nucleus blocks and reverses the enhancement of fear conditioning and interference with escape produced by exposure to inescapable shock. *Behavioral Neuroscience*, 109, 404-412.
- Maier, S. F., Kalman, B. A., & Grahn, R. E. (1994). Chlordiazepoxide microinjected into the region of the dorsal raphe nucleus eliminates the interference with escape responding produced by inescapable shock whether administered before inescapable shock or escape testing. *Behavioral Neuroscience*, *108*, 121-130.
- Maswood, S., Barter, J. E., Watkins, L. R., & Maier, S. F. (1998). Exposure to inescapable but not escapable shock increases extracellular levels of 5-HT in the dorsal raphe nucleus of the rat. *Brain Research*, 783, 115-120.
- Matsuda, Y. & Fujimura, K. (1992). Action of habenular efferents on ventral tegmental area neurons studied in vitro. *Brain Research Bulletin, 28,* 743-749.

- Mayberg, H. S. (1997). Limbic-cortical dysregulation: a proposed model of depression. *The Journal of Neuropsychiatry and Clinical Neurosciences, 9,* 471-481.
- Mayberg, H. S., Brannan, S. K., Mahurin, R. K., Jerabek, P. A., Brickman, J. S., Tekell, J. L. et al. (1997). Cingulate function in depression: a potential predictor of treatment response. *Neuroreport*, 8, 1057-1061.
- Mayberg, H. S., Brannan, S. K., Tekell, J. L., Silva, J. A., Mahurin, R. K., McGinnis, S. et al. (2000). Regional metabolic effects of fluoxetine in major depression: serial changes and relationship to clinical response. *Biological Psychiatry*, 48, 830-843.
- Mayberg, H. S., Lewis, P. J., Regenold, W., & Wagner, H. N., Jr. (1994). Paralimbic hypoperfusion in unipolar depression. *J Nucl.Med.*, 35, 929-934.
- Mayberg, H. S., Liotti, M., Brannan, S. K., McGinnis, S., Mahurin, R. K., Jerabek, P. A. et al. (1999). Reciprocal limbic-cortical function and negative mood: converging PET findings in depression and normal sadness. *The American Journal of Psychiatry*, 156, 675-682.
- Mayeux, R., Stern, Y., Rosen, J., & Leventhal, J. (1981). Depression, intellectual impairment, and Parkinson disease. *Neurology*, *31*, 645-650.
- McCleery, J. M. & Goodwin, G. M. (2001). High and low neuroticism predict different cortisol responses to the combined dexamethasone-CRH test. *Biological Psychiatry*, 49, 410-415.
- McEwen, B. S., Weiss, J. M., & Schwartz, L. S. (1968). Selective retention of corticosterone by limbic structures in rat brain. *Nature*, 220, 911-912.
- McIntosh, A. R. (2000). Towards a network theory of cognition. *Neural Networks, 13,* 861-870.
- McIntosh, A. R., Bookstein, F. L., Haxby, J. V., & Grady, C. L. (1996). Spatial pattern analysis of functional brain images using partial least squares. *Neuroimage.*, 3, 143-157.
- McIntosh, A. R. & Gonzalez-Lima, F. (1994a). Network interactions among limbic cortices, basal forebrain, and cerebellum differentiate a tone conditioned as a Pavlovian excitor or inhibitor: fluorodeoxyglucose mapping and covariance structural modeling. *Journal of Neurophysiology*, 72, 1717-1733.
- McIntosh, A. R. & Gonzalez-Lima, F. (1994b). Structural equation modeling and its application to network analysis in functional brain imaging. *Human Brain Mapping*, *2*, 2-22.

