

**CLONING OF WHEAT GERM EUKARYOTIC INITIATION FACTOR EIF2**

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## Abstract

Eukaryotic Initiation Factor 2 (eIF2) is a protein complex found in eukaryotes involved in the initiation of translation. eIF2 consists of three non-identical subunits ( $\alpha$ ,  $\beta$ , and  $\gamma$ ) and is required for the translation of virtually all mRNA transcripts. eIF2 forms a ternary complex with GTP and Met-tRNA<sub>i</sub><sup>Met</sup>, which then associates with the 40S ribosomal subunit. This 43S preinitiation complex scans the mRNA transcript for the AUG initiation codon, at which time eIF2's GTP is hydrolyzed to GDP and eIF2-GDP dissociates from the 40S subunit. A guanine nucleotide exchange factor called eIF2B is required to reform eIF2-GTP, which can now facilitate a new round of translation initiation.

In many eukaryotic systems, eIF2 has been documented as a site of regulation of translation via phosphorylation at a conserved serine residue on the  $\alpha$  subunit. These studies have found that phosphorylated eIF2 (eIF2[ $\alpha$ P]) binds eIF2B with a much higher affinity than non-phosphorylated eIF2. Since eIF2B is typically present at less than half the cellular concentration of eIF2, the result of this strong binding is sequestration of eIF2B and inhibition of its nucleotide exchange activity. Because only eIF2-GTP can bind Met-tRNA<sub>i</sub><sup>Met</sup>, the result is inhibition of eIF2's ternary complex formation, and thus inhibition of protein synthesis.

Most studies exploring this mechanism of translational control have focused on yeast and mammalian systems. However, in higher plants, regulation of translation by phosphorylation of eIF2 $\alpha$  has not been shown to occur in vivo to date. Although homologous genes of all five eIF2B subunits have been identified in plants, the eIF2B protein complex has not yet been isolated from plants. Similarly, though a gene with homology to an eIF2 $\alpha$ -kinase from yeast (*gcn2*) has been identified in plants,

phosphorylation of eIF2 in a plant translational lysate system has not yet been shown to inhibit translation. The difference of the binding affinity of eIF2 for GDP and GTP is not nearly as severe as that found in other eukaryotic systems, suggesting that a recycling factor (eIF2B) and phosphorylation of eIF2 $\alpha$  may play a less critical regulatory role in plants.

There are significant differences between plants and animals in their responses to environmental stress, and therefore methods of translational regulation may also be different. In order to explore the extent of regulation via phosphorylation of eIF2 $\alpha$  in plants, and to uncover other plant-specific regulation on eIF2, it is necessary to be able to express the eIF2 complex in a manipulable bacterial system. This will allow mutagenesis of the three subunits to probe structure and function. This thesis reports the use of recombinant methods to construct a single expression vector containing all three genes of eIF2 from *T. aestivum* (wheat), with the goal of subsequent expression of eIF2 $\alpha$ ,  $\beta$ , and  $\gamma$  as a functional complex in *E. coli*.

## Background

### Transcriptional vs Translational Regulation

A cell is defined in its function and in its identity by the constellation of proteins it produces, maintains, and secretes. The eukaryotic cell devotes a large portion of its total energy expenditure into the selective synthesis of proteins. For these reasons, protein expression is extensively regulated.

It is economically sensible for a cell to control any process at the initial step in order to preserve its limited energy and materials and, in fact, regulation of gene transcription is the most vastly prevalent and documented type of regulation in the entire pathway from gene to protein. Transcriptional control has the additional property of being highly selective, determining the specific array of proteins that the cell produces, thus conferring identity on to the cell.

However, there are several instances in which a cell might benefit by exhibiting control at the level of translation initiation. By regulating this step, a cell can respond more immediately to a particular physiological stimulus. If the cell senses an environmental stress, it can step down the rate of translation in order to preserve resources until the stress is removed and materials return to abundance. For example, virus-infected cells and cells subjected to a heat-shock treatment both exhibit translational repression. Additionally, some special systems such as early-stage embryos and reticulocytes both rely heavily on translational control, as there is an absence of any gene transcription (Mathews *et al.* 2000).

Regulation at the level of translation tends to have broad and even global effects, affecting the translation of a specified group of mRNAs, or the entire transcriptome

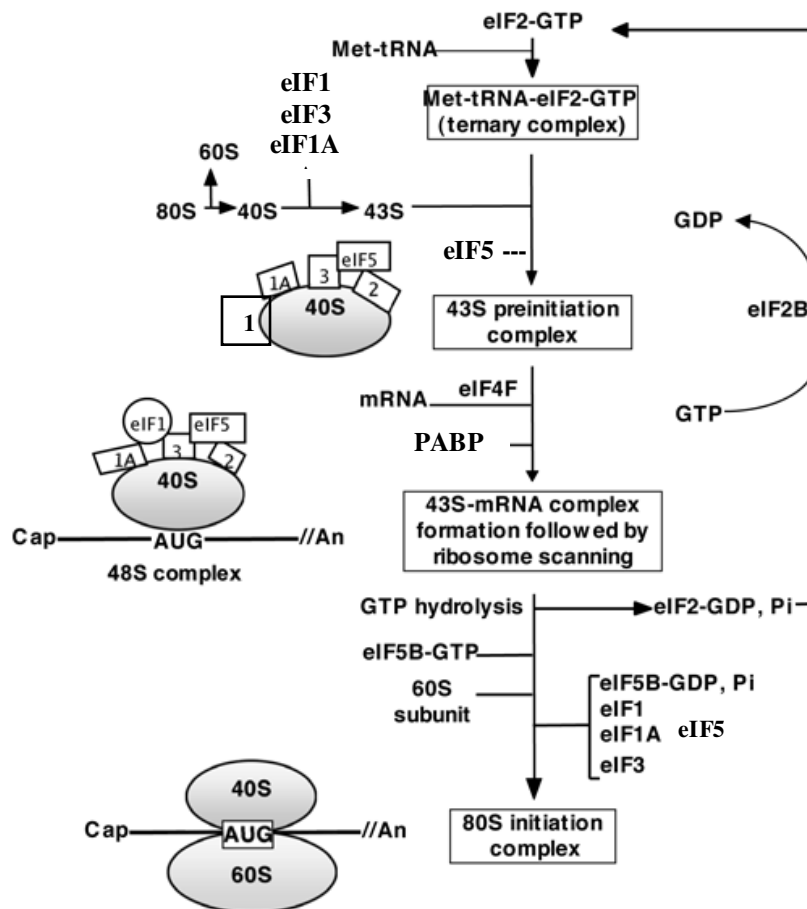
present in the cell cytoplasm. For this reason translational control often reveals something about the total rate of protein synthesis, which in turn can be a measure of the rate of metabolic activity, growth, and reproduction of a cell. Thus, it is important in theory and in practice to understand the mechanisms of translational control, for this is to understand what kinds of cues can increase or decrease protein synthesis and cell growth. The applications – from agricultural yield maximization to cancer therapy – are boundless.

The phenomenon of regulation at the level of translation is most widely appreciated and documented in eukaryotes (Pestova *et al.* 2007). This likely results from the spatial and temporal separation of transcription and translation in eukaryotes, which makes translational control much more necessary and useful. Translational control is most often implemented during the initiation phase of translation, by affecting the activity of an initiation factor via phosphorylation or another mechanism. These two observations warrant a detailed account of eukaryotic initiation of translation, in order to discuss the key sites and mechanisms targeted by translational regulators.

### **The Mechanism of Eukaryotic Translation Initiation**

Eukaryotic translation initiation involves the assembly of the mRNA and Met-tRNA<sub>i</sub><sup>Met</sup> on the ribosome, followed by ribosomal scanning of the mRNA to locate the start codon. This process heavily depends on thirteen eukaryotic translation initiation factors known as eIFs (Figure 1). Before initiation can begin, the two subunits of the ribosome, usually found in association, must be separated from each other in order to allow the build up of the preinitiation complex on the 40S small ribosomal subunit. This

is accomplished by eIF3, eIF1, and eIF1A, which bind the 40S subunit and may hinder its interaction with the larger subunit (Hershey and Merrick 2000). eIF3 is a large complex that consists of thirteen polypeptides. eIF1 and eIF1A are both single polypeptide chains. It is not clear whether these three initiation factors remain associated with the 40S ribosomal subunit throughout initiation or whether they dissociate at some intermediate step. In any case, they have several important roles in later stages of initiation.



**Figure 1. Mechanism of Eukaryotic Translation Initiation (Adapted from www.scielo.cl).**

The first step of initiation is the formation of the ternary complex (consisting of eIF2, GTP, and Met-tRNA<sub>i</sub><sup>Met</sup>) whose function is to deliver the Met-tRNA<sub>i</sub><sup>Met</sup> to the 40S

ribosomal subunit. eIF2 is a heterotrimeric protein complex consisting of  $\alpha$ ,  $\beta$ , and  $\gamma$  subunits. Of these, it is eIF2 $\gamma$  that binds Met-tRNA<sub>i</sub><sup>Met</sup> (assisted by eIF2 $\beta$ ) via a particular A:U base pair (A1:U72) of the tRNA<sub>i</sub><sup>Met</sup>'s acceptor stem (Pestova *et al.* 2007). eIF2 $\gamma$  also binds GTP and is the catalytic subunit with GTPase activity. It is important to note that eIF2-GDP cannot complex with Met-tRNA<sub>i</sub><sup>Met</sup>, and hydrolysis of GTP bound to eIF2 causes a decreased affinity between eIF2 $\gamma$  and Met-tRNA<sub>i</sub><sup>Met</sup> (Hinnebusch 2000). After the ternary complex is formed, an association between eIF3 and eIF2 helps bring the ternary complex to the 40S ribosomal subunit (Hershey and Merrick 2000). eIF5, a GTPase-activating protein (GAP) specific to eIF2, is also recruited at this time via interactions with eIF2 $\beta$  and eIF3.

At this stage the 40S ribosomal subunit, ternary complex, and all other associated translation factors are known collectively as the 43S preinitiation complex. The next step is the association of the 43S preinitiation complex with the mRNA transcript, facilitated by the eIF4 group of initiation factors and poly-A binding protein (PABP). eIF4F, also known as the cap-binding complex, is made up of three proteins in mammals: eIF4E, eIF4G, and eIF4A. eIF4E recognizes and binds the 7-methyl-guanosine cap at the 5' end of the mRNA (Hershey and Merrick 2000). This interaction is enhanced by binding of eIF4E by eIF4G, a large protein that plays important structural and stabilizing roles by causing conformational changes in the mRNA as well as protein factors that it binds. eIF4G also binds eIF4A, an ATP-dependant RNA helicase, effectively localizing the assembly of translation factors at the 5' end and promoting the helicase activity. Additional factors eIF4B and eIF4H increase the helicase's processivity. PABP, by binding the 3' poly-A tail and also the cap binding complex, creates a circular mRNA,

adding to the mRNA's stability (Hershey and Merrick 2000). Although it is known that eIF4G binds eIF3 and therefore that this interaction must assist the 43S preinitiation complex in loading onto the mRNA, it is not known whether the 43S complex initially associates exactly on or somewhat downstream of the 5' cap and associated factors. In the absence of any secondary structure in the 5' leader of the mRNA transcript, the 43S preinitiation complex is able to associate with the mRNA and identify the initiation codon without the presence of eIF4F, eIF4B, or PABP; however, these factors become required if the 5' UTR is even slightly structured (Pestova *et al* 2007).

It is important to note here that most members of the cap-binding complex are targets of translational control. eIF4H, eIF4B, eIF4E, eIF4G, and PABP can all be phosphorylated at several locations. Generally, the degree of phosphorylation positively correlates with the rate of translation (Gallie 2007).

These two steps have accomplished the binding of the Met-tRNA<sub>i</sub><sup>Met</sup> and the mRNA to the 40S ribosome. Although there is some evidence that these events occur in the order indicated, theoretically the mRNA may bind first, or else they may be occurring at more or less the same time (Hershey 2000). The initiation complex is now ready to scan the mRNA for the initiation codon. This process requires eIF4F unless there is no secondary structure present in the 5' leader. During scanning eIF4F maintains its association with eIF3, dissociating with the cap and causing the 5' leader end to feed out of the looped mRNA.

As noted above, the 43S preinitiation complex loads onto the mRNA at the 5' end, assisted by associations with the cap-binding complex. The 43S preinitiation complex then moves downstream along the mRNA until it encounters an AUG codon in



favorable context (Kozak 2002). The interaction between the mRNA AUG start codon and the tRNA<sub>i</sub><sup>Met</sup> UAC anticodon is the primary facilitator of initiation site recognition, at which point GTP is hydrolyzed (Pestova *et al* 2007). This linear scanning mechanism is supported by experiments showing that if an AUG is inserted upstream of the natural start codon, initiation will begin at the newly inserted AUG sequence (Kozak 2002). eIF1 is critical to the fidelity of the codon-anticodon interaction, preventing partial pairing between an incorrect mRNA codon with the tRNA<sub>i</sub><sup>Met</sup> anticodon which would result in false recognition and premature initiation of polypeptide synthesis (Pestova *et al* 2007). eIF1 also checks that the AUG start codon is in a good context on the mRNA, which entails an A or G at the -3 position of the mRNA, a G at the +4 position, and a 5' leader of at least 8 ribonucleotides. eIF2 and eIF5 have also been implicated in the fidelity of start codon recognition, based on evidence that eIF2 $\beta$  interacts directly with the mRNA and that mutations in eIF2 and eIF5 have been shown to allow an upstream UUG to start translation instead (Pestova *et al* 2007).

When the start site is recognized, the binding energy between the start codon and the tRNA<sub>i</sub><sup>Met</sup> anticodon pair causes the complex to stall, and the 40S ribosome switches from an open conformation to a closed conformation involving the rearrangement of the Met-tRNA<sub>i</sub><sup>Met</sup> and the mRNA. At this point, eIF5 stimulates eIF2 $\gamma$  to hydrolyze its bound GTP, causing a weakened interaction between Met-tRNA<sub>i</sub><sup>Met</sup> and eIF2. The energy from this GTP hydrolysis was initially thought to cause all factors to immediately dissociate (Hershey and Merrick 2000); however, this hypothesis has now been reinvestigated to show that rather than immediate dissociation, the factors are displaced from their original positions but remain associated (Pestova 2007). A factor called eIF5B

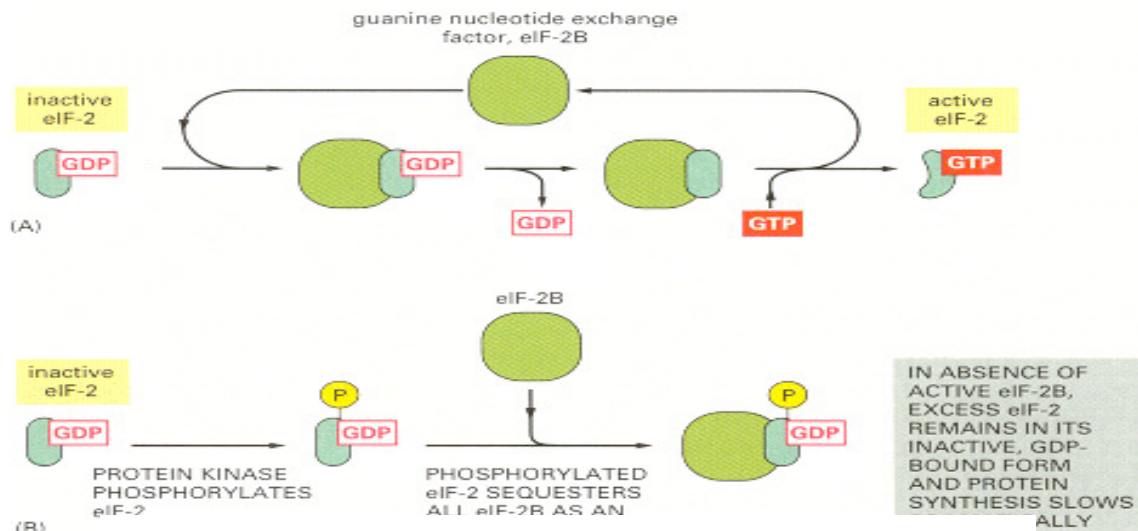
(a G protein) is then recruited and facilitates the displacement of all other translation factors. Finally, eIF5B itself dissociates using the energy of its own bound GTP, leaving the mRNA transcript and Met-tRNA<sub>i</sub><sup>Met</sup> attached to the 40S small ribosomal subunit, now with a bare interaction face which can readily associate with the 60S large ribosomal subunit to form the active 80S ribosome.

