

## REPTILIA: SQUAMATA: SAURIA: TEIIDAE

## CNEMIDOPHORUS SEXLINEATUS

## Catalogue of American Amphibians and Reptiles.

Trauth, S.E. and C.T. McAllister. 1996. *Cnemidophorus sexlineatus*.

***Cnemidophorus sexlineatus* (Linnaeus)**  
Six-lined Racerunner

*Lacerta sexlineata* Linnaeus, 1766:364. Type-locality, "Carolina", restricted to Charleston, Charleston County, South Carolina, by Smith and Taylor (1950b). Holotype lost (see Maslin and Secoy, 1986), collected by A. Garden, date of collection unknown.

*Lacerta 6-lineata*: Müller, 1774:94. Emendation.

*Lacerta sex-lineata*: Bonnaterre, 1789:48. Emendation.

*Seps sexlineata*: Cuvier, 1817 (1816):29.

*Teius lemniscatus*: Merrem, 1820:60 (part).

*Lacerta fallax* Merrem, 1820:63. Type-locality, "Carolina." No type-specimens are known to exist. Collector and date of collection unknown.

*Am.[eiva] sex-lineata*: M'Murtrie, in Cuvier, 1831:22.

*Ameiva sexlineata*: M'Murtrie, in Cuvier, 1831:23. Emendation.

*Lacertam sexlineatam*: Wiegmann, 1834:27.

*Cnemidophorus sexlineatus*: Duméril and Bibron, 1839:131. First use of combination.

*Aspidoscelis sexlineatus*: Fitzinger, 1843:20.

*Cnemidophorus sex-lineatus*: Baird, 1859:38. Emendation.

*Cnemidophorus sexlineatus* var. *gularis*: Cope, 1866 (1867):303 (part).

*Cnemidophorus sexlineilus*: Cope, 1893 (1894):387. *Lapsus*.

*Cnemidophorus sexlineatus sexlineatus*: Cope, 1898 (1900):598.

*Cnemidophorus sexlinsatus*: McLain, 1899:2. *Lapsus*.

*Cnemydophorus sexlineatus*: Tofohr, 1902:201. Emendation.

*Cnemidophorus sexlineatus sexlineatus*: Brown, 1903:546. *Lapsus*.

• **Content.** Three subspecies are recognized: *sexlineatus*, *stephensae*, and *viridis*.

• **Definition.** *Cnemidophorus sexlineatus* is a medium-sized lizard of the *sexlineatus* species group, with adult SVL 66-90

mm (Trauth, 1980). Adult tail length ranges 146-190 mm. Maximum head length of 18.6 mm occurs in males. Scalation is as follows: granules around midbody (GAB) number 62-115, femoral pores 20-42, enlarged subdigital lamellae of the fourth pes 20-39, granules between the paravertebral stripes at mid-body (PVG) 3-24, lateral supraocular granules (LSG) 5-62, circumorbital granules (COG) 1-29, granules from occiput to rump (GOR) 155-267, granules encompassing the vertebral stripe 0-15, and the PVG/GAB ( $\times 100$ ) 9.9-26.7. The postantebrachial scales usually are small but can be enlarged (but not greatly enlarged as in *C. gularis*). Supraocular scales number 4, parietals 3, and frontoparietals 2; the number of scales in the latter two groups may be increased by subdivision, especially in Atlantic Coast populations from Maryland to Florida.

Dorsal adult coloration shows considerable geographic variation. The dorsolateral fields can be dark brown or black with an anterior greenish suffusion on the lateral body surfaces up to and including the dorsolateral stripe (Fig. 1), or the entire trunk may be green. The face and neck may be blue, bluish-green, or yellow. Dorsal longitudinal stripes usually are white or yellow, and may number 6-8 (Fig. 2). Adult males are light blue to bluish green ventrally, whereas adult females vary from a salmon pink to pearly white (or hues of blue). Tail coloration is blue in hatchlings, fading to brown or slate in adults. Juveniles of *sexlineatus* and *viridis* exhibit white stripes upon dark fields, and the venter is white in all subspecies. Juveniles of *stephensae* are yellow on the head and neck.

• **Descriptions.** Trauth (1980) provided the most recent detailed description of this species and included information on geographic variation in scutellation, color, and striping pattern throughout its range (see Comment). Additional descriptions can be found in Burt (1931a), Cope (1898 [1900]), Duellman and Zweifel (1962), Smith (1946), Smith and List (1955), Walker et al. (1990a, b), and Wright (1967). Bickham et al. (1976) and Lowe et al. (1970) described the karyotype ( $2N = 46$ , with 2 large metacentric macrochromosomes, 24 medium-sized telocentric and subtelocentric macrochromosomes, and 20 microchromosomes).