- Meaney, M. J. (2001). Maternal care, gene expression, and the transmission of individual differences in stress reactivity across generations. *Annual Review of Neuroscience, 24,* 1161-1192.
- Minor, T. R., Jackson, R. L., & Maier, S. F. (1984). Effects of task-irrelevant cues and reinforcement delay on choice-escape learning following inescapable shock: evidence for a deficit in selective attention. *Journal of Experimental Psychology.Animal Behavior Processes*, 10, 543-556.
- Modell, S., Yassouridis, A., Huber, J., & Holsboer, F. (1997). Corticosteroid receptor function is decreased in depressed patients. *Neuroendocrinology*, 65, 216-222.
- Moga, M. M., Weis, R. P., & Moore, R. Y. (1995). Efferent projections of the paraventricular thalamic nucleus in the rat. *The Journal of Comparative Neurology*, 359, 221-238.
- Monmaur, P., Sharif, A., & M'Harzi, M. (1997a). Involvement of septal muscarinic receptors in cholinergically mediated changes in rat rearing activity. *Pharmacology, biochemistry, and Behavior, 58,* 577-582.
- Monmaur, P., Sharif, A., & M'Harzi, M. (1997b). Involvement of septal muscarinic receptors in cholinergically mediated changes in rat rearing activity. *Pharmacology, biochemistry, and Behavior, 58,* 577-582.
- Montone, K. T., Fass, B., & Hamill, G. S. (1988). Serotonergic and nonserotonergic projections from the rat interpeduncular nucleus to the septum, hippocampal formation and raphe: a combined immunocytochemical and fluorescent retrograde labelling study of neurons in the apical subnucleus. *Brain Research Bulletin, 20,* 233-240.
- Morris, J. S., Smith, K. A., Cowen, P. J., Friston, K. J., & Dolan, R. J. (1999). Covariation of activity in habenula and dorsal raphe nuclei following tryptophan depletion. *Neuroimage*, 10, 163-172.
- Morris, P. L., Robinson, R. G., Raphael, B., & Hopwood, M. J. (1996). Lesion location and poststroke depression. *The Journal of Neuropsychiatry and Clinical Neurosciences*, 8, 399-403.
- Mulders, W. H., Meek, J., Hafmans, T. G., & Cools, A. R. (1997). Plasticity in the stressregulating circuit: decreased input from the bed nucleus of the stria terminalis to the hypothalamic paraventricular nucleus in Wistar rats following adrenalectomy. *The European Journal of Neuroscience*, 9, 2462-2471.
- Myhrer, T. (1989a). Exploratory behavior and reaction to novelty in rats: effects of medial and lateral septal lesions. *Behavioral Neuroscience*, 103, 1226-1233.

- Myhrer, T. (1989b). Exploratory behavior and reaction to novelty in rats: effects of medial and lateral septal lesions. *Behavioral Neuroscience*, 103, 1226-1233.
- Nair, H. P. & Gonzalez-Lima, F. (1999). Extinction of behavior in infant rats: development of functional coupling between septal, hippocampal, and ventral tegmental regions. *The Journal of Neuroscience*, 19, 8646-8655.
- Nestler, E. J., Barrot, M., DiLeone, R. J., Eisch, A. J., Gold, S. J., & Monteggia, L. M. (2002). Neurobiology of depression. *Neuron*, 34, 13-25.
- Newport, D. J. & Nemeroff, C. B. (2000). Neurobiology of posttraumatic stress disorder. *Current Opinion in Neurobiology*, 10, 211-218.
- Nibuya, M., Nestler, E. J., & Duman, R. S. (1996). Chronic antidepressant administration increases the expression of cAMP response element binding protein (CREB) in rat hippocampus. *The Journal of Neuroscience, 16*, 2365-2372.
- Nishikawa, T., Fage, D., & Scatton, B. (1986). Evidence for, and nature of, the tonic inhibitory influence of habenulointerpeduncular pathways upon cerebral dopaminergic transmission in the rat. *Brain Research*, *373*, 324-336.
- Nofzinger, E. A., Nichols, T. E., Meltzer, C. C., Price, J., Steppe, D. A., Miewald, J. M. et al. (1999). Changes in forebrain function from waking to REM sleep in depression: preliminary analyses of [18F]FDG PET studies. *Psychiatry Research*, *91*, 59-78.
- Nurnberg, H. G., Thompson, P. M., & Hensley, P. L. (1999). Antidepressant medication change in a clinical treatment setting: a comparison of the effectiveness of selective serotonin reuptake inhibitors. J Clin. Psychiatry, 60, 574-579.
- O'Dell, L. E., Sussman, A. N., Meyer, K. L., & Neisewander, J. L. (1999). Behavioral effects of psychomotor stimulant infusions into amygdaloid nuclei. *Neuropsychopharmacology*, 20, 591-602.
- Oddie, S. D., Stefanek, W., Kirk, I. J., & Bland, B. H. (1996). Intraseptal procaine abolishes hypothalamic stimulation-induced wheel-running and hippocampal theta field activity in rats. *The Journal of Neuroscience*, *16*, 1948-1956.
- Ongur, D., Drevets, W. C., & Price, J. L. (1998). Glial reduction in the subgenual prefrontal cortex in mood disorders. *Proceedings of the National Academy of Sciences of the United States of America*, 95, 13290-13295.
- Otake, K., Ruggiero, D. A., Regunathan, S., Wang, H., Milner, T. A., & Reis, D. J. (1998). Regional localization of agmatine in the rat brain: an immunocytochemical study. *Brain Research*, 787, 1-14.