At this point, initiation is now complete, and elongation of the amino acid chain commences via the functioning of the active ribosome and elongation factors. However, in order to begin a new round of translation initiation, eIF2, currently associated with GDP after start-site dependent hydrolysis stimulated by eIF5, must be recycled to its active state, eIF2-GTP. As noted above, only eIF2-GTP can bind the Met-tRNA<sub>i</sub><sup>Met</sup> in order to form the ternary complex. The guanine exchange activity is carried out by eIF2B, a heteropentameric guanine nucleotide exchange factor (GEF) specific to eIF2.

### **Translational Control via eIF2 $\alpha$ Phosphorylation**

In response to certain environmental stresses, eIF2 $\alpha$ -kinases are activated and phosphorylate eIF2 $\alpha$  (yielding eIF2[ $\alpha$ P]) at a conserved residue: Ser-51 (Hinnebush 2000). When phosphorylated at this position, the binding affinity between eIF2 and eIF2B increases greatly, so that eIF2 acts as an inhibitor instead of a substrate. Because the cellular concentration of eIF2 is typically two to five times as high as the cellular concentration of eIF2B (Hinnebush 2000), the effect of this strong binding is that all eIF2B becomes bound in non-functional complexes. This sequestration of eIF2B occurs even when only a fraction of the eIF2 molecules present are phosphorylated. The sequestration and inhibition of eIF2B prevents eIF2 recycling, which in turn causes a

global inhibition of translation in the cell (Figure 2).



**Figure 2. Mechanism of Translational Control via eIF2 $\alpha$  Phosphorylation.**  
**From Albert's Molecular Biology of the Cell, 5<sup>th</sup> ed.**

This recycling mechanism is a central point of translation inhibition, as has been demonstrated mainly in mammalian and yeast systems. There are four well characterized eIF2 $\alpha$ -kinases studied to date (Dever *et al.* 2007). Protein Kinase R (PKR) is activated in response to viral infection via a binding domain for dsRNA; it is found in all vertebrates. PKR-like ER kinase (PERK) is present in the ER lumen of invertebrates and vertebrates and is activated in response to ER stress, as part of the unfolded protein response. GCN2 is activated in response to amino acid starvation by binding uncharged tRNAs; it has been found in all eukaryotes. Heme-regulated kinase (HRI), found in vertebrates, some yeasts, and some insects, is repressed by heme binding and activated in the absence of heme. In response to these kinases, the downstream effects in all four cases are the same: inhibition of GDP/GTP recycling by eIF2B and repression of translation (Dever *et al.* 2007).

In order to understand the nature of the interaction between eIF2 and eIF2B, it is necessary to first examine the structure and functional role of their subunits. The functions of eIF2's three subunits have been noted above as their role in translation initiation arises but are repeated here for detail and clarity. eIF2 $\alpha$  is primarily a regulatory subunit, containing the site of phosphorylation (Serine-51) causing translational inhibition. Yeast eIF2 $\alpha$  contains other serine residues (positions 292, 294, and 301) that are phosphorylated by Casein Kinase II (Pestova *et al.* 2007), and plant eIF2 $\alpha$  is also phosphorylated by CK2 (Dennis 2008). However, these have not been linked directly to translational inhibition. eIF2 $\gamma$  has GTPase activity and binds both GTP (through a G-domain) and Met-tRNA<sub>i</sub><sup>Met</sup>, which it recognizes through the A1:U72 base pair in the tRNA<sub>i</sub><sup>Met</sup>'s acceptor stem. eIF2 $\beta$  contains three strands of lysine residues, known as K-boxes, near its amino-terminal domain that serve as the binding site for eIF5 and eIF2B. eIF2 $\beta$  also assists in Met-tRNA<sub>i</sub><sup>Met</sup> binding and interacts directly with the mRNA transcript, suggesting it plays a role in AUG start-codon recognition (Hinnebush 2000).

eIF2B has five subunits, three of which are regulatory ( $\alpha$ ,  $\beta$ , and  $\delta$ ) and two of which are catalytic for the GEF activity (mainly  $\epsilon$ ; stimulated and enhanced by  $\gamma$ ). The essential interaction between the two translation factors occurs between eIF2B $\epsilon$  and eIF2 $\beta$  (Hinnebush 2000). A sequential mechanism for the guanine exchange reaction has been proposed, which suggests an intermediate quaternary complex which initially consists of eIF2B-GTP and eIF2-GDP, and subsequently rearranges to eIF2B-GDP and eIF2-GTP. The regulatory subunits of eIF2B (primarily  $\alpha$ ; stabilized by  $\beta$  and  $\delta$ ) form a pocket in which the phosphorylated arm of eIF2[ $\alpha$ P] can settle; it is this interaction that

creates high-affinity binding leading to eIF2B inhibition and translational repression. This model is supported by experiments involving mutations near eIF2 $\alpha$ 's Ser-51 that block translational inhibition by eIF2 $\alpha$  phosphorylation: these mutations do not block phosphorylation; rather, they weaken the interaction between eIF2[ $\alpha$ P] and eIF2B, thus preventing sequestration of eIF2B (Hinnebush 2000).

### **Translational Control in Plants**

Translational control in plants is an important area of study for many reasons, including applications to agriculture as well as comparative studies across eukaryotes. These comparative studies are important for understanding the evolutionary modification of translational control to suit the particular lifestyles of plants, animals, and fungi. In addition, the plant translational apparatus has benefits as a choice model for general eukaryotic translation because of easy manipulability from the molecular to the organismal level. Because of their autotrophic, sedentary lifestyle, plants have developed various molecular responses to environmental stress, whereas animals can respond to a given number of stresses simply by moving or finding another food source (Gallie 2007). Specifically, two plant species are typically used to study translational control:

*Arabidopsis thaliana*, because of its sequenced genome and familiarity; and *Triticum aestivum* (wheat) because of its immediate agricultural significance, and because of the ability to easily produce an *in vitro* translation system from wheat germ.

All eukaryotic translation initiation factors discussed above arose before the divergence of the eukaryotic kingdoms and are present in animals, plants, and fungi. Most likely these initiation factors are linked to the evolutionary shift from translation

mediated by the Shine-Dalgarno sequence (prokaryotic) to 5' 7-methyl guanosine cap-dependant translation initiation (eukaryotic). Although the mechanism of translation initiation, elongation, and termination in plants is largely equatable with these processes in all other eukaryotes, it is uncertain how similar are the mechanisms of translational control between plants and the other eukaryotic kingdoms. Here the idiosyncrasies of the plant translational apparatus are presented.

The details and control of the cap-binding complex in plants differs from that in yeast and mammalian systems. Plants possess two isoforms of the cap-binding complex. The first is named eIF4F, composed of eIF4E and eIF4G; the second is named eIFiso4F, composed of eIFiso4E and eIFiso4G (Browning 1996). Both complexes combine with eIF4A. Plant eIF4F shares more sequence homology with mammalian eIF4F, whereas eIFiso4F has a slightly further evolutionary distance (Browning 1996). There exists a phosphorylated form of eIF4E/iso4E that is specific to plants and only active in rapidly growing root and shoot tips of young plants. In addition to this phosphorylated form of eIF4E, plants also express nCBP, a novel cap binding protein that shares similarity in sequence and function to eIF4E/iso4E. Because the cap-binding complex, and specifically eIF4E/iso4E, is responsible for binding and recognizing the mRNA transcript, the existence of several isoforms and phosphorylation states of eIF4E in plants suggests a mechanism of selective translation of separate classes of mRNA by the separate isoforms of the cap-binding complex.

Plant eIF4E/iso4E lack the conserved Ser-209 that in other systems affect its cap-binding affinity. In addition, 4E-BP, which inhibits the association of 4E with 4G, has not been isolated in plants. All of these findings demonstrate repeatedly that the

mechanism of the cap binding complex's activity and regulation differs significantly between plants and other eukaryotic systems, though many details remain to be uncovered.

### **Is Regulation via eIF2 $\alpha$ Phosphorylation Functional in Plants?**

There is evidence that the regulation of eIF2 activity via phosphorylation of eIF2 $\alpha$ 's Ser-51 residue may not be active in plants, or that it may not lead to as severe an inhibition of translation as occurs in other eukaryotic systems (Browning 2004). First, it has been shown that the dissociation constant for eIF2-GTP is only ten times higher than that for eIF2-GDP in plants (Shaikhin *et al* 1991). In contrast, the mammalian system shows as much as a 100-fold difference in the dissociation constant between the two binary complexes. As the dissociation constant measures the propensity for dissociation, this suggests that plant eIF2-GDP may be able to dissociate spontaneously, and therefore that the recycling factor eIF2B may not be as critical to translation initiation (Shaikhin *et al* 1991). This is further supported by the fact that though gene homologs of all five subunits of the recycling factor eIF2B have been identified in plant genomes, the eIF2B protein complex has not been isolated in plants, and whether it is expressed and functioning in plants remains to be shown.

Second, although one eIF2 $\alpha$ -kinase (*gcn2*) gene is present and expressed in several plant species (Zhang *et al* 2008), it has not yet been shown to decrease the rate of translation in plants. The fact that plants do not possess *gcn4* in their genome, nor any other eIF2 $\alpha$ -kinase gene suggests that eIF2 $\alpha$  phosphorylation may not play a significant regulatory role in plants (Browning 2004). Since plants can synthesize all amino acids

from a *de novo* pathway, it is not logical that translational inhibition via amino acid starvation would be active in plants. In addition, four different phosphorylated isoforms of eIF2 $\beta$  have been found in plants, two of which are phosphorylated by Casein Kinase 2 (Dennis 2008). This suggests that it may be possible that the plant translational system may be regulated via eIF2 phosphorylation, but at another amino acid residue, and through a different mechanism.

Thus, it has not been shown conclusively that eIF2 $\alpha$  phosphorylation occurs, is required, or is sufficient to cause translational inhibition in plants. The evidence collected to date is not fully consistent and does not convincingly show that this mechanism of regulation is or is not functional and active in plant systems. In order to resolve these differences, further study needs to be conducted. This study contributes to the clarification of this problem by constructing a single bacterial expression vector containing coding sequences of all three subunits of eIF2 from *T. aestivum* (wheat germ), in order to test the expression of eIF2 $\alpha$ ,  $\beta$ , and  $\gamma$  as a functional complex in *E. coli*. Once this is completed, manipulation and detailed characterization of wheat eIF2 may be conducted.



## Experimental Methods

### Gene Design

Amino acid sequences for wheat eIF2 $\alpha$ , eIF2 $\beta$ , and eIF2 $\gamma$  were collected from NCBI (Appendix A). These amino acid sequences were reverse translated into DNA sequences (DNAworks<sup>®</sup> program at <http://helixweb.nih.gov/dnaworks>) to remove intronic sequences, adjust codon bias for expression in *E.coli*, and remove particular restriction sites via silent mutation. The codon usage tables for *E.coli* and *T.aestivum* (Appendix E) demonstrate the severity of the difference of codon bias of the two organisms; therefore, in order to optimize expression in *E.coli*, nearly all codons had to be adjusted.

Additionally, prefix and suffix DNA sequences were added to each gene that included a mirror of the restriction site complement found in the BioBrick vector that surround the insertion region. The prefix of each gene also contains a Shine-Dalgarno prokaryotic ribosome binding site. Finally, in each gene a specific “backdoor” restriction site was engineered via silent mutation (<http://moby.le.pasteur.fr/cgi-bin/portal.py?form=silent>). After this manipulation, the length of the coding sequences were 1093 base pairs for eIF2 $\alpha$ , 886 base pairs for eIF2 $\beta$ , and 1611 base pairs for eIF2 $\gamma$ . In order to ensure fidelity in the synthetic amplification process, eIF2 $\gamma$  was constructed in two parts:  $\gamma$ 1 and  $\gamma$ 2 (Figure 3, Appendix B).

The DNAworks<sup>®</sup> program generated a set of 5' to 3' overlapping 60-bp oligos spanning the entire length of the gene (both strands), allowing for gene synthesis via overlap extension PCR (Appendix C). Each set of oligos (Invitrogen) was combined in a mix so that each oligo (original concentration 100  $\mu$ M) was diluted to a final concentration of 1  $\mu$ M at a final volume of 200  $\mu$ L.

## **PCR**

Overlap Extension PCR: A PCR reaction was set up using 1  $\mu\text{L}$  of the oligo mix, 5  $\mu\text{L}$  10X KOD<sup>®</sup> reaction buffer, 5  $\mu\text{L}$  2 mM dNTPs, 3  $\mu\text{L}$  25 mM MgSO<sub>4</sub>, 1  $\mu\text{L}$  1 U/ $\mu\text{L}$  KOD<sup>®</sup> HotStart<sup>®</sup> Polymerase, and 35  $\mu\text{L}$  sterile dH<sub>2</sub>O. All PCR reagents were purchased from EMD Chemicals, Novagen<sup>®</sup>. These reactions were thermocycled under the following conditions: 95°C for 2 minutes, followed by 20 cycles (denaturation at 95°C for 20 seconds, primer annealing at 58°C for 10 seconds, and elongation at 70°C for 15 seconds), and finally 70°C for 2 minutes. In overlap extension PCR, each overlapping region functions as a primer for the adjacent oligos, so that PCR products of all different lengths and positions along the gene are created. Out of this mixture of PCR products, some full gene sequence product is also present.

Gene Amplification PCR: To amplify the full gene sequence, a second PCR reaction was set up with 1  $\mu\text{L}$  of the overlap extension PCR reaction, 5  $\mu\text{L}$  10X KOD<sup>®</sup> reaction buffer, 5  $\mu\text{L}$  2 mM dNTPs, 3  $\mu\text{L}$  25 mM MgSO<sub>4</sub>, 1  $\mu\text{L}$  20 mM 5' forward primer (Appendix D), 1  $\mu\text{L}$  20 mM 3' reverse primer (Appendix D), 1  $\mu\text{L}$  1U/ $\mu\text{L}$  KOD<sup>®</sup> HotStart Polymerase<sup>®</sup>, 33  $\mu\text{L}$  sterile dH<sub>2</sub>O. The thermal cycling conditions were as previously described, using a total of 30 cycles instead of 20.

## **RE Digestion**

Double digest reactions consisted of 2.5  $\mu\text{L}$  20,000U/ $\mu\text{L}$  of each RE (New England Biolabs, Inc.<sup>®</sup>), 5  $\mu\text{L}$  of the appropriate 10X NEBuffer<sup>™</sup>, ([http://www.neb.com/nebecomm/DoubleDigestCalculator .asp](http://www.neb.com/nebecomm/DoubleDigestCalculator.asp)), and 3-5  $\mu\text{g}$  of plasmid in a 50  $\mu\text{L}$  volume. Triple digests consisted of 2  $\mu\text{L}$  20,000 U/ $\mu\text{L}$  of each RE, 6  $\mu\text{L}$  of the appropriate 10X

NEBuffer™, and 3-5 µg of plasmid in a 60 µL volume. The reactions were incubated at 37°C overnight. Vector digests were phosphatase-treated by adding 1 µL 5000U/µL Antarctic Phosphatase (NEB.®), 6 µL 10X Antarctic Phosphatase Reaction Buffer, and sterile water to a final volume of 60 µL. After adding the phosphatase, incubation at 37°C was continued for 15 minutes, followed by 20 minutes at 65°C for restriction enzyme heat inactivation.

### **Gel Purification**

All PCR products or digested miniprep plasmid products were separated on a 1% agarose gel, excised under UV visualization, and purified using the GenElute™ Gel Extraction Kit according to the manufacturer's instructions (Sigma-Aldrich®).

### **Ligation**

For blunt-end ligations, 25 ng of Zero Blunt™ TOPO™ (Invitrogen) vector was combined with each gel-purified PCR product in a 100:1 molar ratio of PCR product insert to vector; to this was added 1 µL 400,000U/µL T4® DNA ligase (NEB®), 1 µL 10X T4® DNA ligase buffer, and sterile water to bring the final volume to 10 µL. Reactions were incubated for one hour at 16°C.

For ligations involving restriction-digested plasmids, 50 ng of the digested BioBrick or pET22bb vector was combined with each digested insert in a 3:1 molar ratio of insert to vector, and incubated at 16°C overnight.