• **Illustrations.** Black and white photographs of the various subspecies are numerous and include the following: *sexlineatus*

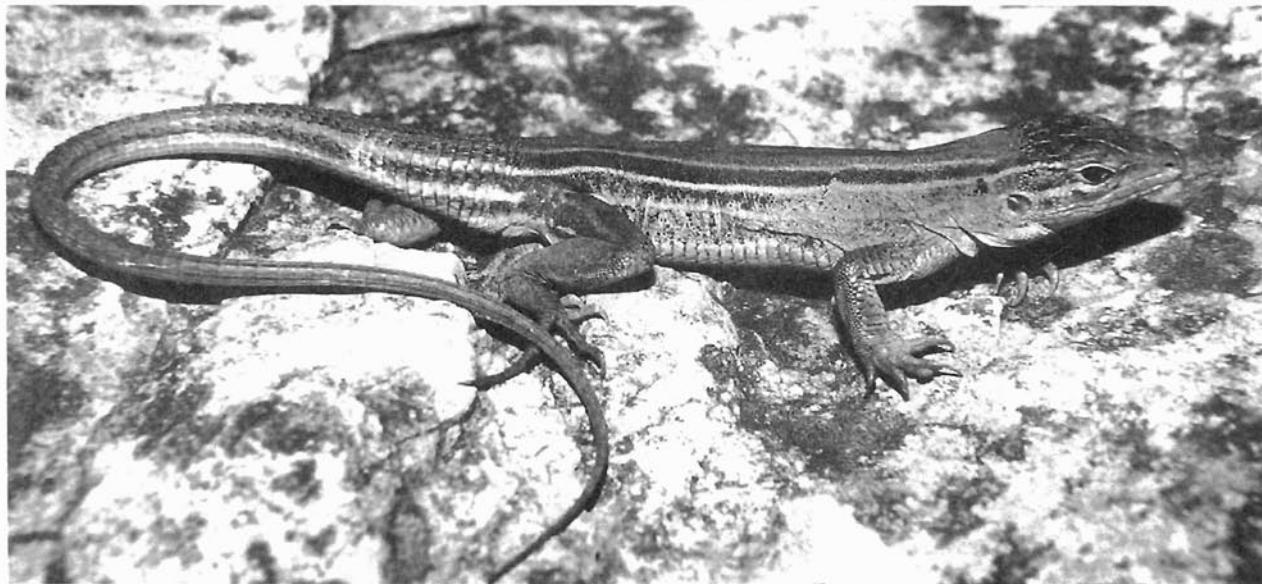
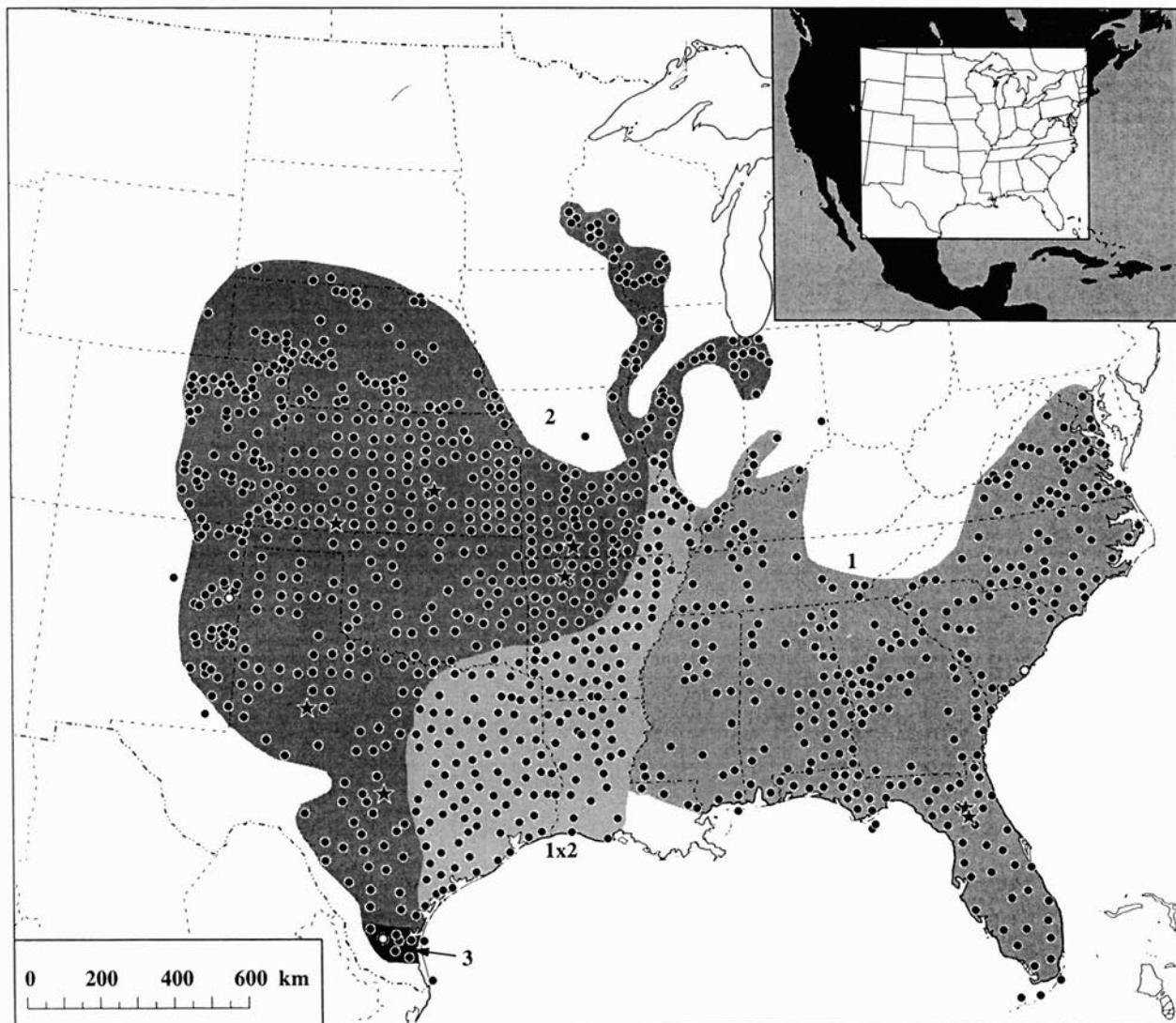