- Overmier, J. B. & Seligman, M. E. P. (1967). Effects of inescapable shock on subsequent escape and avoidance behavior. *Journal of Comparative and Physiological Psychology*, 63, 28-33.
- Palkovits, M. (2000). Stress-induced expression of co-localized neuropeptides in hypothalamic and amygdaloid neurons. *European Journal of Pharmacology, 405,* 161-166.
- Papa, M., Sadile, A. G., Sergeant, J. A., Shumake, J., & Gonzalez-Lima, F. (1998). Functional imaging probes to study the neural bases of behavior in genetic animal models of ADHD. In F.Gonzalez-Lima (Ed.), *Cytochrome oxidase in neuronal metabolism and Alzheimer's disease* (pp. 145-170). New York: Plenum.
- Papolos, D. F., Edwards, E., Marmur, R., Lachman, H. M., & Henn, F. A. (1993). Effects of the antiglucocorticoid RU 38486 on the induction of learned helpless behavior in Sprague-Dawley rats. *Brain Research*, 615, 304-309.
- Parashos, I. A., Tupler, L. A., Blitchington, T., & Krishnan, K. R. (1998). Magneticresonance morphometry in patients with major depression. *Psychiatry Research*, 84, 7-15.
- Pariante, C. M. & Miller, A. H. (2001). Glucocorticoid receptors in major depression: relevance to pathophysiology and treatment. *Biological Psychiatry*, *49*, 391-404.
- Patel, J. G., Bartoszyk, G. D., Edwards, E., & Ashby, C. R., Jr. (2004). The highly selective 5-hydroxytryptamine (5-HT)2A receptor antagonist, EMD 281014, significantly increases swimming and decreases immobility in male congenital learned helpless rats in the forced swim test. *Synapse*, 52, 73-75.
- Paxinos, G., Törk, I., Tecott, L. H., & Valentino, K. L. (1991). Atlas of the Developing Rat Brain. San Diego: Academic Press.
- Paxinos, G. & Watson, C. (1997). *The Rat Brain in Stereotaxic Coordinates*. (Compact Third Edition ed.) San Diego: Academic Press.
- Paxinos, G. & Watson, C. (1998). *The Rat Brain in Stereotaxic Coordinates*. San Diego: Academic Press.
- Petty, F., Davis, L. L., Kabel, D., & Kramer, G. L. (1996). Serotonin dysfunction disorders: a behavioral neurochemistry perspective. *The Journal of Clinical Psychiatry*, 57 Suppl 8, 11-16.
- Petty, F., Kramer, G. L., Wu, J., & Davis, L. L. (1997). Posttraumatic stress and depression. A neurochemical anatomy of the learned helplessness animal model. *Annals of the New York Academy of Sciences*, 821, 529-532.

- Petty, F. & Sherman, A. D. (1981). GABAergic modulation of learned helplessness. *Pharmacology, biochemistry, and Behavior, 15,* 567-570.
- Piazza, P. V., Deminiere, J. M., Le Moal, M., & Simon, H. (1989). Factors that predict individual vulnerability to amphetamine self-administration. *Science*, 245, 1511-1513.
- Piazza, P. V. & Le Moal, M. (1997). Glucocorticoids as a biological substrate of reward: physiological and pathophysiological implications. *Brain Research.Brain Research Reviews*, 25, 359-372.
- Pitman, R. K., van der Kolk, B. A., Orr, S. P., & Greenberg, M. S. (1990). Naloxonereversible analgesic response to combat-related stimuli in posttraumatic stress disorder. A pilot study. *Archives of General Psychiatry*, 47, 541-544.
- Pizzolato, G., Soncrant, T. T., & Rapoport, S. I. (1984). Haloperidol and cerebral metabolism in the conscious rat: relation to pharmacokinetics. *Journal of Neurochemistry*, 43, 724-732.
- Plenge, P., Mellerup, E. T., & Wortwein, G. (2002). Characterization of epibatidine binding to medial habenula: potential role in analgesia. J Pharmacol.Exp.Ther., 302, 759-765.
- Poplawsky, A. & Isaacson, R. L. (1983b). Changes in open-field behaviors following septal lesions in rats. *Behavioral and Neural Biology*, 38, 61-69.
- Poplawsky, A. & Isaacson, R. L. (1983a). Changes in open-field behaviors following septal lesions in rats. *Behavioral and Neural Biology*, 38, 61-69.
- Porrino, L. J., Esposito, R. U., Seeger, T. F., Crane, A. M., Pert, A., & Sokoloff, L. (1984). Metabolic mapping of the brain during rewarding self-stimulation. *Science*, 224, 306-309.
- Porsolt, R. D. (2000). Animal models of depression: utility for transgenic research. *Reviews in the Neurosciences, 11,* 53-58.
- Porsolt, R. D., Le Pichon, M., & Jalfre, M. (1977). Depression: a new animal model sensitive to antidepressant treatments. *Nature*, *266*, 730-732.
- Prisco, S. & Esposito, E. (1995). Differential effects of acute and chronic fluoxetine administration on the spontaneous activity of dopaminergic neurones in the ventral tegmental area. *British Journal of Pharmacology*, *116*, 1923-1931.
- Pruessner, J. C., Hellhammer, D. H., & Kirschbaum, C. (1999). Burnout, perceived stress, and cortisol responses to awakening. *Psychosomatic Medicine*, 61, 197-204.