## **Transformation**

For transformation into Top 10<sup>®</sup> cells (Invitrogen), 2  $\mu\text{L}$  of the ligation reaction was added to thawed Top 10<sup>®</sup> cells (100  $\mu\text{L}$ ). The cells were set on ice for 30 minutes, heat shocked in a water bath at 42°C for 90 seconds and returned to ice for two minutes. 100  $\mu\text{L}$  SOC broth was added and the cells were gently shaken at 37°C for one hour; the entire volume was plated on agar plates containing LB and kanamycin when using Zero Blunt<sup>™</sup>, or ampicillin when using BioBrick or pet22bb, and incubated at 37°C for 16-18 hours. For transformation into BL21<sup>®</sup> cells (Invitrogen), 1  $\mu\text{L}$  of miniprepped plasmid was added to the cells; 500  $\mu\text{L}$  of SOC broth was added and 50  $\mu\text{L}$  was plated.

## **Screening**

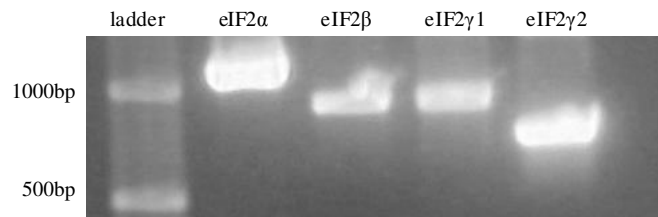
From each plate of transformed cells, 10 colonies were picked with a sterile pipette tip and immersed in a 25  $\mu\text{L}$  PCR reaction containing 2.5  $\mu\text{L}$  10X Taq DNA Polymerase Reaction Buffer, 0.5  $\mu\text{L}$  10 mM dNTPs, 0.15  $\mu\text{L}$  5U/ $\mu\text{L}$  Taq DNA Polymerase (EMD Chemicals, Novagen<sup>®</sup>), and 1.25  $\mu\text{L}$  of 20  $\mu\text{M}$  5' and 3' vector-specific primers (Zero Blunt-specific primers M13F and M13R, BioBrick-specific primers VF2 and VR, or pet22bb-specific primers T7pro and T7term (Appendix D)). When colonies containing large inserts were screened (over 2 kb), an oligo was selected from the middle region of the coding sequence and in the reverse orientation, diluted to 20  $\mu\text{M}$ , and used as a primer along with a vector-specific forward primer in order to decrease the length of the amplified region to about 500 bp and thereby ensure efficient amplification). After immersion in the PCR reaction, each picked colony was then immediately streaked onto replica LB-agar plates containing 30  $\mu\text{g}/\text{mL}$  of kanamycin

(when using Zero Blunt) or ampicillin (when using BioBrick or pet22bb), and the tip was then ejected into a 5 mL culture of LB containing 100 µg/mL kanamycin or ampicillin. The “colony PCR” reactions were thermocycled according to the following parameters: 94°C for 5 minutes, 35 cycles (denaturation at 94°C for 30 seconds, primer annealing at 55°C for 30 seconds, elongation at 72°C for 90 seconds), and 72°C for 5 minutes. The replica plates and cultures were incubated at 37°C for 16-18 hours. The results of the PCR were run on a 1% agarose gel. The cultures corresponding to clones displaying an inserted sequence of the correct size were miniprepmed via the GenElute™ Plasmid Miniprep Kit (Sigma-Aldrich®) and were sequenced with plasmid-specific primers for confirmation.

## Results

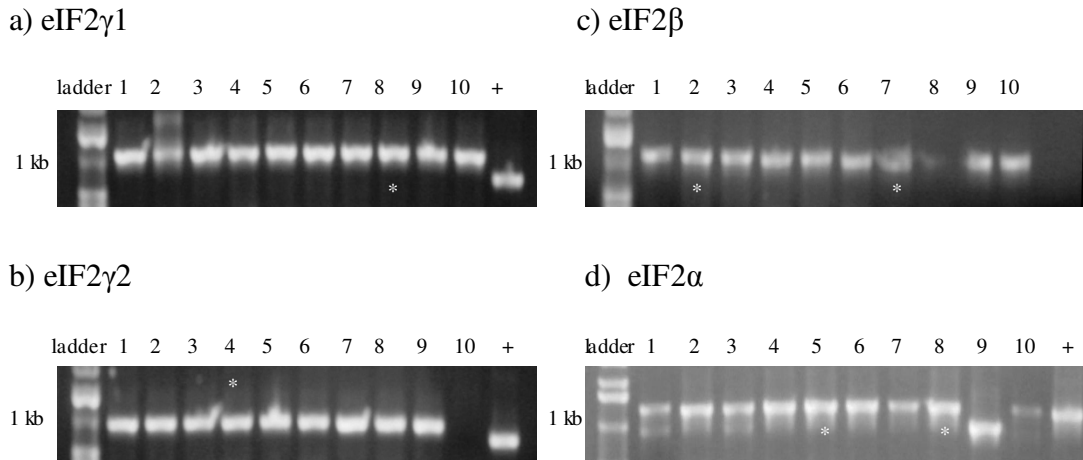
### Gene Synthesis

Complete coding sequences for eIF2 $\alpha$ , eIF2 $\beta$ , eIF2 $\gamma$ 1, and eIF2 $\gamma$ 2 were designed and prepared through overlap extension PCR. The products of the second PCR reaction that uses 5' and 3' end primers specific to each subunit (Appendix C) are shown below (Figure 3).



**Figure 3. Confirmation Gel of Synthesized Coding Sequences. eIF2 $\alpha$  1093 bp; eIF2 $\beta$  886 bp; eIF2 $\gamma$ 1 886 bp; eIF2 $\gamma$ 2 725 bp.**

Each PCR product was ligated into the Zero Blunt<sup>TM</sup> vector and transformed into *E. Coli* strain Top 10<sup>®</sup> competent cells. This strain of cells was chosen because of its susceptibility to the death gene *ccdB* which expresses in Zero Blunt<sup>TM</sup> vectors that have re-ligated without any insert. Because Zero Blunt<sup>TM</sup> confers kanamycin resistance, only transformed cells with inserts should be able to establish colonies on the LB-kanamycin plate. Colonies were screened (Figure 4), and true positives were miniprepped and sequenced (Appendix F). For eIF2 $\alpha$  and eIF2 $\beta$ , complete correct clones were not identified. Two separate clones were chosen, one with a correct 5' end and one with a correct 3' end, for both eIF2 $\alpha$  and eIF2 $\beta$ . For both eIF2 $\gamma$ 1 and eIF2 $\gamma$ 2, complete correct clones were identified (Appendix F).



**Figure 4. Colony PCR confirmation of insert-containing Zero Blunt plasmids using M13F and M13R primers. The + indicates a positive control in which a sequence-confirmed colony transformed with the Zero Blunt vector containing an insert was picked. Note there was no positive control run on image (c) because it is part of the same gel as image (d). Asterisks indicate colonies with correct sequencing results. For eIF2 $\beta$  and eIF2 $\alpha$ , separate colonies were chosen with correct 5' and 3' ends because a complete correct clone was not identified. In c), eIF2 $\beta$  -7 has a correct 5' end, and eIF2 $\beta$  -2 has a correct 3' end. In d), eIF2 $\alpha$  -8 has a correct 5' end, and eIF2 $\alpha$  -5 has a correct 3' end.**

## Cloning Strategy

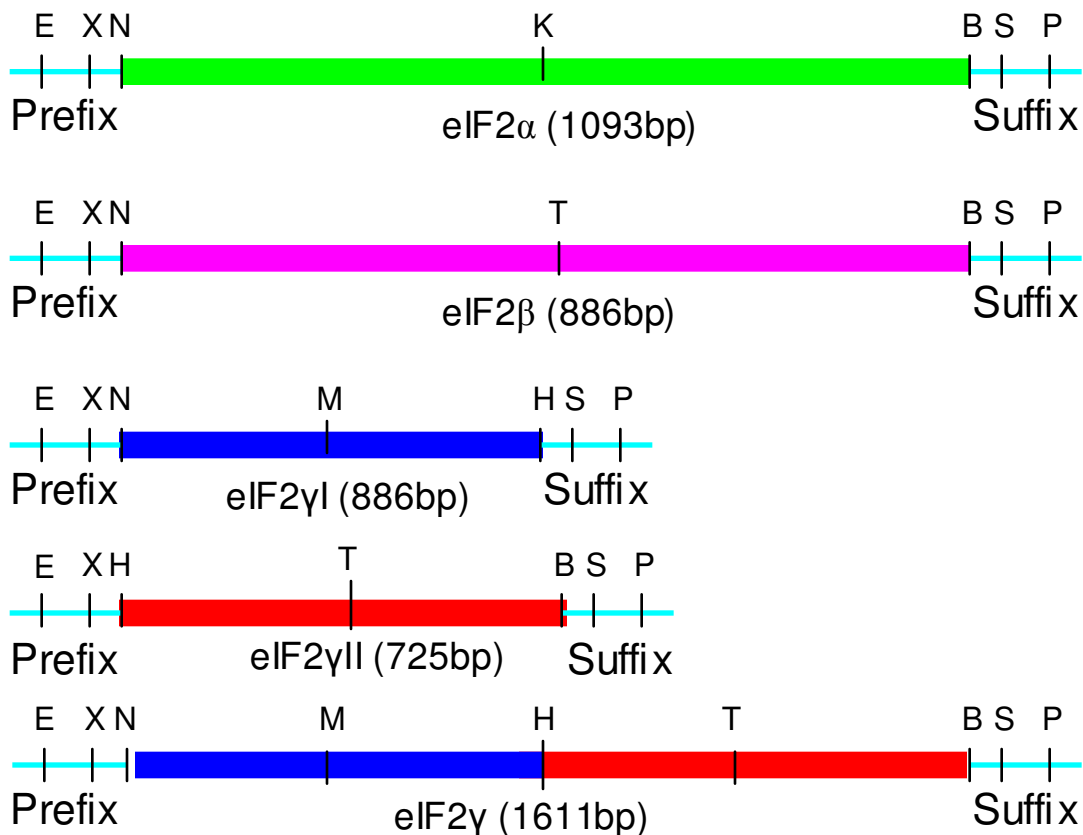
### Overview--

1. Move correct 3' end of eIF2 $\alpha$ , correct 5' end of eIF2 $\beta$ , and eIF2 $\gamma$ 1 from Zero Blunt vectors into BioBrick vectors.
2. Move correct 5' end of eIF2 $\alpha$  in Zero Blunt to BioBrick vector containing correct 3' end of eIF2 $\alpha$ .
3. Move correct 3' end of eIF2 $\beta$  in Zero Blunt to BioBrick vector containing correct 5' end of eIF2 $\beta$ .
4. Use the eIF2 $\gamma$ 1-BioBrick plasmid as a recipient vector for eIF2 $\gamma$ 2 insert.
5. Use the eIF2 $\alpha$ -BioBrick plasmid as a recipient vector for eIF2 $\gamma$  insert.
6. Use the eIF2 $\alpha\gamma$ -BioBrick plasmid as a recipient vector for eIF2 $\beta$  insert.
7. Move the eIF2 $\beta\alpha\gamma$  insert into pET22bb vector for protein expression studies.

**Figure 5. Overview of Cloning Strategy**

The three Zero Blunt plasmids that individually contained eIF2 $\alpha$ -5 (correct 3' end), eIF2 $\beta$ -7 (correct 5' end), and eIF2 $\gamma$ 1-8 were digested with EcoRI and PstI. The BioBrick vector was also digested with EcoRI and PstI; a BioBrick vector with an insert

was used so that a visible band would separate on the gel and confirm the vector was cut to completion. Each insert was ligated into the digested BioBrick vector and transformed. Colonies were screened by PCR and plasmids from positive colonies were isolated and sequenced. As shown in Figure 6, each subunit was engineered with a “backdoor” restriction cut site near the middle of the gene. This “backdoor” allows replacement of a portion of the DNA sequence if needed.

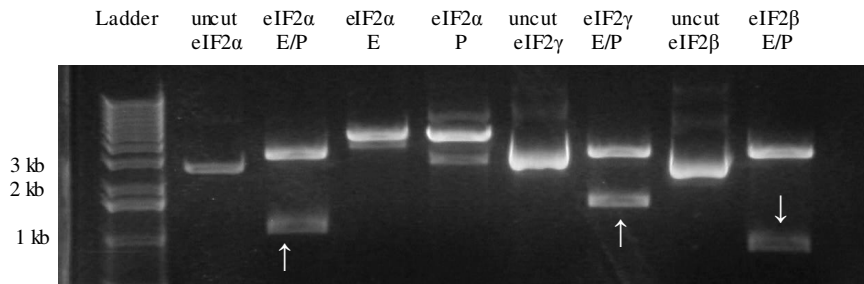


**Figure 6. Gene assembly diagrams showing restriction sites, as designed and synthesized via the DNAworks program, read from left to right as 5' to 3'. Figures are not to scale. Abbreviations: E-EcoR1; X-Xba1; N-Nde1; B-BamH1; S-Spe1; P-Pst1; K-Kpn1; T-Sal1; M-Mlu1; H-HindIII. Schematics are adapted from those provided by Dr. Grace Choy PhD**

To create full length correct-sequence constructs, the correct 3' end clone of eIF2 $\alpha$  now in BioBrick (eIF2 $\alpha$ -5) and the correct 5' end clone of eIF2 $\alpha$  still in Zero Blunt™ (eIF2 $\alpha$ -8) were each digested with KpnI and XbaI. The BioBrick vector band



containing the correct 3' end of eIF2 $\alpha$  (eIF2 $\alpha$ -5) was used as the recipient for the KpnI/XbaI fragment digested out of the Zero Blunt vector that contains the correct 5' end (eIF2 $\alpha$ -8). In the same manner, the correct 5' end clone of eIF2 $\beta$  now in BioBrick (eIF2 $\beta$ -7) and the correct 3' end clone of eIF2 $\beta$  still in Zero Blunt<sup>TM</sup> (eIF2 $\beta$ -2) were each digested with BamHI and Sall, and the defective 3' end of eIF2 $\beta$ -7 was replaced by the correct 3' end of eIF2 $\beta$ -2. To join eIF2 $\gamma$ 1 and  $\gamma$ 2, as shown in Figure 3, eIF2 $\gamma$ 1 in BioBrick and eIF2 $\gamma$ 2 in Zero Blunt<sup>TM</sup> were each digested with HindIII and SpeI. After ligating, the resulting BioBrick plasmids now containing complete eIF2 $\alpha$ , eIF2 $\beta$ , and eIF2 $\gamma$  coding regions were digested with restriction enzymes to confirm the correct size insert is obtained (Figure 7) and were sequenced (Appendix F).



**Figure 7. RE Digests with EcoRI and PstI on single-subunit plasmids in the BioBrick vector to confirm presence and correct size of insert. Expected sizes: eIF2 $\alpha$  1093 bp, eIF2 $\gamma$  1611 bp, and eIF2 $\beta$  886 bp. Abbreviations: E-EcoRI; P-PstI.**

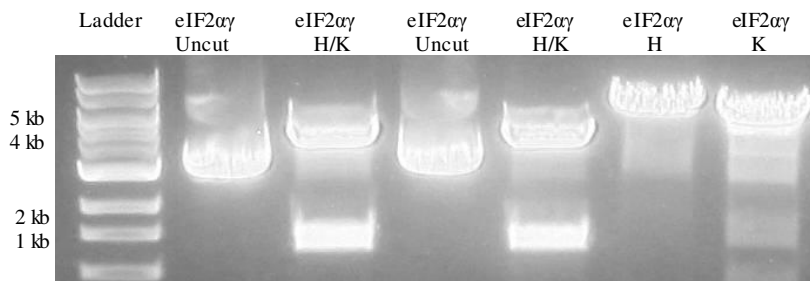
The BioBrick vector was developed at MIT (Knight 2003) and has a unique complement of restriction sites that allows the assembly of several genes in one vector via repeated restriction enzyme digests, ligations, and transformations. To place all the subunit coding regions into an operon for expression, the BioBrick vector containing eIF2 $\alpha$  was used as the recipient for the eIF2 $\gamma$  coding region. Briefly, the eIF2 $\alpha$ -BioBrick

plasmid was digested with SpeI and PstI, and the eIF2 $\gamma$ -BioBrick plasmid was digested with XbaI and PstI. The BioBrick vector is designed with XbaI and SpeI restriction sites on either side of the insertion region. These enzymes produce complementary overhangs that will ligate when mixed; however, the sites have different internal base pairs (those immediately adjacent to the overhang) and will no longer be cleaved by either enzyme (Figure 8). The joined parts are now a new “BioBrick” and may be joined to another DNA fragment using the same restriction enzymes.

|               |               |                        |
|---------------|---------------|------------------------|
| XbaI: T/CTAGA | SpeI: A/CTAGT | After Ligation: TCTAGT |
| AGATC/T       | TGATC/A       | AGATCA                 |

**Figure 8.** The figure shows the cut sites of XbaI and SpeI, and indicates the sequence resulting from a ligation reaction. All sequences are read 5' to 3' on top, 3' to 5' on bottom.