Figure 1. Adult male *Cnemidophorus sexlineatus viridis* (76 mm in SVL) from Izard Co., Arkansas. Photograph by S. E. Trauth.

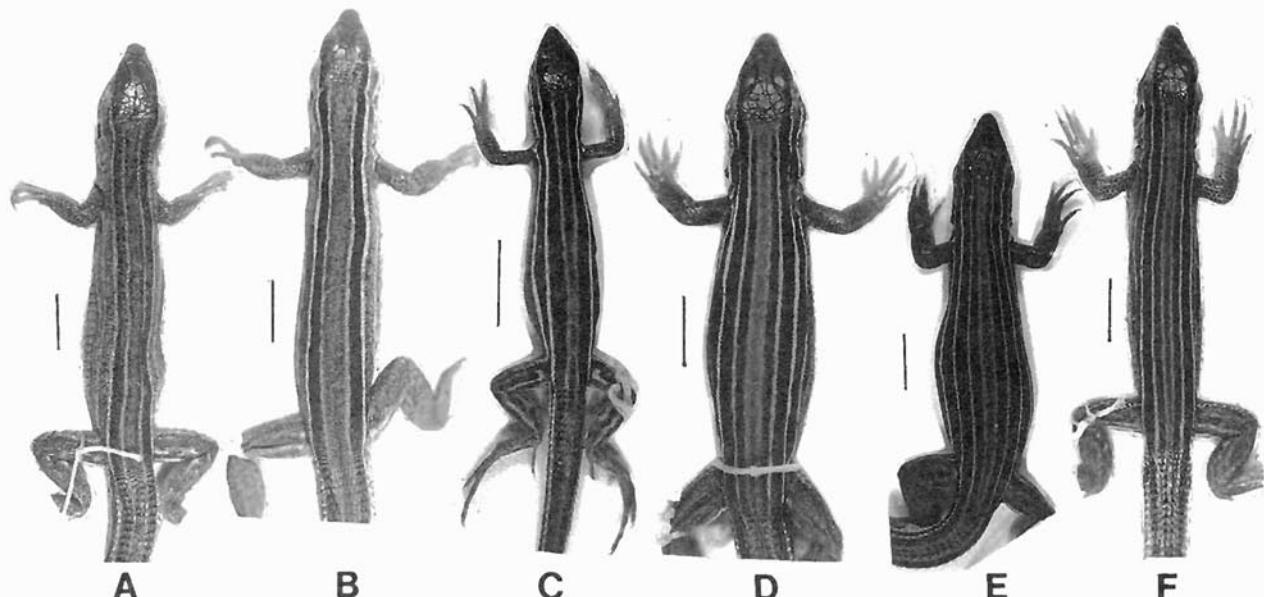
(Barbour, 1971; Cochran and Goin, 1970; Minton, 1972; Mount, 1975; Sellers et al., 1980; Smith, 1946; Trauth, 1980; Wright, 1994); *stephensae* (Trauth, 1992); *viridis* (Anderson, 1965; Baxter and Stone, 1985; Collins, 1993; Collins and Collins, 1991; Fitch, 1958; Hardy, 1962; Lowe, 1966; Lowe et al., 1966; Pough et al., 1989; Smith, 1946; Smith, 1961; Trauth, 1980; Walker et al., 1990a, b). Color photographs of *sexlineatus* were provided by Ashton and Ashton (1991), Behler and King (1979), Leviton (1972), Martof et al. (1980), Palmer and Braswell (1995), and Snyder (1972), and of *viridis* by Busby et al. (1996), Collins and Collins (1993), Conant and Collins (1991), Degenhardt et al. (1996), Garrett and Barker (1987), Hammerson (1982), Johnson (1987), Oldfield and Moriarty (1994), Vermersch (1992), and Vogt (1981). Color illustrations of *sexlineatus* are in Conant (1975), Dundee and Rossman (1989), Mitchell (1994), and Smith and Brodie (1982), and of *viridis* in Stebbins (1985). Line drawings of scutellation were presented by Cope (1892) and Smith (1946); geographic variation in dorsal head scutellation and postantebrachial scales was illustrated by Trauth (1980). Photographs of a nest and eggs are found in Brown (1956). Böhme (1988) illustrated the hemipenes. Newton and Trauth (1992) provided electron micrographs of spermatozoa. The karyotype was illustrated by Bickham et al. (1976) and Lowe