- Prut, L. & Belzung, C. (2003). The open field as a paradigm to measure the effects of drugs on anxiety-like behaviors: a review. *Eur.J Pharmacol.*, 463, 3-33.
- Purba, J. S., Hoogendijk, W. J., Hofman, M. A., & Swaab, D. F. (1996). Increased number of vasopressin- and oxytocin-expressing neurons in the paraventricular nucleus of the hypothalamus in depression. *Archives of General Psychiatry*, 53, 137-143.
- Raadsheer, F. C., Hoogendijk, W. J., Stam, F. C., Tilders, F. J., & Swaab, D. F. (1994). Increased numbers of corticotropin-releasing hormone expressing neurons in the hypothalamic paraventricular nucleus of depressed patients. *Neuroendocrinology*, 60, 436-444.
- Rauch, S. L., van der Kolk, B. A., Fisler, R. E., Alpert, N. M., Orr, S. P., Savage, C. R. et al. (1996). A symptom provocation study of posttraumatic stress disorder using positron emission tomography and script-driven imagery. *Archives of General Psychiatry*, 53, 380-387.
- Rauch, S. L., Whalen, P. J., Shin, L. M., McInerney, S. C., Macklin, M. L., Lasko, N. B. et al. (2000). Exaggerated amygdala response to masked facial stimuli in posttraumatic stress disorder: a functional MRI study. *Biological Psychiatry*, 47, 769-776.
- Rawlins, J. N., Feldon, J., & Gray, J. A. (1979). Septo-hippocampal connections and the hippocampal theta rhythm. *Experimental Brain Research*, *37*, 49-63.
- Reiser, L. W. & Reiser, M. F. (1995). Endocrine disorders. In H.I.Kaplan & B. J. Sadock (Eds.), *Comprehensive Textbook of Psychiatry* (IV ed., pp. 1170-1171). Baltimore: William and Wilkins.
- Rhodes, J. S., Garland, T., Jr., & Gammie, S. C. (2003). Patterns of brain activity associated with variation in voluntary wheel-running behavior. *Behavioral Neuroscience*, *117*, 1243-1256.
- Richman, H. & Frueh, B. C. (1996). Personality disorder symptomatology among Vietnam veterans with combat-related PTSD. *Anxiety.*, *2*, 286-295.
- Riemann, D., Berger, M., & Voderholzer, U. (2001). Sleep and depression--results from psychobiological studies: an overview. *Biological Psychology*, *57*, 67-103.
- Risold, P. Y. & Swanson, L. W. (1997). Connections of the rat lateral septal complex. *Brain Research.Brain Research Reviews, 24,* 115-195.
- Rogers, M. A., Bradshaw, J. L., Pantelis, C., & Phillips, J. G. (1998). Frontostriatal deficits in unipolar major depression. *Brain Research Bulletin, 47, 297-310.*