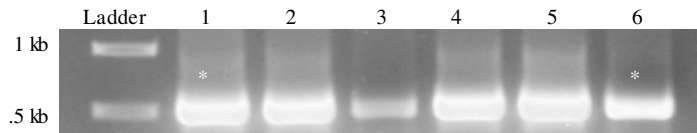
The eIF2 $\gamma$  insert was ligated into the eIF2 $\alpha$ -BioBrick plasmid and correct clones were identified via screening and confirmed via restriction enzyme digestion (Figure 9).



**Figure 9.** RE Digests with HindIII (eIF2 $\gamma$  backdoor enzyme) and KpnI (eIF2 $\alpha$  backdoor enzyme) on eIF2 $\alpha\gamma$  in the BioBrick vector to confirm presence and correct size of insert. Abbreviations: H – HindIII; K – KpnI.

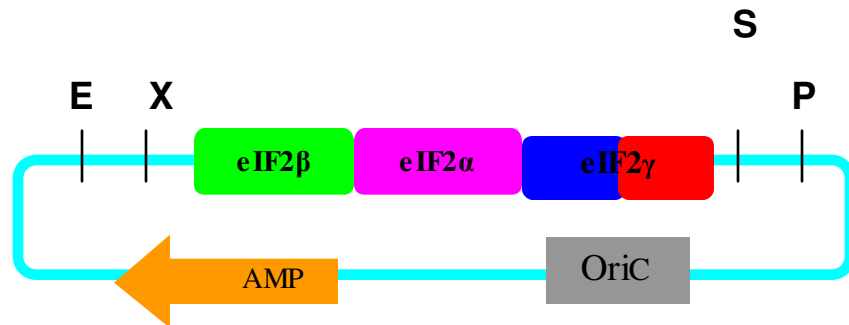
In the same way, the eIF2 $\alpha\gamma$ -BioBrick plasmid was used as the recipient for the eIF2 $\beta$  insert. The eIF2 $\alpha\gamma$ -BioBrick was digested with EcoRI and XbaI, and the eIF2 $\beta$ -

BioBrick plasmid was digested with EcoRI and SpeI. The eIF2 $\beta$  insert was ligated into the eIF2 $\alpha\gamma$ -BioBrick plasmid and resulting colonies were screened (Figure 10) and sequenced (Appendix F).



**Figure 10. Colony PCR screening on eIF2 $\beta\alpha\gamma$  in BioBrick using VF2 and an oligo from the middle of the eIF2 $\beta$  coding region. Asterisks indicate colonies with correct sequencing results used to continue cloning.**

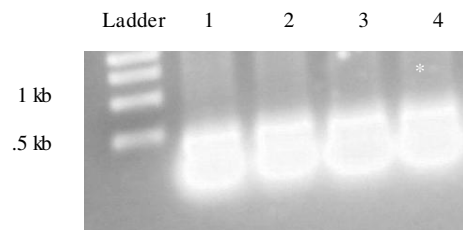
A schematic of the completed eIF2 operon in the BioBrick vector is shown below (Figure 11).



**Figure 11. Completed eIF2 $\beta\alpha\gamma$  operon in BioBrick. Each subunit contains its own Shine-Dalgarno ribosome binding site at its 5' end.**

To m ector, the eIF2 $\beta\alpha\gamma$ -BioBrick plasmid and the pET22bb vector were both digested with EcoRI, PstI, and XhoI. XhoI was added to cut the BioBrick vector backbone into three parts of around 1 kb, so that it would be easily separated from the eIF2 $\beta\alpha\gamma$  insert (3.6 kb). The eIF2 $\beta\alpha\gamma$  insert was ligated into the pET22bb vector and transformed into Top 10 cells.

The resulting colonies were screened (Figure 12) and sequenced (Appendix F). The eIF2 $\beta$  $\alpha$  $\gamma$ -pET22bb plasmid was transformed into BL21 for expression.



**Figure 12. Colony PCR screening on eIF2 $\beta$  $\alpha$  $\gamma$  in pET22bb using T7pro and an oligo from the middle of the eIF2 $\beta$  coding region. Asterisk indicates colony with correct sequencing results.**

## Discussion

Through the research experiments detailed here, plasmids have been constructed carrying coding sequences for wheat germ eIF2 $\alpha$ , eIF2 $\beta$ , and eIF2 $\gamma$  individually in the BioBrick vector. Additionally, all three coding sequences have been assembled adjacent to one another in an operon in the BioBrick vector and transferred to the pET22bb vector for expression of the operon in *E. coli*. A discussion of the difficulties, summary of the strategies, and introduction to the initial stages of expression analysis follows in this section.

Difficulties were encountered at several steps throughout the gene-assembly process. Several of the double digest reactions called for a specific combination of restriction enzymes that did not share an optimal buffer. In these cases, the amount ratios of the two enzymes were adjusted from 1:1 to 3:2, using more of the enzyme that had lower performance in the chosen buffer. Another difficulty was obtaining sufficient recovery of the pET22bb plasmid from bacterial cultures because of its low copy number. In order to overcome this, the standard manufacturer's protocol was adjusted so that elution volume was reduced to nearly half the recommended volume, and the incubation time (with the elution solution on the column) was increased to five minutes. A final difficulty of note was encountered in the colony PCR protocol. In order for the reaction to occur, the cells need to be broken open. It was found that this can be accomplished manually by rubbing the pipette tip against the wall of the PCR tube.

Though technical problems in individual protocols were faced, the overall strategies and procedures of the cloning strategy were successful. By designing DNA sequences from reverse-translated wheat eIF2 amino acid sequences and adjusting codons

to match usage in *E. coli*, the resulting DNA sequences encode identical proteins to those found in wild type wheat germ, and at the same time have optimal expression in *E. coli*. The engineering of “backdoor” restriction sites into the middle of each DNA sequence by silent mutation enabled the piecing together of partial correct clones, removing the need to screen and sequence large numbers of transformed colonies in search of a correct clone. This ability to pair 5’ and 3’ correct clones proved useful in the case of eIF2 $\beta$  and eIF2 $\alpha$ , though it turned out to be unnecessary in the case of eIF2 $\gamma$ 1 and eIF2 $\gamma$ 2. The complement of restriction sites in the BioBrick plasmid itself allowed for the assembly of the wheat eIF2 $\beta$ ,  $\alpha$ , and  $\gamma$  coding sequences into a single operon by sequential digestion and ligation reactions.

Initial studies of protein expression have been conducted on the eIF2 $\beta\alpha\gamma$  operon in the pET22bb vector in BL21 cells, including analysis of SDS-PAGE gel by Coomassie Stain and Western Blot. These results cannot yet be reported because of their inconclusive nature. The background of *E. coli* proteins seen on the Coomassie stain does not allow for distinctive bands to be seen, and the Western Blot is also not meaningful due to low-activity antibodies. Therefore, further testing needs to be completed before experimental data can be interpreted or reported. A partial protein purification will be done prior to repeating the SDS-PAGE gel analysis in order to add confidence to and simplify the interpretation of Coomassie Stain and Western Blot results. Expression of all three subunits of the eIF2 operon is expected, and in that light a brief discussion of future studies is presented in the following section.

## Future Directions

It is the intent to express the fully functional eIF2 protein complex in *E. coli*. If it can be confirmed that all three subunits are expressed and are soluble, there are several further directions in which to take this research.

The purification of the expressed eIF2 protein complex from *E. coli* would be attempted. If this is successful, the function of the purified eIF2 complex can be assayed. This would include measuring the ability of eIF2 to support polypeptide synthesis in eIF2-depleted wheat germ extracts (Benkowski *et al.* 1995) or to bind [<sup>35</sup>S]Met-tRNA<sub>i</sub><sup>Met</sup> or [3H]GTP as a measure of ternary complex formation.

After the activity of the expressed eIF2 is established, an exogenous eIF2 $\alpha$ -kinase, perhaps from a yeast or mammalian system, may be used to determine whether or not eIF2 $\alpha$ -phosphorylation causes translation in a wheat germ extract to be significantly hindered. In addition, we can explore other possible mechanisms of translational control via eIF2 in plant systems by introducing mutations via site-directed mutagenesis that may shed light on eIF2 function.

In conclusion, the completion of the current research project detailed in this thesis will allow for the study of a number of aspects of the mechanism and regulation of eIF2 function in wheat. This will add to the scientific body of knowledge concerning translational regulation in plants, which has significant implications for agricultural biotechnology as well as for comparative analyses of the mechanisms of protein synthesis across the kingdoms of life.

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## Appendix A – Wheat germ eIF2 Amino Acid Sequences

### eIF2 $\alpha$ :

MANLECRMYEPRFPEVDAAVMIQVKHIADMGAYVSLLEYNNVEGMILFSELSRRRIRSISSLIKVGRQEPAILVR  
VDRDKGYIDLSKRRVSEEEARSCEDKYNKSKLVHSIMRHVAETLEIDLEPIYQRIGWPLYRKYGHAFKFLIVA  
DPDAILDVLT YEERETGPDGQEVTKVVPVAVTPEIKETLVQNI RRRMTPQPLKIRADVEMKCFQFDGVLHIKQAMR  
KAEAAGNTNCPVKIKLVAPPLYVLTQTLDKQGISVLTDAVKACTAEIEKHKGKLVVKEAPRAVSEREDKLLNA  
QLDTLVEQNAEVAGDDDSEDEEDTGMGDI DLTNSGVHAD

### eIF2 $\beta$ :

MADEEQMERKEEATEIAPFDPTKKKKKKVVIQDPADEVDKLAEKTEGLSVTESGEASFVGLKKKKKLVLDPS  
LVEAGDGEDTLDQVGEDEQGEIIVLGGATQYPWEGTDRDYKYDELLGRVFNILRENNPDLAGDRRRTVMRPPQV  
LREGTKKTVFVNFMDLCKTMHRQPEHVMMFLAEMGTSGSLDGQQLVIKGRFAPKNFEAILRRYINEYVICNGC  
KSPDTILSKENRLLFFLRCEQCGSSRSVAPIKAGFVAQVGRRKAGT

### eIF2 $\gamma$ :

MARRGLMEQDLTKLDVTKLHPLSPEVISRQATINIGTIGHVAHGKSTVVKAISGVQTVRFKNELERNITIKLGYA  
NAKIYKCEDDRCPREMCYKAYGSGKEDTPACDVPGFENTRMKLLRHVSFVDCPGHDI LMATMLNGAAIMDGALLL  
IAANESCPQPQTSEHLAAVEIMRLQHLII LQNKIDLIOESAAMNQHEAIQKFIQGTIAEGAPVVPI SAQLKYNID  
VICEYIIKKIPIPENFTSPPNMIVIRSFVNKPGSE  
FTSPPNMIVIRSFVNKPGSEVDEIRGGVAGGSI LRGVLRVNQNI EVRPGIVMKDESGNIKCTPIYSRIVSLYAE  
QNELQFAVPGGLIGVGT TMDPTLTRADRLVGQVLGEIGSLPDVFVELEINFFLLRRL LGVRTKGTEKAGKVKSLT  
KGEIIMLNIGSMS TGARVAVKNDLAKLQLTAPVCTSKGEKVALSRRVEKHWRLIGWGQIQAGATLEVPPCPL

## Appendix B – Wheat germ eIF2 DNA sequences.

These were modified using DNAworks to adjust codon bias for expression in E.coli, and remove particular restriction sites via silent mutation. Additionally a prefix and suffix (in red) were added to each gene that included a mirror of BioBrick restriction site complement and an IRES element in the prefix. Note that here eIF2 $\gamma$  is artificially broken up into two parts for the sake of efficient and accurate cloning. It is pieced back together at a later stage.

### eIF2 $\alpha$ :

```
GAATTCGCGGCCGCTTCTAGAGATTAAAGAGGAGAAAACCATATGGCGAACCTGGAATGCCGTATGTATGAACCGCGTTTTCCGAAGTGGATGCTGCGGTGATGATTCAGGTGAAACATATTGCGGATATGGGCGCGTATGTGAGCCTGCTGGAATATAACAATGTGGAAGGCATGATTCTGTTTTCCGAGCTGAGCCGTCGCCGCATTTCGGAGCATAAGCAGCCTGATAAAAAGTGGGTGCTCAAGAACCGGCATAGTTCTTCGTGTGGATCGTGATAAAAGGCTATATTGACTTGTC CAAGCGCCGTGTTTTCCGAAGAAGAAGCGGTAGCTGCGAGGATAAATACAATAAGAGCAAGCTGGTCCATAGCAT TATGCGTCACGTGCTGAGACCCTGGAGATTGATCTGGAACCGATCTATCAGCGGATTGGCTGGCCGCTGTATCG TAAATACGGCCACGCATTTGAAGCGTTCAGCTGATGTGTGCGGACCCTGACGCCATATAGACGTGCTGACCTA TGAAAGAGCGTGAACCGGGCCAGATGGCCAAGAAGTGACCAAAGTGGTACCAGCAGTCAACCCGAAATAAAGGA AACCCCTGGTGCAAGAACATTTCGTGTCGGATGACACCACAACCGTGAAAATTCGTGCGGACGTGGAAATGAAATG CTTTTAGTTTGTAGGCGTGTGCATATCAAACAGGCGATGCGTAAGGCGGAGGCTGCGGGCAATACCAATTGTCC GGTAAAGATTAAAGTTAGTGGCGCCTCCACTGTATGTACTGACCACCCAGACACTGGACAAAGATCAGGGCATTAG CGTTTTAACCAGTGGCGTCAAAGCGTGCACGCGGAAATCGAAAAACATAAGGGCAAACCTGGTGGTGAAGAAAGC CCCACGCGCGGTGTGCAAGCGTGAAGATAAAGTGTGAATGCCAACGATGATACTCTGGTAGAACAAAATGCGGA AGTTCGAGGCGACGATGATAGCGAGGACGAAGAGGATACCGGCATGGGCGACATCGATCTGACCAATAGCGGCGT GCATGCGGATTAATAAGGATCCTACTAGTAGCGGCCGCTGCAG
```

### eIF2 $\beta$ :

```
GAATTCGCGGCCGCTTCTAGAGATTAAAGAGGAGAAAACCATATGGCGGACGAAGAACAGATGGAACGTAAAGGA AGAAGCGACCGAGATTGCGCCGTTTTGACCCGACTAAGAAGAAGAAGAAGAAGAAAGTGGTGATTTCAGGACCCAGC GGATGAAGTGGATAAATTGGCCGAAAAGACCGAAGGCCTGAGCGTGACCAGAAAGCGGCGAGGCGAGCCTTTGTGGG CTTGAAGAAAAAGAAAAAGAAGTTAGTAGAAGTGGACCCAGCCTTGTGCAAGCCGGTGATGGCGAAGATACCCCT GGACGATCAGGTGGCGGAGGATGAACAAGGAGAGGGCATTGTGCTGGGCGGTGCCACCCAGTATCCGTGGGAAGG AACCGATCGTGATTACAAATATGATGAGCTGTGGGCGGTGTGTTCAATATTCGCGCGAGAATAATCCAGACCT GGCGGGAGACCGTCGACGTACCGTGTGCGTCCACCGCAGGTCTTGCCTGAAGGCACCAAAAAAACCGTGTGTTGT GAATTTTCATGGATCTGTGCAAAACCATGCACTGTCAGCCGGAACATGTGATGATGTTCTGCTTGCAGGAAATGGG CACCAGCGGCGAGCTGGATGGGCGAGCAGCGTCTGGTCAATTAAGGCCGCTTCGCTCCCAAGAATTTTGAAGCGAT CCTGCGGCGTTACATCAATGAATATGTGATTTGCAATGGCTGCAAGAGCCCGGATACCATCTGAGCAAAAGAGAA TCGTCTGTTTTTCCGCGTTGCGAACAGTGCAGCAGTAGCCGTAGCGTGGCGCCGATTAAGCCGGGTTCTGTG C CAGGTAGGCCGTCGTAAGGCGGGCACCTAATAAGGATCCTACTAGTAGCGGCCGCTGCAG
```