et al. (1970), and that of a triploid laboratory hybrid (*C. neomexicanus* × *C. sexlineatus*) by Dessauer and Cole (1984). Trauth (1988) provided electron micrographs of the eggtooth. Light and electron micrographs of the eggshell are found in Trauth and Fagerberg (1984). Photomicrographs of histological sections of the testes and adrenal glands are found in Brackin (1979). Electrophoretic patterns are available in Dessauer and Cole (1984, 1989) and Wright et al. (1983). Photographs of winter burrows are found in Trauth (1983).

• **Distribution.** *Cnemidophorus sexlineatus* ranges from Maryland to the Florida Keys in the eastern United States, from the upper Mississippi River region (southeastern Minnesota and adjacent Wisconsin) in the north to extreme southern Texas in the south, and from southern South Dakota and southeastern Wyoming through eastern Colorado to eastern New Mexico in the west. The eastern race (*sexlineatus*) occurs in the eastern United States, east of the Mississippi River and south of the Ohio River at their confluence. The western race (*viridis*) occurs in Illinois, northern and southern Indiana, and Wisconsin as well as areas west of the Mississippi River. A broad zone of intergradation exists between *sexlineatus* and *viridis* in the south-central United States (see Map). Burt (1931a), Conant (1958,



**Map.** Distribution of *Cnemidophorus sexlineatus*. Large circles indicate type-localities; dots mark counties of occurrence or specific localities at the edge of the range. Stars indicate fossil records.



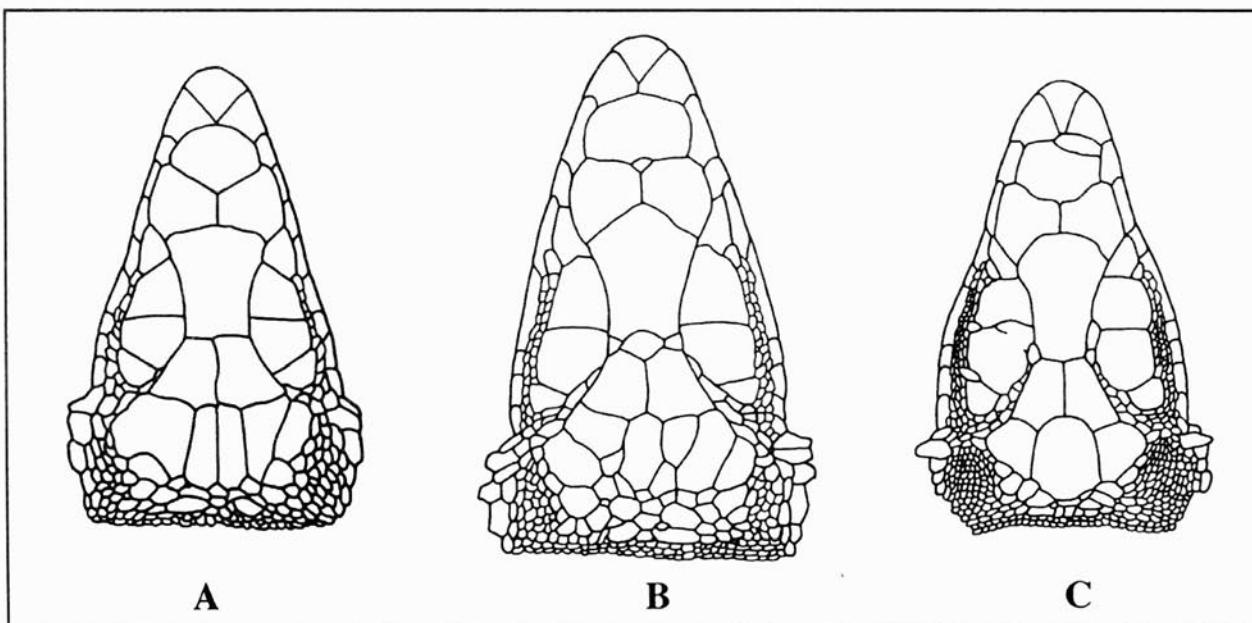
**Figure 2.** Dorsal pattern in *Cnemidophorus sexlineatus*. *C. s. viridis*: A - (male, Bourbon Co., Kansas); B - (male, Baxter Co., Arkansas). *C. s. stephensae*: C - (male, Kenedy Co., Texas). *C. s. sexlineatus*: D - (male, Wake Co., North Carolina); E - (female, Lancaster Co., South Carolina). *C. s. sexlineatus* × *viridis*: F - (male, Cameron Parish, Louisiana). Line = 10 mm. Photographs from Trauth (1980).