- Ronan, P. J., Steciuk, M., Kramer, G. L., Kram, M., & Petty, F. (2000). Increased septal 5-HIAA efflux in rats that do not develop learned helplessness after inescapable stress. *Journal of Neuroscience Research*, 61, 101-106.
- Rubin, R. T., Phillips, J. J., McCracken, J. T., & Sadow, T. F. (1996). Adrenal gland volume in major depression: relationship to basal and stimulated pituitary-adrenal cortical axis function. *Biological Psychiatry*, 40, 89-97.
- Sachinvala, N., Kling, A., Suffin, S., Lake, R., & Cohen, M. (2000). Increased regional cerebral perfusion by 99mTc hexamethyl propylene amine oxime single photon emission computed tomography in post-traumatic stress disorder. *Mil.Med.*, 165, 473-479.
- Sakata, J. T., Coomber, P., Gonzalez-Lima, F., & Crews, D. (2000). Functional connectivity among limbic brain areas: differential effects of incubation temperature and gonadal sex in the leopard gecko, Eublepharis macularius. *Brain, Behavior and Evolution, 55,* 139-151.
- Sanders, A. R., Detera-Wadleigh, S. D., & Gershon, E. S. (1999). Molecular genetics of mood disorders. In D.S.Charney, E. J. Nestler, & B. S. Bunney (Eds.), *Neurobiology of Mental Illness* (pp. 299-316). New York: Oxford.
- Sandyk, R. (1991). Relevance of the habenular complex to neuropsychiatry: a review and hypothesis. *The International Journal of Neuroscience*, *61*, 189-219.
- Sapolsky, R. M. (2000). Glucocorticoids and hippocampal atrophy in neuropsychiatric disorders. *Archives of General Psychiatry*, *57*, 925-935.
- Sapolsky, R. M., Krey, L. C., & McEwen, B. S. (1986). The neuroendocrinology of stress and aging: the glucocorticoid cascade hypothesis. *Endocrine Reviews*, 7, 284-301.
- Saxena, S., Brody, A. L., Ho, M. L., Alborzian, S., Ho, M. K., Maidment, K. M. et al. (2001). Cerebral metabolism in major depression and obsessive-compulsive disorder occurring separately and concurrently. *Biological Psychiatry*, 50, 159-170.
- Scarpa, A. & Luscher, K. A. (2002). Self-esteem, cortisol reactivity, and depressed mood mediated by perceptions of control. *Biological Psychology*, 59, 93-103.
- Schafer, M. K., Eiden, L. E., & Weihe, E. (1998). Cholinergic neurons and terminal fields revealed by immunohistochemistry for the vesicular acetylcholine transporter. I. Central nervous system. *Neuroscience*, 84, 331-359.
- Schultz, W. (2000). Multiple reward signals in the brain. *Nature Reviews.Neuroscience*, *1*, 199-207.

Schultz, W. (2002). Getting formal with dopamine and reward. Neuron, 36, 241-263.

- Schultz, W. & Dickinson, A. (2000). Neuronal coding of prediction errors. *Annual Review of Neuroscience, 23,* 473-500.
- Scott, L. V. & Dinan, T. G. (1998). Vasopressin and the regulation of hypothalamicpituitary-adrenal axis function: implications for the pathophysiology of depression. *Life Sciences*, 62, 1985-1998.
- Seggie, J., Uhlir, I., & Brown, G. M. (1974). Adrenal stress responses following septal lesions in the rat. *Neuroendocrinology*, 16, 225-236.
- Seligman, M. E. P. (1975). *Helplessness: On Depression, Development, and Death.* San Francisco: W.H. Freeman and Company.
- Seligman, M. E. P. & Maier, S. F. (1967). Failure to escape traumatic shock. *Journal of Experimental Psychology*, 74, 1-9.
- Semple, W. E., Goyer, P., McCormick, R., Morris, E., Compton, B., Muswick, G. et al. (1993). Preliminary report: brain blood flow using PET in patients with posttraumatic stress disorder and substance-abuse histories. *Biological Psychiatry*, 34, 115-118.
- Semple, W. E., Goyer, P. F., McCormick, R., Compton-Toth, B., Morris, E., Donovan, B. et al. (1996). Attention and regional cerebral blood flow in posttraumatic stress disorder patients with substance abuse histories. *Psychiatry Research*, 67, 17-28.
- Shah, P. J., Ebmeier, K. P., Glabus, M. F., & Goodwin, G. M. (1998). Cortical grey matter reductions associated with treatment-resistant chronic unipolar depression. Controlled magnetic resonance imaging study. *Br.J Psychiatry*, 172, 527-532.
- Sheline, Y. I., Wang, P. W., Gado, M. H., Csernansky, J. G., & Vannier, M. W. (1996). Hippocampal atrophy in recurrent major depression. *Proceedings of the National Academy of Sciences of the United States of America*, 93, 3908-3913.
- Sherman, A. D. & Petty, F. (1980). Neurochemical basis of the action of antidepressants on learned helplessness. *Behavioral and Neural Biology, 30,* 119-134.
- Sherman, A. D. & Petty, F. (1984). Learned helplessness decreases [3H]imipramine binding in rat cortex. *Journal of Affective Disorders, 6,* 25-32.
- Sherman, A. D., Sacquitne, J. L., & Petty, F. (1982). Specificity of the learned helplessness model of depression. *Pharmacology, biochemistry, and Behavior, 16,* 449-454.