### eIF2 $\gamma$ 1:

```
GAATTCGCGGCCGCTTCTAGAGATTAAAGAGGAGAAAACCATATGGCGCGTCGGGGACTGATGGAACAGGATCT TACCAAATTTGGATGTCACAAAACCTGCATCCGCTGAGCCCGGAAGTGATTAGCCGTCAGGCCACGATTAACATAGG GACCATTTGGCCATGTGGCGCATGGCAAAGCACTGTGGTAAAGGCTATTAGCGGGGTGCAGACCGTGCCTTTCAA GAATGAATTTGGAGCGTAACATAACCATTAACCTGGGCTATGCGAACGCGAAAACTACAAATGCGAAGATGATCG TTGCCCGCGTCCCATGTGCTACAAAGCGTATGGCAGCGGTAAAGAGGATACCCCTGCGTGCATGTCCCGGGTTT TGAGAATACGCGTATGAAACTGTTGCGCCATGTTAGCTTTGTGGATTGCCCTGGTCAATGATATTTTGTGAGGCGAC CATGCTGAACGGAGCCGCTATTATGGATGGCGCCCTGTTGCTGATTGCGGCAAAATGAATCTTGCCTCAGCCGCA GACCAGCGAACATCTGGCTGCGGTGGAATTTATGCGTCTTCAGCACCTGATAATCTTCAGAATAAGATTGATCT GATTTCAGGAATCCGAGCGATGAATCAACATGAGGCGATTCAAAGTTCATTCAAGGCACCATAGCGGAGGGCGC TCCAGTGGTTCCGATTAGCGCGCAGTTGAAGTATAACATCGACGTAATTTGCGAATACATCATAAAGAAGATTCC GATCCCTGAACGTAACCTTACCAGCCACCGAATATGATGTTATTCGAAGCTTCGATGTGAATAAACCAGGCGAG TGAAGTGGATGAAATTCGTGGCGGCGTGGCTGGGGTTTCCTACTAGTAGCGGCCGCTGCAG
```

### eIF2 $\gamma$ 2:

GAATTCGCGGCCGCTTCTAGAGCTTTACCAGCCACCGAATATGATTGTTATTCGAAGCTTCGATGTGAATAAAC  
CGGGCAGTGAAGTGGATGAAATTCGTGGCGGCGTGGCTGGGGGTTCATTTACGTGGCGTGTTCGGGGTGAACC  
AGAACATTGAAGTGCCTCCGGGCATTGTGATGAAAGACGAATCGGGGAACATAAAGTGCACGCCCATTTATAGCC  
GTATTGTGAGCTTATATGCGGAACAAAATGAGCTGCAATTCGCGGTGCC TGGGGCC TGATTGGGGTGGGCACGA  
CTATGGACCCGACCCTGACTCGTGCGGATCGTCTGGTGGGCCAAGTGCTGGGAGAAATTGGTAGCCTGCCGGATG  
TCTTTGTTGAATTAGAGATTAAC TTTTTT CAGCTGCGTCTGCTGGGTGTTCGGACCAAAGGGACCGAGAAAG  
CGGGGAAAGTGAGCAAATTGACCAAGGGGAAAT TCTGATGTTAAACATCGGAAGCATGAGCACCGGAGCGCGTG  
TGGTTGCGGTCAAAAATGACCTGGCGAAGTTACAGTTAACAGCCCCAGTATG TACTTCGAAAGGCGAGAAGGTAG  
CGCTGTCCCCTCGTGTGGAAAAGCATTGGCGTCTGATCGGTTGGGGCCAGATTC AAGCTGGAGCTACCC TGGAAAG  
TGCCGCCGTGCCGCTGTAATAAGGATCCTACTAGTAGCGGCCGCTGCAG

## Appendix C- DNA works-generated Overlapping Oligos

Note again that here eIF2 $\gamma$  is artificially broken up into two parts for the sake of efficient and accurate cloning. It is pieced back together at a later stage. In each gene a “backdoor” restriction site was engineered via silent mutation (bolded, with silent mutation in red and recognizing enzyme noted to the side).

### eIF2 $\alpha$ :

```

1 ATGGCGAACC TGGAA T GCCGTATGTATGAACCGCGT TTTCCCG 43
2 TCCGCAATATGTTTCACTGAATCATCACCGCAGCATCCACTTCGGGAAAACGCGGTTC 60
3 TCAGGTGAAACATAT T GCGGATATGGGCGCGTATGTGAGCCTGCTGGAATATAACAATGT 60
4 GGCGACGGCTCAGCTCGGAAAACAGAA TCATG CCTTCCACAT TGTATATCCAGCAGGC 60
5 AGCTGAGCCGTCGCCGCATTCGGAGCATAAGCAGCCTGATAAAAAGTGGGT CGTCAAGAAC 60
6 TAGCCTTTATACGATCCACACGAAGA ACTATCGCCGGT TCTTGACGACCCACTTTTATC 60
7 TGTGGATCGTATAAAGGCTATATTGACTTGTCCAAGCGCCGTGTTTCCGAAGAAGAAGC 60
8 GACCAGCTTGCTCTTATGTATTTATCCTCGCAGCTACGCGCTTCTTCTTCGGAAACACG 60
9 TAAATACAATAAGAGCAAGCTGGTCCATAGCATTATGCGTCACGTTGCTGAGACCC T GGA 60
10 AGCGGCCAGCCAA TCCGCTGATAGATCGGTTCCAGATCAATCTCCAGGGTCTCAGCAACG 60
11 GGATTGGCTGGCCGCTGATTCGTAATAACGGCCACGCAT TGAAGCGTTCAAGCTGATTG 60
12 CATAGGT CAGCACGCTAATATGGCGTCAGGGTCCGCAACAATCAGCTTGAACGCTTCAA 60
13 CATATTAGACGTGCTGACCTATGAAGAGCGTGAACC GGGCCAGATGGCCAAAGAAGTGAC 60
14 GGTTTCTTTATTTCCGGGGTGACTGCTGGTACCACTTTGGTCACTTCTTGGCCATCTGG 60 kpnI
15 ACCCCGGAATAAAGGAACCCTGGTGCAGAACAT TCGTCGTGGATGACACCACAACCG 60
16 AAAC TGAAGCATTTCATTCCACGTCCGCACGAATTTTCAGCGGTTGTGGTGCATCCG 60
17 GTGGAATGAAATGCTTTTAGT TTAGTGGCGTGTGCATATCAAACAGGC GATGCGTAAAG 60
18 TTAATCTTTACCGACAATTGGTATGCCC GCAGCCTCCGCCTTACGCATCGCCTGTTTG 60
19 ATACCAATTGTCGGTAAAGATTAAGT TAGTGGCGCCTCCACTGTATGACTGACCACCC 60
20 CGGTTAAAACGCTAATGCCC TGATCTTTGTCCAGTGTCTGGGTGGT CAGTACATACAGTG 60
21 GGGCATTAGCGTTTAAACC GATGCGGTCAAAGCTGCACTGCGGAAATCGAAAACATAA 60
22 CACCGCGCGTGGGGCTTCTTTCAACCACAGTTTGCCCTTATGTTTTTCGATTTCCGCAGT 60
23 CCCACGCGCGGTGTCAGAACGTGAAGATAAAC TGCTGAATGCCAACTGGATACTCTGG 60
24 CTCGCTATCATCGTCGCTGCGACTTCCGCATTTTGT TCTACCAGAGTATCCAGTTGGGC 60
25 GGCGACGATGATAGCGAGGACGAAGAGGATACCGGCATGGGC GACATCGATCTGACCAAT 60
26 ATCCGCATGCACGCCGCTATTTGGTCAGATCGATGTCGC 38

```

### eIF2 $\beta$ :

```

1 ATGGCGGACGAAGAACAGATGGAACGTAAGGAAGAAGCGACCAGATTCGCGCCTTTGA 59
2 TCCTGAA TCACTCTTCTTCTTCTTCTTCTTTAGTCGGGTCAAACGGCGCAATCTCG 60
3 AGAAGAAAGTGGT GATTCAGGACCCAGCGGATGAAGTGGATAAAT TGGCCGAAAAGACCG 60
4 CAAAGCTCGCCTCGCCGCTTTCGGT CAGCTCAGGCC TTCGGTCTTTTCGGCCAATTTAT 60
5 GGCAGGGCAGCTTTGTGGGCTTGAAGAAAAGAAAAGAAGTTAGTAGAACTGGACCCC 60
6 TCCAGGTATCTTCGCCATCACC GCTTCGCACAGGCTGGGGTCCAGTTCTACTAATTC 60
7 ATGGCGAAGATACCTTGGACGATCAGGTGGGC GAGGATGAACAAGGAGAGGGCATGTGTC 60
8 ACGATCGGTTCC TCCACGGATAC TGGGTGGCACCGCCAGCACAA TGCCCTCTCCTTG 60
9 TGGGAAGGAACCGATCGTGATTA CAAATATGATGAGCTGT TGGGCCGTGTGTCAATAT 60
10 GTCGACGGTCTCCCGCCAGGCTG GATTATCTCGCGCAGAATATGAACACAGGCCCA 60 SalI
11 GCGGGAGACCGTCGACTACCGTGATGCGTCCACCGCAGGCTTGCCTGAAGGCACCAAAA 60 SalI
12 CATGGTTTTGCACAGATCCATGAAATTCACAAAACAGGTTTTTTGGTGCCTTCACGCAA 60
13 ATGGATCTGTGCAAAACCATGCATCGTCAGCCGGAACATGTGATGATGTTCTGCTTGCG 60
14 ACGTGTGTC CCA TCCAGGTGCCGCTGGTGCCATTTCCGCAAGCAGAAACATCATCAC 60
15 GGATGGGCAGCAGCGTCTGGTCA TTAAGCCGCTTCGCTCCCAAGAATTTTGAAGCGAT 60
16 CATTGCCAAATCACATATTCATTGATGTAACGCCG CAGGATCGCTTCAAATTC TTGGGAG 60
17 CATCAATGAAATATGTGATTTGCAATGGCTGCAAGACCCGGATACCATTC TGAGCAAAGA 60
18 TGCCGCACTGTTCCAAACG CAGGAAAACAGACGAT TCTCTTGTCTCAGAATGGTATCCG 60
19 TGCGAACAGTGC GGCAGTAGCCGTAGCGTGGCGCCGATTAAGCCGGGTTCGTTGCCCG 60
20 GGTGCCCGCCTTACGACGGCCTACCTGGGCAACGAACCCG 40

```

### eIF2 $\gamma$ 1:

```

1 ATGGCGCGTCGGGGACTGATGGAACAGGATCTTACCAAATGGATGTCACAAAAC T GCA 59
2 ATCTGTGGCCTGACGGCTAATCACTTCCGGGCTCAGCGGATGCAGTTTTGTGACATCCAAT 60
3 GCCGTCAGGCCACGATTAAACATAGGGACCATTGCCATGTGGCGCATGGCAAAGCACTG 60

```

4 TCTTCAAACGCACGGTCTGCACCCCGCTAATAGCCTTAACCACAGTGCTTTTGCCATGCG 60  
 5 CAGACCGTGCCTTCAAGAATGAATGGAGCGTAACATAACCATTAACCTGGGCTATGCG 60  
 6 GCGGGCAACGATCATCTTCGCATTTGTAGATTTTCGCGTTTCGCATAGCCAGTTAATGG 60  
 7 AAGATGATCGTTGCCCGCGTCCCATGTGCTACAAAAGCGTATGGCAGCGGTAAAGAGGATA 60  
 8 AT**ACGCGT**ATTTCTAAAACCCGGGACATCGCACGCAGGGGTATCCTCTTTACCGCTGCCA 60 **MluI**  
 9 CGGGTTTTGAGAA**ACGCGT**ATGAAACGTGTGCGCCATGTTAGCTTTGTGGATTGCCCTG 60 **MluI**  
 10 AGCGGCTCCGTTCAGCATGGTCGCCATCAAAATATCATGACCAGGGCAATCCACAAAGCT 60  
 11 GCTGAACGGAGCCGCTATTATGGATGGCGCCCTGTTGCTGATTGCGGCAAAATGAATCTTG 60  
 12 TCCACCGCAGCCAGATGTTTCGCTGGTCTGCGGCTGAGGGCAAGATTCATTTGCCGCAATC 60  
 13 CATCTGGCTGCGGTGGAAATTATGCGTCTTCAGCACCTGATAATTCTTCAGAAATAAGATT 60  
 14 TTGATTCATCGCTGCGGATTCCTGAATCAGATCAATCTTATTCTGAAGAAATATCAGGTG 60  
 15 ATCCGCAGCGATGAATCAACATGAGGCGATTCAAAAAGTTCATTCAAGGCACCATAGCGGA 60  
 16 TTATACTTCAACTGCGCGCTAATCGGAACCACTGGAGCGCCCTCCGCTATGGTGCCTTGA 60  
 17 TAGCGCGCAGTTGAAGTATAACATCGACGTAATTTGCGAATACATCATAAAGAAGATTCC 60  
 18 TATTCGGTGGCTGGTAAAGTTACGTTTCAGGATCGGAATCTTCTTTATGATGATTTGCG 60  
 19 CTTTACCAGCCACCAGAAATATGATTGTTATTCG**AAGCTT**CGATGTGAATAAACCGGGCAG 60 **HindIII**  
 20 GGAACCCCGACGCACGCCACGAATTTCACTTCACTGCCCAGTTTATTCACATC 60

eIF2 $\gamma$ 2:

19 CTTTACCAGCCACCAGAAATATGATTGTTATTCG**AAGCTT**CGATGTGAATAAACCGGGCAG 60 **HindIII**  
 20 GGAACCCCGACGCACGCCACGAATTTCACTTCACTGCCCAGTTTATTCACATC 60  
 21 CGTGGCTGGGGTTCCATTTTACGTGGCGTGTGCGGGTGAACCAGAACATTGAAGTGCG 60  
 22 TTTATGTTCCCGATTCGTCTTTTATCACAATGCCCGACGCACTTCAATGTTCTGGTTC 60  
 23 AGACGAATCGGGGAACATAAAGTGCACGCCATTTATAGCCGATTTGTGAGCTTATATGC 60  
 24 GGCCCCCAGGACCCGCAATTGCAGCTCATTTTGTTCGCATATAAGCTCACAATACGGC 60  
 25 GGTGCTGGGGCCTGATTGGGGTGGGCACGACTATGGACCCGACCTGACTCGTGCGGA 60  
 26 CCGGCAGGCTACCAATTTCTCCAGCACTTGGCCCACAGACGATCCGCACGAGTCAGGG 60  
 27 AAATTGGTAGCTGCCGGATGCTTTTGTGAAATAGAGATTAACCTTTTT**CAGCTG**CGTC 60 **SalI**  
 28 GCTTTCTCGTCCCTTTGGTCCGAACACCCAGCAGACGACG**CAGCTG**AAAAAAGTTAATC 60 **SalI**  
 29 CCAAAGGGACCGAGAAAAGCGGGGAAAGTGAGCAAAATGACCAAGGGGGAAATTTCTGATGT 60  
 30 CAACCACACGCGCTCCGGTGTCTATGCTTCCGATGTTTAAACATCAGAATTTCCCCCTTGG 60  
 31 GGAGCGCGTGTGGTTGCGGTCAAAAATGACCTGGCGAAGTTACAGTTAACAGCCCCAGTA 60  
 32 CGACGGGACAGCGCTACCTTCTCGCCTTTTCAAGTACATACTGGGGCTGTTAACTGTAAC 60  
 33 TAGCGCTGTCCGTCGTGTGAAAAGCATTTGGCGTCTGATCGGTTGGGGCCAGATTC AAG 60  
 34 CAGCGGGCACGGCGCACTTCCAGGGTAGCTCCAGCTTGAATCTGGCCCCAACC 54

## Appendix D: Primers

EIF2•-F:

5'-GAATTCGCGGCCGCTTCTAGAGATTAAAGAGGAGAAATACCATATGGCGAACCTGGAATGCCG-3'

EIF2•-R:

5'-CTGCAGCGGCCGCTACTAGTAGGATCCTTATTAATCCGCATGCACGCCGCTAT-3'

EIF2•-F:

5'-GAATTCGCGGCCGCTTCTAGAGATTAAAGAGGAGAAATACCATATGGCGGACGAAGAACAGAT-3'

EIF2•-R:

5'-CTGCAGCGGCCGCTACTAGTAGGATCCTTATTAGGTGCCCGCCTTACGACGGC-3'

EIF2•I-F:

5'-GAATTCGCGGCCGCTTCTAGAGATTAAAGAGGAGAAATACCATATGGCGCGTCGGGGACTGAT-3'

EIF2•I-R:

5'-CTGCAGCGGCCGCTACTAGTAGGAACCCCCAGCCACGCCGC-3'