1975), Conant and Collins (1991), Smith (1946), Trauth (1980), Vance (1978), and Wright (1994) summarized the distribution in the United States.

*Cnemidophorus sexlineatus* inhabits areas with dry, friable soils comprised mostly of sand, sandy-loam or red clay substrates in which they seek shelter via burrowing; these habitats include sandy areas along and within river floodplains, sand dunes, cedar glades, and rock outcrops. Racerunners are also common in a variety of physically or floristically-altered environments such as along highway and railroad right-of-ways, beneath power lines, the borders of cultivated fields or pastures, and open canopy areas within pine and oak-hickory forests.

• **Fossil Record.** *Cnemidophorus sexlineatus* has been reported from the Pleistocene of Arkansas (Davis, 1973), Florida (Auffenberg, 1956; Gut and Ray, 1963; Martin, 1974), Kansas (Etheridge, 1958, 1960; Holman, 1971, 1979; Rickart, 1976), Texas (Holman, 1966, 1969; Rogers, 1976), and Missouri (Holman, 1974).

• **Pertinent Literature.** Comprehensive works on the biology of *Cnemidophorus sexlineatus* include Burt (1931a), Fitch (1958), and Trauth (1980). Taxonomic and systematic discussions are found in Burt (1931a), Cope (1892), Duellman and Zweifel (1962), Gadow (1903), Trauth (1980), and Wright



**Figure 3.** Dorsal head scutellation of *Cnemidophorus sexlineatus*. A. *C. s. viridis* (USD 3581), Knox Co., Nebraska. B. *C. s. sexlineatus* (USNM 22178), St. Mary's Co., Maryland. C. *C. s. sexlineatus* (AMNH 64548), Highlands Co., Florida.

(1993). Chromosomes were examined by Bickham et al. (1976), Cole (1984), Dessauer and Cole (1984), Lowe and Wright (1966), and Lowe et al. (1970). Electrophoretic patterns and biochemical genetics are discussed in Dessauer and Cole (1984, 1989), Dessauer et al. (1962), Guttman (1971), McKinney et al. (1973), Neaves (1969), Parker and Selander (1976), and Wright et al. (1983). Mitochondrial DNA analyses are detailed in the works of Densmore et al. (1989a, b) and Moritz et al. (1992). Parker (1979) provided detailed morphometric analyses within a phylogenetic framework.