- Shin, L. M., Kosslyn, S. M., McNally, R. J., Alpert, N. M., Thompson, W. L., Rauch, S. L. et al. (1997a). Visual imagery and perception in posttraumatic stress disorder. A positron emission tomographic investigation. *Archives of General Psychiatry*, 54, 233-241.
- Shin, L. M., McNally, R. J., Kosslyn, S. M., Thompson, W. L., Rauch, S. L., Alpert, N. M. et al. (1997b). A positron emission tomographic study of symptom provocation in PTSD. Annals of the New York Academy of Sciences, 821, 521-523.
- Shin, L. M., McNally, R. J., Kosslyn, S. M., Thompson, W. L., Rauch, S. L., Alpert, N. M. et al. (1999). Regional cerebral blood flow during script-driven imagery in childhood sexual abuse-related PTSD: A PET investigation. *American Journal of Psychiatry*, 156, 575-584.
- Shin, L. M., Whalen, P. J., Pitman, R. K., Bush, G., Macklin, M. L., Lasko, N. B. et al. (2001). An fMRI study of anterior cingulate function in posttraumatic stress disorder. *Biological Psychiatry*, 50, 932-942.
- Shinoda, K., Inagaki, S., Shiosaka, S., Kohno, J., & Tohyama, M. (1984). Experimental immunohistochemical studies on the substance P neuron system in the lateral habenular nucleus of the rat: distribution and origins. *J Comp Neurol.*, 222, 578-588.
- Shinoda, K. & Tohyama, M. (1987). Analysis of the habenulopetal enkephalinergic system in the rat brain: an immunohistochemical study. J Comp Neurol., 255, 483-496.
- Shumake, J., Edwards, E., & Gonzalez-Lima, F. (2003). Opposite metabolic changes in the habenula and ventral tegmental area of a genetic model of helpless behavior. *Brain Research*, 963, 274-281.
- Shumake, J. & Gonzalez-Lima, F. (2003). Brain systems underlying susceptibility to helplessness and depression. *Behavioral and Cognitive Neuroscience Reviews*, 2, 198-221.
- Silverman, A. J., Hoffman, D. L., & Zimmerman, E. A. (1981). The descending afferent connections of the paraventricular nucleus of the hypothalamus (PVN). *Brain Research Bulletin, 6,* 47-61.
- Slawinska, U. & Kasicki, S. (1998b). The frequency of rat's hippocampal theta rhythm is related to the speed of locomotion. *Brain Research*, *796*, 327-331.
- Slawinska, U. & Kasicki, S. (1998a). The frequency of rat's hippocampal theta rhythm is related to the speed of locomotion. *Brain Research*, *796*, 327-331.

- Speciale, S. G., Neckers, L. M., & Wyatt, R. J. (1980). Habenular modulation of raphe indoleamine metabolism. *Life Sciences*, *27*, 2367-2372.
- Steciuk, M., Kram, M., Kramer, G. L., & Petty, F. (1999). Decrease in stress-induced c-Fos-like immunoreactivity in the lateral septal nucleus of learned helpless rats. *Brain Research*, 822, 256-259.
- Stein, M. B., Koverola, C., Hanna, C., Torchia, M. G., & McClarty, B. (1997). Hippocampal volume in women victimized by childhood sexual abuse. *Psychological Medicine*, 27, 951-959.
- Steingard, R. J., Renshaw, P. F., Yurgelun-Todd, D., Appelmans, K. E., Lyoo, I. K., Shorrock, K. L. et al. (1996). Structural abnormalities in brain magnetic resonance images of depressed children. *Journal of the American Academy of Child and Adolescent Psychiatry*, 35, 307-311.
- Steingard, R. J., Yurgelun-Todd, D. A., Hennen, J., Moore, J. C., Moore, C. M., Vakili, K. et al. (2000). Increased orbitofrontal cortex levels of choline in depressed adolescents as detected by in vivo proton magnetic resonance spectroscopy. *Biological Psychiatry*, 48, 1053-1061.
- Stern, Y. & Langston, J. W. (1985). Intellectual changes in patients with MPTP-induced parkinsonism. *Neurology*, 35, 1506-1509.
- Sutherland, R. J. (1982). The dorsal diencephalic conduction system: a review of the anatomy and functions of the habenular complex. *Neuroscience and Biobehavioral Reviews*, *6*, 1-13.
- Sutherland, R. J. & Nakajima, S. (1981). Self-stimulation of the habenular complex in the rat. *Journal of Comparative and Physiological Psychology*, 95, 781-791.
- Sutton, L. C., Grahn, R. E., Wiertelak, E. P., Watkins, L. R., & Maier, S. F. (1997). Inescapable shock-induced potentiation of morphine analgesia in rats: involvement of opioid, GABAergic, and serotonergic mechanisms in the dorsal raphe nucleus. *Behavioral Neuroscience*, 111, 816-824.
- Swanson, L. W. & Risold, P. (2000). On the basic architecture of the septal region. In R.Numan (Ed.), *The behavioral neuroscience of the septal region* (pp. 1-14). New York: Springer-Verlag.
- Tononi, G., Sporns, O., & Edelman, G. M. (1994). A measure for brain complexity: relating functional segregation and integration in the nervous system. *Proceedings* of the National Academy of Sciences of the United States of America, 91, 5033-5037.