EIFGII-F:

5'-GAATTCGCGGCCGCTTCTAGAGCTTTACCAGCCCACCGAATA-3'

EIFGII-R:

5'-CTGCAGCGGCCGCTACTAGTAGGATCCTTATTACAGCGGGCACGGCGGCACTT-3'

M13F:

5'-GTAAAACGACGGCCAGT-3'

M13R:

5'-AACAGCTATGACCATG-3'

VF2:

5'-TGCCACCTGACGTCTAAGAA-3'

VR:

5'-ATTACCGCCTTTGAGTGAGC-3'

T7pro:

5'-TAATACGACTCACTATAGGG -3'

T7term:

5'-GCTAGTTATTGCTCAGCGG -3'

## Appendix E – Codon Usage Tables

For each species is listed the three nucleotide codon, the single letter abbreviation for the amino acid it encodes, and the codon percent usage. The percent usage is determined by how often the codon appears in the coding regions of the species' genome. (adapted from Codon Usage Database at <http://www.kazusa.or.jp/codon/>)

### *Triticum aestivum*

|            |            |            |            |
|------------|------------|------------|------------|
| TTT F 0.31 | TCT S 0.16 | TAT Y 0.29 | TGT C 0.30 |
| TTC F 0.69 | TCC S 0.25 | TAC Y 0.71 | TGC C 0.70 |
| TTA L 0.05 | TCA S 0.12 | TAA * 0.00 | TGA * 1.00 |
| TTG L 0.18 | TCG S 0.22 | TAG * 0.00 | TGG W 1.00 |
| CTT L 0.14 | CCT P 0.15 | CAT H 0.40 | CGT R 0.05 |
| CTC L 0.31 | CCC P 0.27 | CAC H 0.60 | CGC R 0.26 |
| CTA L 0.07 | CCA P 0.27 | CAA Q 0.53 | CGA R 0.02 |
| CTG L 0.26 | CCG P 0.32 | CAG Q 0.47 | CGG R 0.23 |
| ATT I 0.26 | ACT T 0.20 | AAT N 0.3  | AGT S 0.04 |
| ATC I 0.52 | ACC T 0.33 | AAC N 0.63 | AGC S 0.22 |
| ATA I 0.23 | ACA T 0.16 | AAA K 0.16 | AGA R 0.02 |
| ATG M 1.00 | ACG T 0.31 | AAG K 0.84 | AGG R 0.41 |
| GTT V 0.17 | GCT A 0.17 | GAT D 0.49 | GGT G 0.14 |
| GTC V 0.29 | GCC A 0.29 | GAC D 0.51 | GGC G 0.51 |
| GTA V 0.14 | GCA A 0.22 | GAA E 0.38 | GGA G 0.07 |
| GTG V 0.40 | GCG A 0.32 | GAG E 0.62 | GGG G 0.28 |

### *E. coli*

|            |            |            |            |
|------------|------------|------------|------------|
| TTT F 0.59 | TCT S 0.17 | TAT Y 0.60 | TGT C 0.47 |
| TTC F 0.41 | TCC S 0.14 | TAC Y 0.40 | TGC C 0.53 |
| TTA L 0.15 | TCA S 0.15 | TAA X 0.60 | TGA X 0.31 |
| TTG L 0.13 | TCG S 0.13 | TAG X 0.09 | TGG W 1.00 |
| CTT L 0.12 | CCT P 0.19 | CAT H 0.59 | CGT R 0.34 |
| CTC L 0.10 | CCC P 0.14 | CAC H 0.41 | CGC R 0.34 |
| CTA L 0.04 | CCA P 0.21 | CAA Q 0.34 | CGA R 0.07 |
| CTG L 0.46 | CCG P 0.47 | CAG Q 0.66 | CGG R 0.12 |
| ATT I 0.49 | ACT T 0.19 | AAT N 0.52 | AGT S 0.17 |
| ATC I 0.37 | ACC T 0.38 | AAC N 0.48 | AGC S 0.23 |
| ATA I 0.14 | ACA T 0.19 | AAA K 0.73 | AGA R 0.08 |
| ATG M 1.00 | ACG T 0.24 | AAG K 0.27 | AGG R 0.05 |
| GTT V 0.29 | GCT A 0.19 | GAT D 0.64 | GGT G 0.34 |
| GTC V 0.20 | GCC A 0.26 | GAC D 0.36 | GGC G 0.35 |
| GTA V 0.17 | GCA A 0.24 | GAA E 0.67 | GGA G 0.15 |
| GTG V 0.34 | GCG A 0.31 | GAG E 0.33 | GGG G 0.16 |



## Appendix F – Sequencing Data

For each sequence, the correct coding sequences as presented in Appendix C are listed as ‘Query’ (top line), and the actual sequenced sample as ‘Sbjct’ (bottom line). The sequences were aligned using the NCBI BLAST specialized nucleotide alignment tool: ([http://blast.ncbi.nlm.nih.gov/Blast.cgi?PAGE\\_TYPE=BlastSearch&PROG\\_DEF=blastn&BLAST\\_PROG\\_DEF=megaBlast&SHOW\\_DEFAULTS=on&BLAST\\_SPEC=blast2seq&LINK\\_LOC=align2seq](http://blast.ncbi.nlm.nih.gov/Blast.cgi?PAGE_TYPE=BlastSearch&PROG_DEF=blastn&BLAST_PROG_DEF=megaBlast&SHOW_DEFAULTS=on&BLAST_SPEC=blast2seq&LINK_LOC=align2seq)). For Sequences 1-4, the engineered ‘backdoor’ restriction site is noted in red, and only the 3’ or 5’ end of that sequence was used, as indicated. Primers used for sequencing are noted in ().

### Sequence 1: eIF2 $\alpha$ in Zero Blunt, Correct 5’ end (M13F)

```
Query 79 GAATTCGCGCGCGCTTCTAGAGATTAAGAGGAGAAATACCATATGGCGAACCTGGAATG 138
|
|
|
Sbjct 1 GAATTCGCGCGCGCTTCTAGAGATTAAGAGGAGAAATACCATATGGCGAACCTGGAATG 60

Query 139 CCGTATGTATGAACCGCGTTTTCCCGAAGTGGATGCTGCGGTGATGATTCAGGTGAAACA 198
|
|
|
Sbjct 61 CCGTATGTATGAACCGCGTTTTCCCGAAGTGGATGCTGCGGTGATGATTCAGGTGAAACA 120

Query 199 TATTGCGGATATGGGCGCGTATGTGAGCCTGCTGGAATATAACAATGTGGAAGGCATGAT 258
|
|
|
Sbjct 121 TATTGCGGATATGGGCGCGTATGTGAGCCTGCTGGAATATAACAATGTGGAAGGCATGAT 180

Query 259 TCTGTTTTCCGAGCTGAGCCGTCGCCGCATTCGGAGCATAAGCAGCCTGATAAAAGTGGG 318
|
|
|
Sbjct 181 TCTGTTTTCCGAGCTGAGCCGTCGCCGCATTCGGAGCATAAGCAGCCTGATAAAAGTGGG 240

Query 319 TCGTCAAGAACCGGCGATAGTTCTTCGTGTGGATCGTGATAAAGGCTATATTGACTTGTC 378
|
|
|
Sbjct 241 TCGTCAAGAACCGGCGATAGTTCTTCGTGTGGATCGTGATAAAGGCTATATTGACTTGTC 300

Query 379 CAAGCGCCGTGTTTTCCGAAGAAGAAGCGCGTAGCTGCGAGGATAAATACAATAAGAGCAA 438
|
|
|
Sbjct 301 CAAGCGCCGTGTTTTCCGAAGAAGAAGCGCGTAGCTGCGAGGATAAATACAATAAGAGCAA 360

Query 439 GCTGGTCCATAGCATTATGCGTACAGTTGCTGAGACCCTGGAGATTGATCTGGAACCGAT 498
|
|
|
Sbjct 361 GCTGGTCCATAGCATTATGCGTACAGTTGCTGAGACCCTGGAGATTGATCTGGAACCGAT 420

Query 499 CTATCAGCGGATTGGCTGGCCGCTGTATCGTAAATACGGCCACGCATTTGAAGCGTTCAA 558
|
|
|
Sbjct 421 CTATCAGCGGATTGGCTGGCCGCTGTATCGTAAATACGGCCACGCATTTGAAGCGTTCAA 480

Query 559 GCTGATTGTTGCGGACCCTGACGCCATATTAGACGTGCTGACCTATGAAGAGCGTGAAAC 618
|
|
|
Sbjct 481 GCTGATTGTTGCGGACCCTGACGCCATATTAGACGTGCTGACCTATGAAGAGCGTGAAAC 540

Query 619 CGGGCCAGATGGCCAAGAAGTGACCAAAGTGGTACCAGCAGTCACCCCGAAATAAAGGA 678
|
|
|
Sbjct 541 CGGGCCAGATGGCCAAGAAGTGACCAAAGTGGTACCAGCAGTCACCCCGAAATAAAGGA 600

Query 679 AACCCTGGTGCAGAACATTTCGTCGTCGGA-CACACCACAACCGCTGAAAATTCGTGCGGA 737
|
|
|
Sbjct 601 AACCCTGGTGCAGAACATTTCGTCGTCGGA GACACCACAACCGCTGAAAATTCGTGCGGA 660

Query 738 CGTGAAAATGAAAATGCTTTTCAGTTTGATGG-G--C---ATATCAAACAGGCGATGCGTAA 791
|
|
|
Sbjct 661 CGTGAAAATGAAAATGCTTTTCAGTTTGATGGCGTGCATATCAAACAGGCGATGCGTAA 720

Query 792 GGCGGAGGCTGCGGGCAATACCAATTGTCCGGTAAAGATTAAGTTAGTGGCGCCTCCACT 851
|
|
|
Sbjct 721 GGCGGAGGCTGCGGGCAATACCAATTGTCCGGTAAAGATTAAGTTAGTGGCGCCTCCACT 780
```

Query 852 GTATGTACTGACCACCCAGACACTGGACAAAGATCAGGGCATTAGCGTTTTAACCGATGC 911  
 |||||||  
 Sbjct 781 GTATGTACTGACCACCCAGACACTGGACAAAGATCAGGGCATTAGCGTTTTAACCGATGC 840

Query 912 GGTCAAAGCGTGCACTGCGGAAATCGAAAAACATANGG-CAAACGGTGGTGAAAGAAGC 970  
 |||||||  
 Sbjct 841 GGTCAAAGCGTGCACTGCGGAAATCGAAAAACATAAGGGCAAACGGTGGTGAAAGAAGC 900

Query 971 CCCACGCGCGNGTCAGAACGTGAAGATAAACTGCTGAATGCCCACTGGNTACTCTGGT 1030  
 |||||||  
 Sbjct 901 CCCACGCGCGGTGTGAGAACGTGAAGATAAACTGCTGAATGCCCACTGGATACTCTGGT 960

Query 1031 AGA-CAAA-TGNNGAAGTCGCNNNN-ACGATGATAGCNNNgnnnan-anGNTACCNGCAT 1086  
 ||| ||| || ||||| ||||| || | | ||| ||||  
 Sbjct 961 AGAACAAAATGCGGAAGTCGAGGCGACGATGATAGCGAGGACGAAGAGGATACCGGCAT 1020

Query 1087 GGGCGACNTCGATCTGANNA-TANCGGCGTGCATG 1120  
 ||||| ||||| | || |||||  
 Sbjct 1021 GGGCGACATCGATCTGACCAATAGCGGCGTGCATG 1055

**Sequence 2: eIF2 $\alpha$  in Zero Blunt, Correct 3' end (M13R)**

Query 80 CTGCAGCGCCGCTACTAGTAGGATCCTTATTAATCCGCATGCACGCCGCTATTGGTCAG 139  
 |||||||  
 Sbjct 1093 CTGCAGCGCCGCTACTAGTAGGATCCTTATTAATCCGCATGCACGCCGCTATTGGTCAG 1034

Query 140 ATCGATGTCGCCCATGCCGGTATCCTCTTCGTCCTCGCTATCATCGTCGCCTGCGACTTC 199  
 |||||||  
 Sbjct 1033 ATCGATGTCGCCCATGCCGGTATCCTCTTCGTCCTCGCTATCATCGTCGCCTGCGACTTC 974

Query 200 CGCATTTTGTCTTACCAGAGTATCCAGTTGGGCATTACAGAGTTTATCTTCACGTTCTGA 259  
 |||||||  
 Sbjct 973 CGCATTTTGTCTTACCAGAGTATCCAGTTGGGCATTACAGAGTTTATCTTCACGTTCTGA 914

Query 260 CACCGCGCGTGGGGCTTCTTTCACCACCAGTTTGCCTTATGTTTTTCGATTTCCGCAGT 319  
 |||||||  
 Sbjct 913 CACCGCGCGTGGGGCTTCTTTCACCACCAGTTTGCCTTATGTTTTTCGATTTCCGCAGT 854

Query 320 GCACGCTTTGACCGCATCGGTTAAAAAGCTAATGCCCTGATCTTTGTCAGTGTCTGGGT 379  
 |||||||  
 Sbjct 853 GCACGCTTTGACCGCATCGGTTAAAAAGCTAATGCCCTGATCTTTGTCAGTGTCTGGGT 794

Query 380 GGTCAGTACATACAGTGGAGGCGCCACTAACTTAATCTTTACCGGACAATTGGTATTGCC 439  
 |||||||  
 Sbjct 793 GGTCAGTACATACAGTGGAGGCGCCACTAACTTAATCTTTACCGGACAATTGGTATTGCC 734

Query 440 CGCAGCCTCCGCCTTACGCATCGCCTGTTGATATGCAGCAGCCATCAAACGAAAGCA 499  
 |||||||  
 Sbjct 733 CGCAGCCTCCGCCTTACGCATCGCCTGTTGATATGCAGCAGCCATCAAACGAAAGCA 674

Query 500 TTTCAATTTCCACGTCGCGACGAATTTTACGCGGTTGTGGTGTATCCGACGACGAATGTT 559  
 |||||||  
 Sbjct 673 TTTCAATTTCCACGTCGCGACGAATTTTACGCGGTTGTGGTGTATCCGACGACGAATGTT 614

Query 560 CTGCACCAGGGTTTCCTTTATTTCCGGGGTGACTGCTGGTACCCTTTGGTCACTTCTTG 619  
 |||||||  
 Sbjct 613 CTGCACCAGGGTTTCCTTTATTTCCGGGGTGACTGCTGGTACCCTTTGGTCACTTCTTG 554

Query 620 GCCATCTGGCCCGGTTTACGCTCTTCATAGGTGAGCAGCTCTAATATGGCGTCAGG-TC 678  
 |||||||  
 Sbjct 553 GCCATCTGGCCCGGTTTACGCTCTTCATAGGTGAGCAGCTCTAATATGGCGTCAGGGTC 494

Query 679 CGCAACAATCAGCTTGAACGCTTCAAATGCGTGGCCGTATTTACGATACAGCGCCAGCC 738  
 |||||||  
 Sbjct 493 CGCAACAATCAGCTTGAACGCTTCAAATGCGTGGCCGTATTTACGATACAGCGCCAGCC 434

Query 739 AATCCGCTGATAGATCGGTTCCAGATCAATCTCCAGGGTCTCAGCAACGTGACGCATAAT 798  
 |||||||  
 Sbjct 433 AATCCGCTGATAGATCGGTTCCAGATCAATCTCCAGGGTCTCAGCAACGTGACGCATAAT 374

```

Query 799 GCTATGGACCAGCTTGCTCTTAT-GTATTTATCCTCGCAGCTACGCGCTTCTTCTTCGGA 857
          |||
Sbjct 373 GCTATGGACCAGCTTGCTCTTATTGTATTTATCCTCGCAGCTACGCGCTTCTTCTTCGGA 314

Query 858 AACACGGCGCTTGACAAAGTCAATATAGCCTTTATCACGATCCACACGAAGAACTATCGC 917
          |||
Sbjct 313 AACACGGCGCTTGACAAAGTCAATATAGCCTTTATCACGATCCACACGAAGAACTATCGC 254

Query 918 CGGTTCTTGACGACCCACTTTTATCANN-TGCTTATGCTCCGAATGCGGCGANGGNTCAG 976
          |||
Sbjct 253 CGGTTCTTGACGACCCACTTTTATCAGGCTGCTTATGCTCCGAATGCGGCGACGGCTCAG 194

Query 977 CTCGNAAAACAGAATCATGCCTTCNACATTGTNATATTNCAGC 1019
          |||
Sbjct 193 CTCGGAAAACAGAATCATGCCTTCCACATTGTTATATTCCAGC 151