Ecological relationships with unisexual congeners were examined by Bickham et al. (1976), Cole et al. (1988), McKinney et al. (1973), Paulissen et al. (1992), Taylor et al. (1989), Walker (1987), and Walker et al. (1990a, b, 1994). Specific habitat affinities are documented in Anderson et al. (1952), Dodd (1992a), Fouquette and Lindsay (1955), Mushinsky (1985), and Rudolph and Dickson (1990). Discussion of interspecific competition can be found in Bohlen (1976) and Werth (1972). Life history parameters are discussed by Ballinger and Jones (1985), Ballinger et al. (1990), Bellis (1964), Carpenter (1959), Clark (1976), Fitch (1958), Hardy (1962), Mushinsky (1985), Paulissen (1988c), Walker (1964), and Werth (1969, 1972). Relationships among life-history attributes including growth pattern, survival rate, and age of maturity are reported in Shine and Charnov (1992). The effects of habitat alteration are mentioned in Means and Campbell (1981). Symbiotic relationships are discussed by Lips (1991), Pearson and Nelson (1952), and Witz et al. (1991). Predation by the red imported fire ant was discussed in Mount et al. (1981), whereas other instances of predation were discussed in Camper (1986) and Whiting et al. (1991). Field marking techniques include the use of bird bands (Paulissen, 1986) and fluorescent dyes (Dodd, 1992b). Durtsche (1996) presented a novel capture technique. Aquatic habits were documented by Dillon and Baldauf (1945), Stille (1947), and Trauth et al. (1996). Arboreality was observed by Paulissen and Harvey (1985). Dietary preferences, feeding activity, feeding behavior, and nutrition are detailed in Burt (1928), Dearing (1993), Hamilton and Pollack (1961), Harney (1953), McCauley (1939), Paulissen (1987a, b), Pough (1973), Pritchett (1903), Punzo (1990), and Witz and Lawrence (1993). Whole-body protein content was reported by Boyd and Goodyear (1971). Cannibalism was reported in Etheridge and Wit (1982). Hatch and Stein (1994) reported that adults will eat eggs laid in captivity. Ballinger et al. (1992) discussed chemosensory searching. Hibernation, hibernation denning sites, and overwintering of eggs are described in Etheridge et al. (1983, 1986), Sellers et al. (1982), Smith et al. (1965), Trauth (1974, 1977, 1983), Trauth and Mount (1978), and Wit and Sellers (1993). Data on eggs, nests, and nesting habitat are presented in Ballinger and Clark (1973), Brown (1956), Carpenter (1960a), Mount et al. (1981), Mullen et al. (1984), and Trauth (1977, 1983). The female reproductive cycle and clutch characteristics are reported in Carpenter (1960a), Clark (1976), Etheridge et al. (1986), Fitch (1970, 1985), Hoddenbach (1966), and Trauth (1983). The male reproductive cycle was examined in Brackin (1979), Etheridge et al. (1986), and Johnson and Jacob (1984). Clark et al. (1995) examined contamination by pesticides. Witz (1996) presented a collecting technique.

Mating behavior is presented in Noble and Bradley (1933). Social behavior and activity patterns were discussed by Ayres (1973), Barden (1942), Brackin (1978), Carpenter (1960b, 1962), Etheridge and Wit (1993), Hardy (1962), Leuck (1980, 1982, 1985), and Paulissen (1988a). Gove (1979) discussed tongue-flicking behavior.

Requirements regarding ultraviolet light are presented in Townsend and Cole (1985). Brackin (1979) and Grassman and Hess (1992a, b) addressed adrenal gland activity. Sellers et al.

(1980) discussed methods of blood collection. Body temperature relationships were presented by Brattstrom (1965), Edgren (1955), Fitch (1956), McKenna and Packard (1975), Paulissen (1988b), and Ragland et al. (1981). Discussion of locomotion, including running speed, sprint performance, and energetics, can be found in Ballinger et al. (1979), Dodd (1993), and White and Anderson (1994), respectively. Pituitary activity is documented in Johnson and Jacob (1984). Guillette et al. (1991) induced oviposition with the use of prostaglandins.

Ectoparasites were studied by Farner and Seaman (1946), Loomis (1956), and Loomis and Crossley (1963), whereas descriptions of endoparasites can be found in the studies of Brooks and Mayes (1976), Dollahon and Janovy (1971), Dyer (1971), Harwood (1932), Hughes et al. (1941), McAllister et al. (1991a, b), Moskowitz (1951), Shoop and Janovy (1978), and Zimmerman and Brown (1952).

Morphology has been studied as follows: adrenal gland and testicular histology (Brackin, 1979), mesonephros-adrenal association (Hardy and Cole, 1981), eggshell (Trauth and Fagerberg, 1984, 1993), eggtooth (Trauth, 1988), oral anatomy and teeth (Presch, 1974; Taylor, 1940), spermatozoan ultrastructure (Newton and Trauth, 1992; Trauth and Buchanan, 1996), circum-testicular tunic (Lowe and Goldberg, 1966), thyroid gland (Lynn et al., 1966), and ultimobranchial bodies (Sehe, 1965). Sex identification using scutellation was presented by Pietruszka (1981). Sexual dimorphism was examined by Trauth (1980) and Fitch (1981). Maslin (1967) described skin grafting techniques.