- Treit, D. & Menard, J. (2000). The septum and anxiety. In R.Numan (Ed.), *The behavioral neuroscience of the septal region* (pp. 210-233). New York: Springer-Verlag.
- True, W. R., Rice, J., Eisen, S. A., Heath, A. C., Goldberg, J., Lyons, M. J. et al. (1993). A twin study of genetic and environmental contributions to liability for posttraumatic stress symptoms. *Archives of General Psychiatry*, 50, 257-264.
- Trugman, J. M., James, C. L., & Wooten, G. F. (1991). D1/D2 dopamine receptor stimulation by L-dopa. A [14C]-2-deoxyglucose autoradiographic study. *Brain: A Journal of Neurology, 114 (Pt 3)*, 1429-1440.
- Uhlir, I., Seggie, J., & Brown, G. M. (1974). The effect of septal lesions on the threshold of adrenal stress response. *Neuroendocrinology*, *14*, 351-355.
- Ullsperger, M. & Von Cramon, D. Y. (2003). Error monitoring using external feedback: specific roles of the habenular complex, the reward system, and the cingulate motor area revealed by functional magnetic resonance imaging. *The Journal of Neuroscience*, 23, 4308-4314.
- Uylings, H. B., Groenewegen, H. J., & Kolb, B. (2003). Do rats have a prefrontal cortex? *Behavioural Brain Research*, *146*, 3-17.
- Uylings, H. B. & van Eden, C. G. (1990). Qualitative and quantitative comparison of the prefrontal cortex in rat and in primates, including humans. *Progress in Brain Research*, 85, 31-62.
- Valjakka, A., Vartiainen, J., Tuomisto, L., Tuomisto, J. T., Olkkonen, H., & Airaksinen, M. M. (1998). The fasciculus retroflexus controls the integrity of REM sleep by supporting the generation of hippocampal theta rhythm and rapid eye movements in rats. *Brain Research Bulletin, 47*, 171-184.
- van der Kolk, B. A., Greenberg, M. S., Orr, S. P., & Pitman, R. K. (1989). Endogenous opioids, stress induced analgesia, and posttraumatic stress disorder. *Psychopharmacology Bulletin, 25,* 417-421.
- Van Pett, K., Viau, V., Bittencourt, J. C., Chan, R. K., Li, H. Y., Arias, C. et al. (2000). Distribution of mRNAs encoding CRF receptors in brain and pituitary of rat and mouse. *The Journal of Comparative Neurology*, 428, 191-212.
- Vazquez-Palacios, G., Bonilla-Jaime, H., & Velazquez-Moctezuma, J. (2004a). Antidepressant-like effects of the acute and chronic administration of nicotine in the rat forced swimming test and its interaction with flouxetine. *Pharmacology, biochemistry, and Behavior, 78,* 165-169.

- Vazquez-Palacios, G., Bonilla-Jaime, H., & Velazquez-Moctezuma, J. (2004b). Antidepressant-like effects of the acute and chronic administration of nicotine in the rat forced swimming test and its interaction with flouxetine. *Pharmacology*, *biochemistry*, and Behavior, 78, 165-169.
- Vazquez-Palacios, G., Bonilla-Jaime, H., & Velazquez-Moctezuma, J. (2004c). Antidepressant-like effects of the acute and chronic administration of nicotine in the rat forced swimming test and its interaction with flouxetine. *Pharmacology*, *biochemistry*, and Behavior, 78, 165-169.
- Vermetten, E., Charney, D. S., & Bremner, J. D. (2001). Post-traumatic stress disorder. In F.Henn, N. Sartorius, H. Helmchen, & H. Lauter (Eds.), Part 2: Personality Disorders, Anxiety and Related Disorders, Behavioural and Addictive Disorders (4th ed., pp. 37-76). Heidelberg: Springer Verlag.
- Videbech, P. (2000). PET measurements of brain glucose metabolism and blood flow in major depressive disorder: a critical review. *Acta Psychiatrica Scandinavica*, 101, 11-20.
- Villarreal, G., Hamilton, D. A., Petropoulos, H., Driscoll, I., Rowland, L. M., Griego, J. A. et al. (2002). Reduced hippocampal volume and total white matter volume in posttraumatic stress disorder. *Biological Psychiatry*, 52, 119-125.
- Vogt, B. A. & Peters, A. (1981). Form and distribution of neurons in rat cingulate cortex: areas 32, 24, and 29. *The Journal of Comparative Neurology*, 195, 603-625.
- Vollmayr, B., Bachteler, D., Vengeliene, V., Gass, P., Spanagel, R., & Henn, F. (2004). Rats with congenital learned helplessness respond less to sucrose but show no deficits in activity or learning. *Behavioural Brain Research*, 150, 217-221.
- Wang, R. Y. & Aghajanian, G. K. (1977). Physiological evidence for habenula as major link between forebrain and midbrain raphe. *Science*, 197, 89-91.
- Wang, S., Mason, J., Charney, D., Yehuda, R., Riney, S., & Southwick, S. (1997). Relationships between hormonal profile and novelty seeking in combat-related posttraumatic stress disorder. *Biological Psychiatry*, 41, 145-151.
- Wang, T., Palkovits, M., Rusnak, M., Mezey, E., & Usdin, T. B. (2000). Distribution of parathyroid hormone-2 receptor-like immunoreactivity and messenger RNA in the rat nervous system. *Neuroscience*, 100, 629-649.
- Watts, A. G., Swanson, L. W., & Sanchez-Watts, G. (1987). Efferent projections of the suprachiasmatic nucleus: I. Studies using anterograde transport of Phaseolus vulgaris leucoagglutinin in the rat. *The Journal of Comparative Neurology*, 258, 204-229.