```

### Sequence 3: eIF2 $\beta$ in Zero Blunt, Correct 5' end (M13F)

```

Query 81 CTGCAGCGGCCGCTACTAGTAGGATCCTTATTAGGTGCCCGCCTTACGACGGCCTACCTG 140
          |||
Sbjct 886 CTGCAGCGGCCGCTACTAGTAGGATCCTTATTAGGTGCCCGCCTTACGACGGCCTACCTG 827

Query 141 GGCAACGAACCCGGCTTTAATCGGCGCCACGCTA-GGCTACTGCCGACTGTTTCGCAACG 199
          |||
Sbjct 826 GGCAACGAACCCGGCTTTAATCGGCGCCACGCTACGGCTACTGCCGACTGTTTCGCAACG 767

Query 200 CA-GAAAAACAGACGATTCTCTTTGCTCAGAAATGGTATCCGGGCTCTTGACGCCATTGCA 258
          |||
Sbjct 766 CAGGAAAAACAGACGATTCTCTTTGCTCAGAAATGGTATCCGGGCTCTTGACGCCATTGCA 707

Query 259 AATCACATATCATTGATGTAACGCCGAGGATCGCTTCAAAATCTTGGGAGCGAAGCG 317
          |||
Sbjct 706 AATCACATATTCATTGATGTAACGCCGAGGATCGCTTCAAAATCTTGGGAGCGAAGCG 647

Query 318 GCCTTTAATGACCAGACGCTGCTGCCATCCAGGCTGCCGCTGGTGCCATTTCCGCAAG 377
          |||
Sbjct 646 GCCTTTAATGACCAGACGCTGCTGCCATCCAGGCTGCCGCTGGTGCCATTTCCGCAAG 587

Query 378 CAGAAACATCATCACATGTTCCGGCTGACGATGCATGGTTTTGCACAGATCCATGAAATT 437
          |||
Sbjct 586 CAGAAACATCATCACATGTTCCGGCTGACGATGCATGGTTTTGCACAGATCCATGAAATT 527

Query 438 CACAAACACGGtttttttGGTGCCCTTCACGCAAGACCTGCGGTGGACGCATCA---TACG 494
          |||
Sbjct 526 CACAAACACGGTTTTTTTGGTGCCCTTCACGCAAGACCTGCGGTGGACGCATCACGGTACG 467

Query 495 TCGACGGTCTCCCGCCAGGCTGGATTATTCTCGCGCAGAATATTGAACACACGGCCCAA 554
          |||
Sbjct 466 TCGACGGTCTCCCGCCAGGCTGGATTATTCTCGCGCAGAATATTGAACACACGGCCCAA 407

Query 555 CAGCTCATCATATTTGTAATCACGATCGGTTCCCTTCCCACGGATACTGGGTGGCACCGCC 614
          |||
Sbjct 406 CAGCTCATCATATTTGTAATCACGATCGGTTCCCTTCCCACGGATACTGGGTGGCACCGCC 347

Query 615 CAGCACAAATGCCCTCTCCTTGTTTCATCCTCGCCACCTGATCGTCCAGGGTATCTTCGCC 674
          |||
Sbjct 346 CAGCACAAATGCCCTCTCCTTGTTTCATCCTCGCCACCTGATCGTCCAGGGTATCTTCGCC 287

Query 675 ATCACC GGCTTCGACAAGGCTGGGGTCCAGTTCTACTAACTTCTTTTTCTTTTCTTCAA 734
          |||
Sbjct 286 ATCACC GGCTTCGACAAGGCTGGGGTCCAGTTCTACTAACTTCTTTTTCTTTTCTTCAA 227

Query 735 GCCCACAAAGCTCGCCTCGCCGCTTTCCGGTACGCTCAGGCCTTCGGTCTTTTCGGCCAA 794
          |||
Sbjct 226 GCCCACAAAGCTCGCCTCGCCGCTTTCCGGTACGCTCAGGCCTTCGGTCTTTTCGGCCAA 167

Query 795 TTTATCCAATTTCATCCGCTGGGTCCTGAATCACCActttcttcttcttcttcttAGT 854
          |||

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Sbjct 166 TTTATCCACTTCATCCGCTGGGTCTGAATCACCACCTTCTTCTTCTTCTTCTTAGT 107
Query 855 CGGGTCAAACGGCGCAATCTCGGTTCGCTTCTTCTTACGTTCCATCTGTTCTTCGTCGGC 914
      |||
Sbjct 106 CGGGTCAAACGGCGCAATCTCGGTTCGCTTCTTCTTACGTTCCATCTGTTCTTCGTCGGC 47
Query 915 CATATGG 921
      |||
Sbjct 46 CATATGG 40

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**Sequence 4: eIF2β in Zero Blunt, Correct 3' end (M13R)**

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Query 101 CTGCAGCGGCCGCTACTAGTAGGATCCTTATTAGGTGCCCGCCTTACGACGGCCTACCTG 160
      |||
Sbjct 886 CTGCAGCGGCCGCTACTAGTAGGATCCTTATTAGGTGCCCGCCTTACGACGGCCTACCTG 827
Query 161 GGCAACGAACCCGGCTTTAATCGGGCCACGCTACGGCTACTGCCGCACTGTTCCGAACG 220
      |||
Sbjct 826 GGCAACGAACCCGGCTTTAATCGGGCCACGCTACGGCTACTGCCGCACTGTTCCGAACG 767
Query 221 CAGGAAAAACAGACGATTCTCTTTGCTCAGAAATGGTATCCGGGCTCTTGACGCCATTGCA 280
      |||
Sbjct 766 CAGGAAAAACAGACGATTCTCTTTGCTCAGAAATGGTATCCGGGCTCTTGACGCCATTGCA 707
Query 281 AATCACATATTGATGTAAACGCCGAGGATCGCTTCAAATTTCTGGGAGCGAAGCG 340
      |||
Sbjct 706 AATCACATATTGATGTAAACGCCGAGGATCGCTTCAAATTTCTGGGAGCGAAGCG 647
Query 341 GCCTTTAATGACCAGACGCTGCTGCCCATCCAGGCTGCCGCTGGTGCCCATTTCCGCAAG 400
      |||
Sbjct 646 GCCTTTAATGACCAGACGCTGCTGCCCATCCAGGCTGCCGCTGGTGCCCATTTCCGCAAG 587
Query 401 CAGAAACATCATCACATGTTCCGGCTGACGATGCATGGTTTGCACAGATCCATGAAATT 460
      |||
Sbjct 586 CAGAAACATCATCACATGTTCCGGCTGACGATGCATGGTTTGCACAGATCCATGAAATT 527
Query 461 CACAAACACGGTttttttGGTGCCCTCACGCAAGACCTGCGGTGGACGCATCACGGTACG 520
      |||
Sbjct 526 CACAAACACGGTTTTTTGGTGCCCTCACGCAAGACCTGCGGTGGACGCATCACGGTACG 467
Query 521 TCGACGGTCTCCCGCCAGGCTGGATTATTCTCGCGCAGAATATTGAACACACGGCCCAA 580
      |||
Sbjct 466 TCGACGGTCTCCCGCCAGGCTGGATTATTCTCGCGCAGAATATTGAACACACGGCCCAA 407
Query 581 CAGCTCATCATATTGTAATCAGATCGGTTTCCACGGATACTGGGTG-CACCGCC 638
      |||
Sbjct 406 CAGCTCATCATATTGTAATCAGATCGGTTTCCACGGATACTGGGTGGCACCGCC 347
Query 639 CAGCACA-TGCCCTCTCCTTGTTTCATCCTCGCCACCTGATCGTCCAGGGTATCTTCGCC 697
      |||
Sbjct 346 CAGCACAATGCCCTCTCCTTGTTTCATCCTCGCCACCTGATCGTCCAGGGTATCTTCGCC 287
Query 698 ATCACCGGCTTCGACAAGGCTGGGGTCCAGTTCTACTAACTTCTTTTCTTTTCTTCAA 757
      |||
Sbjct 286 ATCACCGGCTTCGACAAGGCTGGGGTCCAGTTCTACTAACTTCTTTTCTTTTCTTCAA 227
Query 758 GCCCACAAAGCTCGCCTCGCCGCTTTCGGTACGCTCAGGCCTTCGGTCTTTTCGGCCAA 817
      |||
Sbjct 226 GCCCACAAAGCTCGCCTCGCCGCTTTCGGTACGCTCAGGCCTTCGGTCTTTTCGGCCAA 167
Query 818 TTTATCCACTTCATCCGCTGGGTCTGAATCACCActtttcttcttcttcttcttAGT 877
      |||
Sbjct 166 TTTATCCACTTCATCCGCTGGGTCTGAATCACCACCTTCTTCTTCTTCTTCTTAGT 107
Query 878 CGGGTCAAACGGCGCAATCTCGGTTCGCTTCTTCTTACGTTCCATCTGTTCTTCGTCGGC 937
      |||
Sbjct 106 CGGGTCAAACGGCGCAATCTCGGTTCGCTTCTTCTTACGTTCCATCTGTTCTTCGTCGGC 47
Query 938 CATATGGTATTCTCCTCTTAATCTC 964
      |||

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Sbjct 79 GAATTCGCGGCCGGCTTCTAGAGCTTTACCAGCCACCGAATATGATTGTTATTCGAAGC 138

Query 60 TTCGATGTGAATAAACCGGGCAGTGAAGTGGATGAAATTCGTGGCGCGTGGCTGGGGT 119  
 |||  
 Sbjct 139 TTCGATGTGAATAAACCGGGCAGTGAAGTGGATGAAATTCGTGGCGCGTGGCTGGGGT 198

Query 120 TCCATTTTACGTGGCGTGTTCGGGTGAACCAGAACATTGAAGTGCCTCCGGCATTGTG 179  
 |||  
 Sbjct 199 TCCATTTTACGTGGCGTGTTCGGGTGAACCAGAACATTGAAGTGCCTCCGGCATTGTG 258

Query 180 ATGAAAGACGAATCGGGGAACATAAAGTGCACGCCCATTTATAGCCGTATTGTGAGCTTA 239  
 |||  
 Sbjct 259 ATGAAAGACGAATCGGGGAACATAAAGTGCACGCCCATTTATAGCCGTATTGTGAGCTTA 318

Query 240 TATGCGGAACAAAATGAGCTGCAATTCGCGGTGCCTGGGGCCTGATTGGGGTGGGCACG 299  
 |||  
 Sbjct 319 TATGCGGAACAAAATGAGCTGCAATTCGCGGTGCCTGGGGCCTGATTGGGGTGGGCACG 378

Query 300 ACTATGGACCCGACCTGACTCGTGC GGATCGTCTGGTGGGCAAGTG-CTGGGAGAAAT 358  
 |||  
 Sbjct 379 ACTATGGACCCGACCTGACTCGTGC GGATCGTCTGGTGGGCAAGTGCTGGGAGAAAT 438

Query 359 TGGTAGCCTGCCGGATGTCTTTGTTGAATTAGAGATTAAC TTTTTCAGCTGCGTCTCT 418  
 |||  
 Sbjct 439 TGGTAGCCTGCCGGATGTCTTTGTTGAATTAGAGATTAAC TTTTTCAGCTGCGTCTCT 498

Query 419 GCTGGGTGTTCCGACCAAAGGGACCGAGAAAGCGGGAAAGT GAGCAAATTGACCAAGGG 478  
 |||  
 Sbjct 499 GCTGGGTGTTCCGACCAAAGGGACCGAGAAAGCGGGAAAGT GAGCAAATTGACCAAGGG 558

Query 479 GGAAATCTGATGTTAAACATCGGAAGCATGAGCACCGGAGCGCGTGTGGTTGCGGTCAA 538  
 |||  
 Sbjct 559 GGAAATCTGATGTTAAACATCGGAAGCATGAGCACCGGAGCGCGTGTGGTTGCGGTCAA 618

Query 539 AAATGACCTGGCGAAGTTACAGTTAACAGCCCCAGTATGTACTTCGAAAGGCGAGAAGGT 598  
 |||  
 Sbjct 619 AAATGACCTGGCGAAGTTACAGTTAACAGCCCCAGTATGTACTTCGAAAGGCGAGAAGGT 678

Query 599 AGCGCTGTCCCGTCTGTGGAAAAGCATTGGCGTCTGATCGGTTGGGGCCAGATTCAAGC 658  
 |||  
 Sbjct 679 AGCGCTGTCCCGTCTGTGGAAAAGCATTGGCGTCTGATCGGTTGGGGCCAGATTCAAGC 738

Query 659 TGGAGCTACCTGGAAGTGCCGCCGTGCCCGCTGTAATAAGGATCCTACTAGTAGCGGCC 718  
 |||  
 Sbjct 739 TGGAGCTACCTGGAAGTGCCGCCGTGCCCGCTGTAATAAGGATCCTACTAGTAGCGGCC 798

Query 719 GCTGCAG 725  
 |||  
 Sbjct 799 GCTGCAG 805

### Sequence 7: Complete eIF2 $\alpha$ in BioBrick (VF2)

Query 1 GAATTCGCGGCCGGCTTCTAGAGATTAAGAGGAGAAATACCATATGGCGAACCTGGAATG 60  
 |||  
 Sbjct 89 GAATTCGCGGCCGGCTTCTAGAGATTAAGAGGAGAAATACCATATGGCGAACCTGGAATG 148

Query 61 CCGTATGTATGAACCGCGTTTTCCCGAAGTGGATGCTGCGGTGATGATTCAGGTGAAACA 120  
 |||  
 Sbjct 149 CCGTATGTATGAACCGCGTTTTCCCGAAGTGGATGCTGCGGTGATGATTCAGGTGAAACA 208

Query 121 TATTGCGGATATGGGCGCGTATGTGAGCCTGCTGGAATATAACAATGTGGAAGGCATGAT 180  
 |||  
 Sbjct 209 TATTGCGGATATGGGCGCGTATGTGAGCCTGCTGGAATATAACAATGTGGAAGGCATGAT 268

Query 181 TCTGTTTTCCGAGCTGAGCCGTCCCGCATTCGGAGCATAAGCAGCCTGATAAAAGTGGG 240  
 |||  
 Sbjct 269 TCTGTTTTCCGAGCTGAGCCGTCCCGCATTCGGAGCATAAGCAGCCTGATAAAAGTGGG 328

Query 241 TCGTCAAGAACCGGCGATAGTTCTTCGTGIGGATCGTGATAAAGGCTATATTGACTTGTG 300

|       |     |  |     |
|-------|-----|--|-----|
| Sbjct | 329 | <br>TCGTC AAGAACC GCGATAGTTCCTTCGTGTGGATCGTGATAAAGGCTATATTGACTTGTCTC | 388 |
| Query | 301 | CAAGCGCCGTGTTTCCGAAGAAGAAGCGCGTAGCTGCGAGGATAAAATACAATAAGAGCAA        | 360 |
| Sbjct | 389 | <br>CAAGCGCCGTGTTTCCGAAGAAGAAGCGCGTAGCTGCGAGGATAAAATACAATAAGAGCAA    | 448 |
| Query | 361 | GCTGGTCCATAGCATTATGCGTACGTTGCTGAGACCCTGGAGATTGATCTGGAACCGAT          | 420 |
| Sbjct | 449 | <br>GCTGGTCCATAGCATTATGCGTACGTTGCTGAGACCCTGGAGATTGATCTGGAACCGAT      | 508 |
| Query | 421 | CTATCAGCGGATTGGCTGGCCGCTGTATCGTAAATACGGCCACGCATTTGAAGCGTTCAA         | 480 |
| Sbjct | 509 | <br>CTATCAGCGGATTGGCTGGCCGCTGTATCGTAAATACGGCCACGCATTTGAAGCGTTCAA     | 568 |
| Query | 481 | GCTGATTGTTGCGGACCCTGACGCCATATTAGACGTGCTGACCTATGAAGAGCGTGAAAC         | 540 |
| Sbjct | 569 | <br>GCTGATTGTTGCGGACCCTGACGCCATATTAGACGTGCTGACCTATGAAGAGCGTGAAAC     | 628 |
| Query | 541 | CGGGCCAGATGGCCAAGAAGTGACCAAAGTGGTACCAGCAGTCACCCCGGAAATAAAGGA         | 600 |
| Sbjct | 629 | <br>CGGGCCAGATGGCCAAGAAGTGACCAAAGTGGTACCAGCAGTCACCCCGGAAATAAAGGA     | 688 |
| Query | 601 | AACCTGGTGCAGAACATTTCGTCTCGGATGACACCACAACCGCTGAAAATTCGTGCGGA          | 660 |
| Sbjct | 689 | <br>AACCTGGTGCAGAACATTTCGTCTCGGATGACACCACAACCGCTGAAAATTCGTGCGGA      | 748 |
| Query | 661 | CGTGAAAATGAAAATGCTTTCAGTTTGATGGCGTGCATATCAAACAGGCGATGCGTAA           | 720 |
| Sbjct | 749 | <br>CGTGAAAATGAAAATGCTTTCAGTTTGATGGCGTGCATATCAAACAGGCGATGCGTAA       | 808 |
| Query | 721 | GGCGGAGGCTGCGGGCAATACCAATTGTCCGGTAAAGATTAAGTTAGTGGCGCCTCCACT         | 780 |
| Sbjct | 809 | <br>GGCGGANGCTGCGGGCAATACCAATTGTCCGGTAAAGATTAAGTTAGTGGCGCCTCCACT     | 868 |
| Query | 781 | GTATGTACTGACCACCAGACACTGGACAAAGATCAGGGCATTAGCGTTTAAACCGATGC          | 840 |
| Sbjct | 869 | <br>GTATGTACTGACCACCAGACACTGGACAAAGATCAGGGCATTAGCGTTTAAACCGATGC      | 928 |
| Query | 841 | GGTCAAAGCGTGCCTGCGGAAATCGAAAAACATAAGGGCAAAGTGG-TGGTGAAAGAAG          | 899 |
| Sbjct | 929 | <br>GGTCAAAGCGTGCCTGCGGAAATCGAAAAACATAANNNN-AACTGGGTGGTGAAAGAAG      | 987 |
| Query | 900 | CCCCACGCGGGTGTGAGAACGTGAAGATAAACTGCTGAATGCCCAACT 948                 |     |
| Sbjct | 988 | <br>CCCCACGCGGGTGTGAGAACGTGAAGATAAACTGCTGAATGCCCAACT 1034            |     |

### Sequence 8: Complete eIF2 $\alpha$ in BioBrick (VR)

|       |      |   |     |
|-------|------|---|-----|
| Query | 165  | ATGTGGAAGGCATGATTCTGTTTCCGAGCTGAGCCGTCGCGCATTTCGGAGCATAAGCA       | 224 |
| Sbjct | 1049 | <br>ATGTGGANGGCATGNTTCTGTTTTCGAGCTGAGCCGTCGNCGCATTTCGNAGCATAAGCA  | 990 |
| Query | 225  | GCCTGATAAAAAGTGGGTCGTCAAGAACC GCGATAGTTCCTCGTGTGGATCGTGATAAAG     | 284 |
| Sbjct | 989  | <br>-NNNGATAAAAAGTGGGTCGTCAAGANCCGGCGATAGTTCCTCGTGTGGATCGTGATAAAG | 931 |
| Query | 285  | GCTATATTGACTTGTCCAAGCGCCGTGTTTCCGAAGAAGAAGCGCGTAGCTGCGAGGATA      | 344 |
| Sbjct | 930  | <br>GCTATATTGACTTNNCCAAGCGCCGTGTTTNGAAGAAGAAGCGCGTAGCTGCGAGGATA   | 871 |
| Query | 345  | AATACAATAAGAGCAAGCTGGTCCATAGCATTATGCGTCACGTTGCTGAGACCCTGGAGA      | 404 |
| Sbjct | 870  | <br>AATACAATAAGAGCAAGCTGGTCCATAGCATTATGCGTCACGTTGCTGAGACCCTGGAGA  | 811 |
| Query | 405  | TTGATCTGGAACCGATCTATCAGCGGATTGGCTGGCCGCTGTATCGTAAATACGGCCACG      | 464 |
| Sbjct | 810  | <br>TTGATCTGGAACCGATCTATCAGCGGATTGGCTGGCCGCTGTATCGTAAATACGGCCACG  | 751 |
| Query | 465  | CATTTGAAGCGTTC AAGCTGATTGTTGCGGACCCTGACGCCATATTAGACGTGCTGACCT     | 524 |

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Sbjct 750      |||||||||||||||||||||||||||||||||||||||||||||||||||||||||| 691
                CATTTGAAGCGTTCAAGCTGATTGTTGCGGACCCTGACGCCATATTAGACGTGCTGACCT
Query 525      ATGAAGAGCGTGAAACCGGGCCAGATGGCCAAGAAGTGACCAAAGTGGTACCAGCAGTCA 584
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 690      ATGAAGAGCGTGAAACCGGGCCAGATGGCCAAGAAGTGACCAAAGTGGTACCAGCAGTCA 631
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 585      CCCCAGAAATAAAGGAAACCCCTGGTGCGAACATTTCGTTCGTTCGGATGACACCACAACCGC 644
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 630      CCCCAGAAATAAAGGAAACCCCTGGTGCGAACATTTCGTTCGTTCGGATGACACCACAACCGC 571
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 645      TGAAAATTCGTGCGGACGTGGAAATGAAATGCTTTCAGTTTGATGGCGTGCTGCATATCA 704
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 570      TGAAAATTCGTGCGGACGTGGAAATGAAATGCTTTCAGTTTGATGGCGTGCTGCATATCA 511
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 705      AACAGGCGATGCGTAAGGCGGAGGCTGCGGGCAATACCAATTGTCCGGTAAAGATTAAGT 764
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 510      AACAGGCGATGCGTAAGGCGGAGGCTGCGGGCAATACCAATTGTCCGGTAAAGATTAAGT 451
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 765      TAGTGGCGCCTCCACTGTATGTACTGACCACCCAGACACTGGACAAAGATCAGGGCATT 824
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 450      TAGTGGCGCCTCCACTGTATGTACTGACCACCCAGACACTGGACAAAGATCAGGGCATT 391
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 825      GCGTTTTAACCGATGCGGTCAAAGCGTGCACTGCGGAAATCGAAAAACATAAGGGCAAAC 884
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 390      GCGTTTTAACCGATGCGGTCAAAGCGTGCACTGCGGAAATCGAAAAACATAAGGGCAAAC 331
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 885      TGGTGGTGAAAGAAGCCCCACGCGCGGTGTGAGAACGTGAAGATAAACTGCTGAATGCC 944
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 330      TGGTGGTGAAAGAAGCCCCACGCGCGGTGTGAGAACGTGAAGATAAACTGCTGAATGCC 271
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 945      AACTGGATACTCTGGTAGAACAAAATGCGGAAGTCGCAGGCGACGATGATAGCGAGGACG 1004
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 270      AACTGGATACTCTGGTAGAACAAAATGCGGAAGTCGCAGGCGACGATGATAGCGAGGACG 211
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 1005     AAGAGGATACCGGCATGGGCGACATCGATCTGACCAATAGCGGCGTGCATGCGGATTAAT 1064
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 210      AAGAGGATACCGGCATGGGCGACATCGATCTGACCAATAGCGGCGTGCATGCGGATTAAT 151
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 1065     AAGGATCCTACTAGTAGCGGCCGCTGCAG 1093
                ||||||||||||||||||||||||||||||||||
Sbjct 150      AAGGATCCTACTAGTAGCGGCCGCTGCAG 122