Annotated bibliographies which contained *Cnemidophorus sexlineatus* include Carpenter and Krupa (1989), Dixon (1993), Enge and Dodd (1986), Lohofener and Altig (1983), Price (1983), and Vance (1985). Regional summaries occur for the following states: Alabama (Mount, 1975), Colorado (Hammerson, 1982), Florida (Ashton and Ashton, 1991), Illinois (Smith, 1961), Indiana (Minton, 1972), Kansas (Collins, 1974, 1982, 1993), Kentucky (Barbour, 1971), Louisiana (Dundee and Rossman, 1989), Minnesota (Oldfield and Moriarty, 1994), Missouri (Anderson, 1965; Johnson, 1987), Nebraska (Lynch, 1985), New Mexico (Degenhardt et al., 1996), North Carolina (Palmer and Braswell, 1995), Oklahoma (Webb, 1970), Tennessee (Snyder, 1972); Texas (Dixon, 1987), Virginia (Mitchell, 1994; Tobey, 1985), Wisconsin (Vogt, 1981), and Wyoming (Baxter and Stone, 1985). Additional noteworthy distributional or ecological information is available in Burt (1935) and Minton and Minton (1948), and for the following states: Alabama (Jackson and Jackson, 1970 [1971]), Arkansas (Dellinger and Black, 1938; Dowling, 1957; Schwart, 1938; Taylor, 1935), Florida (Blaney, 1971; Franz, 1995; Funderburg and Lee, 1968; Greenberg et al., 1994; Lee, 1968), Georgia (Martof, 1963), Illinois (Cagle, 1942; Munyer and Parmalee, 1967), Kansas (Burt, 1927; Busby and Parmalee, 1996; Clarke, 1958, 1965; Collins and Collins, 1991), Missouri (Hurter, 1893), Nebraska (Hudson, 1972), New Mexico (Lowe, 1955), North Carolina (Engels, 1942, 1952; Lewis, 1946), Oklahoma (Ortenburger and Freeman, 1930), Tennessee (Savage, 1967), Texas (Strecker, 1902, 1909, 1922, 1929; Strecker and Williams, 1928; Vermersch, 1992), Tennessee (Blanchard, 1922), and Virginia (Richmond and Goin, 1938).

• **Nomenclatural History.** As with many other members of "Cope's most difficult genus" (Lowe, 1993), the taxonomic history of *Cnemidophorus sexlineatus* is replete with misapplied names. The early synonyms were generated prior to the general acceptance of Darwin's work placing biological species within a phylogenetic framework, and largely resulted from differing interpretations of systematic classification at levels above the species category. *Cnemidophorus sexlineatus* became taxonomically intertwined with *C. gularis* and *C. perplexus* following

the description of the latter two taxa by Baird and Girard in 1852. Cope (1866 [1867]) considered *gularis* and *sexlineatus* to be conspecific, an arrangement followed by most other workers (e.g., Bocourt, 1874; Boulenger, 1885; Cope, 1875, 1883, 1898 [1900]; Coues, 1875; Günther, 1885; Strecker, 1902, 1909, 1922; Yarrow, 1875, 1882 [1883]; but not Cope, 1892) for almost a century. Cope (1892, 1898 [1900]) considered *perplexus* a subspecies of *tessellatus* (= *tigris* Burger, 1950), but Brown (1903) and Smith and Taylor (1950a) suggested a close relationship between *sexlineatus* and *perplexus*. Burt (1931a, b) recognized distinct character differences between *sexlineatus* and *gularis*, but remained unconvinced of the distinctness of the two taxa and stated that both *sexlineatus* and *perplexus* possessed close affinities with *gularis*. He codified this view by treating the three as subspecies of *C. sexlineatus* in his classic treatise on the genus *Cnemidophorus*. Although some workers (e.g. Gadow, 1906; Strecker, 1930) recognized the fact, it remained for Taylor (1936) to explicitly make clear the distinction between *sexlineatus* and *gularis*. Schmidt and Smith (1944) recognized that *gularis* was distinct from *perplexus*, and Wright (1969) has detailed the confusing misapplications of the latter name. Only with the advent of sophisticated univariate and multivariate statistical analyses of character variation in large numbers of specimens from throughout the ranges of various taxa and the application of molecular genetic techniques to systematic problems in the genus *Cnemidophorus* have the taxonomic relationships between *gularis*, *perplexus*, and *sexlineatus* been solidly established.

• **Etymology.** The name *sexlineatus* (L. "sex" = six or six-fold; L. "lineatus" = streaked or marked with lines) refers to the number of longitudinal stripes running the length of the body. The name *viridis* (L. "viridis" = green) alludes to the characteristic body color of this subspecies. The matronym *stephensae* honors Hazel J. Stephens Hickey for her financial support toward the herpetological endeavors of the senior author.

• **Comment.** Trauth (1980) noted the distinctiveness of populations of *C. sexlineatus* in Florida and Texas, and recently (Trauth, 1992) designated a new race from Texas (see below). The diagnostic traits for the new Texas race were not applicable to the undescribed Florida race. This latter form possesses extremely high scale counts for several characters (e.g., Figure 3C) and is generally restricted to central peninsular Florida and several coastal scrub habitats. Zones of intergradation between the new forms and the other races remain to be established. Trauth (1980) postulated that Florida was the evolutionary center (storehouse of genetic variation) for the species because of the range of variation of several character states in Florida.