- Weissman, A. D., Marquis, K. L., Moreton, J. E., & London, E. D. (1989). Effects of self-administered phencyclidine on regional uptake of 2- deoxy-D-[1-14C]glucose in brain. *Neuropharmacology*, 28, 575-583.
- Wilkerson, G. & London, E. D. (1989). Effects of methylenedioxymethamphetamine on local cerebral glucose utilization in the rat. *Neuropharmacology*, *28*, 1129-1138.
- Willner, P. (1990). Animal models of depression: an overview. *Pharmacology & Therapeutics*, 45, 425-455.
- Willner, P., Towell, A., Sampson, D., Sophokleous, S., & Muscat, R. (1987). Reduction of sucrose preference by chronic unpredictable mild stress, and its restoration by a tricyclic antidepressant. *Psychopharmacology*, *93*, 358-364.
- Wong-Riley, M. T. (1989). Cytochrome oxidase: an endogenous metabolic marker for neuronal activity. *Trends Neurosci.*, 12, 94-101.
- Wong-Riley, M. T., Nie, F., Hevner, R. F., & Liu, S. (1998). Brain cytochrome oxidase: Functional significance and bigenomic regulation in the CNS. In F.Gonzalez-Lima (Ed.), Cytochrome oxidase in neuronal metabolism and Alzheimer's disease (pp. 1-54). New York: Plenum Press.
- Woolf, N. J. & Butcher, L. L. (1985). Cholinergic systems in the rat brain: II. Projections to the interpeduncular nucleus. *Brain Research Bulletin*, 14, 63-83.
- Yadid, G., Nakash, R., Deri, I., Tamar, G., Kinor, N., Gispan, I. et al. (2000). Elucidation of the neurobiology of depression: insights from a novel genetic animal model. *Progress in Neurobiology*, 62, 353-378.
- Yehuda, R. (1999). Linking the neuroendocrinology of post-traumatic stress disorder with recent neuroanatomic findings. *Seminars in Clinical Neuropsychiatry*, 4, 256-265.
- Yehuda, R. & Antelman, S. M. (1993). Criteria for rationally evaluating animal models of posttraumatic stress disorder. *Biological Psychiatry*, 33, 479-486.
- Yehuda, R., Levengood, R. A., Schmeidler, J., Wilson, S., Guo, L. S., & Gerber, D. (1996). Increased pituitary activation following metyrapone administration in post-traumatic stress disorder. *Psychoneuroendocrinology*, 21, 1-16.
- Yehuda, R., McFarlane, A. C., & Shalev, A. Y. (1998). Predicting the development of posttraumatic stress disorder from the acute response to a traumatic event. *Biological Psychiatry*, 44, 1305-1313.
- Yehuda, R., Southwick, S. M., Krystal, J. H., Bremner, D., Charney, D. S., & Mason, J. W. (1993). Enhanced suppression of cortisol following dexamethasone

administration in posttraumatic stress disorder. The American Journal of Psychiatry, 150, 83-86.

- Zhang, K. Z., Westberg, J. A., Holtta, E., & Andersson, L. C. (1996). BCL2 regulates neural differentiation. *Proceedings of the National Academy of Sciences of the United States of America*, 93, 4504-4508.
- Zilles, K. & Wree, A. (1985). Cortex: Areal and laminar structure. In G.Paxinos (Ed.), *The rat nervous system* (pp. 375-415). Sydney: Academic Press.
- Zorrilla, E. P., DeRubeis, R. J., & Redei, E. (1995). High self-esteem, hardiness and affective stability are associated with higher basal pituitary-adrenal hormone levels. *Psychoneuroendocrinology*, 20, 591-601.

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