### 1. *Cnemidophorus sexlineatus sexlineatus* (Linnaeus) Six-lined Racerunner

*Lacerta sexlineata* Linnaeus, 1766:364. See species synonymy. *Cnemidophorus sexlineatus*: Duméril and Bibron, 1839:131. See species account.

*Cnemidophorus sexlineatus oligoporus*: Hoffman, 1957a:153.

*Cnemidophorus s.[exlineatus] sexlineatus*: Hoffman, 1957a:154.

First use of trinomial.

*Cnemidophorus sexlineatus pauciporus*: Hoffman, 1957b:423.

• **Definition.** This subspecies is characterized by six well-defined longitudinal light stripes. An additional vertebral stripe may be present, and it may be single, thinly-divided, or separated into two or more weakly-defined stripes. The latter condition results from expansion and separation of the vertebral stripe and occurs mostly in southeastern populations. The ver-

tebral stripe is broad in *sexlineatus* × *viridis* intergrades. The ground color is dark brown or black. The face and neck are blue. Lateral supraocular granules average 28 (12-62), circumorbital granules average 11 (4-29), and granules from occiput to rump average 218 (182-267).

• **Remarks.** Hoffman (1944, 1949) reported on the phenotypic variation in *sexlineatus* in the eastern United States and noted subdivision of parietal scales; later, Hoffman (1957a, b) named a new subspecies, *pauciporus* (= *oligoporus*), characterized by high parietal counts and reduced femoral pore number (mostly less than 30). The range of *pauciporus* was given as Maryland, Virginia, North Carolina, and northern South Carolina. Duellman and Zweifel (1962) noted that scales other than the parietals (e.g. frontoparietals and supraoculars) are often subdivided in *sexlineatus* and, by showing that femoral pore counts in *sexlineatus* elsewhere in its range were below 30, refuted the recognition of *pauciporus*. Trauth (1980) showed that supernumerary frontoparietals occurred in Alabama, Georgia, and Florida populations.

### 2. *Cnemidophorus sexlineatus viridis* Lowe Prairie Racerunner

*Cnemidophorus sexlineatus viridis* Lowe, 1966:44. Type-locality, "7.6 mi. S Tucumcari, along state rd 18, Quay County, New Mexico." Holotype, University of Arizona, Department of Zoology (UAZ) 14800, an adult female, collected by C.H. Lowe on 13 August 1949 (not examined by authors).

• **Definition.** This subspecies possesses six well-defined longitudinal stripes and an additional vertebral stripe that is single or narrowly divided. The vertebral stripe is broad in *sexlineatus* × *viridis* intergrades. The ground color is mostly green. The face and neck are bluish-green. Lateral supraocular granules average 21 (5-36), circumorbital granules average 9 (1-12), and granules from occiput to rump average 192 (155-240).

• **Remarks.** Trauth (1980) examined geographic variation in morphological characters of *C. sexlineatus* from throughout its range. He found that this subspecies exhibits a broad contact zone with that of the eastern form (see Map). Intergrade populations possess an array of intermediate meristic counts as well as body coloration and striping patterns. Trauth (1980) also noted that 15 of 18 characters examined varied clinally. Clines were usually oriented in a northwest to southeasterly direction with meristic counts concordant from smaller to larger, respectively.

### 3. *Cnemidophorus sexlineatus stephensae* Trauth Texas Yellow-faced Racerunner

*Cnemidophorus sexlineatus stephensi* Trauth 1992:438. Type-locality, "4.8 km S Hebbronville, Jim Hogg Co., Texas." Holotype, National Museum of Natural History (USNM) 278270, an adult male, collected by S.E. Trauth on 18 May 1983 (examined by authors).

*Cnemidophorus sexlineatus stephensae*: Trauth, 1995:68. Emendation.

• **Description.** This subspecies is characterized by the following combination of features: small adult body size (maximum SVL 68 mm), absence of a vertebral stripe (a vestige may be present in the region of the nape or on the anterior vertebral field in some specimens), and yellow coloration on the face and lateral surfaces of the neck in both adults and juveniles.

• **Remarks.** The range of this subspecies as illustrated by Trauth (1992) is generally restricted to the South Texas Plains, a mostly mesquite-live oak savanna characterized by moderately tall, white-to-brown colored sand dunes in the eastern portion to a region of low red-clay sand dunes in the western portion (Mahler, 1981). Trauth (1992) also found this subspecies on South Padre Island.

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