```

### Sequence 9: Complete eIF2 $\beta$ in BioBrick (VF2)

```

Query 40      CCATATGGCGGACGAAGAACAGATGGAACGTAAGGAAGAAGCGACCGAGATTGCGCCGTT 99
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 103      CCATATGGCGGACGAAGAACAGATGGAACGTAAGGAAGAAGCGACCGAGATTGCGCCGTT 162
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 100     TGACCCGACTTaagaagaagaagaagaagaagTGGTGATTCAGGACCCAGCGGATGAAGT 159
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 163      TGACCCGACTAAGAAGAAGAAGAAGAAGAAAGTGGTGATTCAGGACCCAGCGGATGAAGT 222
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 160     GGATAAATTGGCCGAAAAGACCGAAGGCTGAGCGTGACCGAAAAGCGGCGAGGCGAGCTT 219
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 223      GGATAAATTGGCCGAAAAGACCGAAGGCTGAGCGTGACCGAAAAGCGGCGAGGCGAGCTT 282
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 220     TGTGGGCTTGAAGAAAAAGAAAAAGAAGTTAGTAGAACTGGACCCAGCCTTGTTCGAAGC 279
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 283      TGTGGGCTTGAAGAAAAAGAAAAAGAAGTTAGTAGAACTGGACCCAGCCTTGTTCGAAGC 342
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 280     CGGTGATGGCGAAGATACCTGGACGATCAGGTGGGCGAGGATGAACAAGGAGAGGGCAT 339
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 343      CGGTGATGGCGAAGATACCTGGACGATCAGGTGGGCGAGGATGAACAAGGAGAGGGCAT 402
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 340     TGTGCTGGGCGGTGCCACCCAGTATCCGTGGGAAGGAACCGATCGTGATTACAAATATGA 399
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 403      TGTGCTGGGCGGTGCCACCCAGTATCCGTGGGAAGGAACCGATCGTGATTACAAATATGA 462
                ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Query 400     TGAGCTGTTGGGCCGTGTGTTCAATATTCTGCGCGAGAATAATCCAGACCTGGCGGAGA 459

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|       |     |   |     |
|-------|-----|---|-----|
| Sbjct | 463 | <br>TGAGCTGTTGGGCCGTGTGTTCAATATTCTGCGCGAGAATAATCCAGACCTGGCGGGGAGA | 522 |
| Query | 460 | CCGTCGACGTACCGTGATGCGTCCACCGCAGGTCTTGCGTGAAGGCACCaaaaaaCCGT       | 519 |
| Sbjct | 523 | <br>CCGTCGACGTACCGTGATGCGTCCACCGCAGGTCTTGCGTGAAGGCACCAAAAAAACCGT  | 582 |
| Query | 520 | GTTTGTGAATTTTCATGGATCTGTGCAAAACCATGCATCGTCAGCCGGAACATGTGATGAT     | 579 |
| Sbjct | 583 | <br>GTTTGTGAATTTTCATGGATCTGTGCAAAACCATGCATCGTCAGCCGGAACATGTGATGAT | 642 |
| Query | 580 | GTTTCTGCTTGCAGAAATGGGCACCGCAGCCTGGATGGGCAGCAGCGTCTGGTCAT          | 639 |
| Sbjct | 643 | <br>GTTTCTGCTTGCAGAAATGGGCACCGCAGCCTGGATGGGCAGCAGCGTCTGGTCAT      | 702 |
| Query | 640 | TAAAGGCCGCTTCGCTCCCAAGAAATTTGAAGCGATCCTGCGGCGTTACATCAATGAATA      | 699 |
| Sbjct | 703 | <br>TAAAGGCCGCTTCGCTCCCAAGAAATTTGAAGCGATCCTGCGGCGTTACATCAATGAATA  | 762 |
| Query | 700 | TGTGATTGCAATGGCTGCAAGAGCCCGGATACCATTCTGAGCAAAGAGAATCGTCTGTT       | 759 |
| Sbjct | 763 | <br>TGTGATTGCAATGGCTGCAAGAGCCCGGATACCATTCTGAGCAAAGAGAATCGTCTGTT   | 822 |
| Query | 760 | TTTCTGCGTTGCGAACAGTGCAGCAGTAGCCGTAGCGTGGCGCCGATTAAGCCGGG-T        | 818 |
| Sbjct | 823 | <br>TTTCTGCGTTGCGAACAGTGCAGCAGTAGCCGTAGCGTGGCGCCNATTAAGCCNGGNT    | 882 |
| Query | 819 | TCGTGCGCCAGGTAGCCGTCGTAAGCGGGCACCTAATAAGGATCCTACTAGTAGCGGC        | 878 |
| Sbjct | 883 | <br>TCGTGCGCCAGGTAGCCGTCGTAAGCGGGCACCTAATAAGGATCCTACTAGTAGCGGN    | 942 |
| Query | 879 | CGCTGCAG 886  |     |
| Sbjct | 943 | <br>CGCTGCAG 950  |     |

### Sequence 10: Complete eIF2 $\beta$ in BioBrick (VR)

|       |     |   |     |
|-------|-----|---|-----|
| Query | 40  | CCATATGGCCGACGAAGAACAGATGGAACGTAAGGAAGAAGCGACCGAGATTGCGCCGTT      | 99  |
| Sbjct | 970 | <br>CCATATGGCCGACGANGAACAGATGGAACGTAAGGAAGAAGCGACCGAGATTGCGCCGTT  | 911 |
| Query | 100 | TGACCCGACTaagaagaagaagaagaagaagTGGTGATTTCAGGACCCAGCGGATGAAGT      | 159 |
| Sbjct | 910 | <br>TGACCCGACTAAGAAGAAGAAGAAGAAGTGGTGATTTCAGGACCCAGCGGATGAAGT     | 851 |
| Query | 160 | GGATAAATTGGCCGAAAAGACCGAAGCCTGAGCGTGACCGAAAAGCGGCGAGGCGAGCTT      | 219 |
| Sbjct | 850 | <br>GGATAAATTGGCCGAAAAGACNGAAGCCTGAGCGTGACCGAAAAGCGGCGAGGCGAGCTT  | 791 |
| Query | 220 | TGTGGGCTTGAAGAAAAAGAAAAAGAAGTTAGTAGAACTGGACCCAGCCTTGTCTGAAGC      | 279 |
| Sbjct | 790 | <br>TGTGGGCTTGAAGAAAAAGAAAAAGAAGTTAGTAGAACTGGACCCAGCCTTGTCTGAAGC  | 731 |
| Query | 280 | CGGTGATGGCGAAGATAACCCTGGACGATCAGGTGGGCGAGGATGAACAAGGAGAGGGCAT     | 339 |
| Sbjct | 730 | <br>CGGTGATGGCGAAGATAACCCTGGACGATCAGGTGGGCGAGGATGAACAAGGAGAGGGCAT | 671 |
| Query | 340 | TGTGCTGGGCGGTGCCACCCAGTATCCGTGGGAAGGAACCGATCGTGATTACAAATATGA      | 399 |
| Sbjct | 670 | <br>TGTGCTGGGCGGTGCCACCCAGTATCCGTGGGAAGGAACCGATCGTGATTACAAATATGA  | 611 |
| Query | 400 | TGAGCTGTTGGGCCGTGTGTTCAATATTCTGCGCGAGAATAATCCAGACCTGGCGGGGAGA     | 459 |
| Sbjct | 610 | <br>TGAGCTGTTGGGCCGTGTGTTCAATATTCTGCGCGAGAATAATCCAGACCTGGCGGGGAGA | 551 |
| Query | 460 | CCGTCGACGTACCGTGATGCGTCCACCGCAGGTCTTGCGTGAAGGCACCaaaaaaCCGT       | 519 |
| Sbjct | 550 | <br>CCGTCGACGTACCGTGATGCGTCCACCGCAGGTCTTGCGTGAAGGCACCAAAAAAACCGT  | 491 |

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Query 520  GTTTGTGAATTTTCATGGATCTGTGCAAAACCATGCATCGTCAGCCGGAACATGTGATGAT 579
          |||
Sbjct 490  GTTTGTGAATTTTCATGGATCTGTGCAAAACCATGCATCGTCAGCCGGAACATGTGATGAT 431

Query 580  GTTTCTGCTTGCAGAAATGGGCACCGAGCGGCAGCCTGGATGGGCAGCAGCGTCTGGTCAT 639
          |||
Sbjct 430  GTTTCTGCTTGCAGAAATGGGCACCGAGCGGCAGCCTGGATGGGCAGCAGCGTCTGGTCAT 371

Query 640  TAAAGCCGCTTCGCTCCCAAGAATTTTGAAGCGATCCTGCGGCGTTACATCAATGAATA 699
          |||
Sbjct 370  TAAAGCCGCTTCGCTCCCAAGAATTTTGAAGCGATCCTGCGGCGTTACATCAATGAATA 311

Query 700  TGTGATTTGCAATGGCTGCAAGAGCCCGGATACCATTCTGAGCAAAGAGAATCGTCTGTT 759
          |||
Sbjct 310  TGTGATTTGCAATGGCTGCAAGAGCCCGGATACCATTCTGAGCAAAGAGAATCGTCTGTT 251

Query 760  TTTCTGCGTTGCGAACAGTGCAGCAGTAGCCGTAGCGTGGCGCCGATTAAAGCCGGGTT 819
          |||
Sbjct 250  TTTCTGCGTTGCGAACAGTGCAGCAGTAGCCGTAGCGTGGCGCCGATTAAAGCCGGGTT 191

Query 820  CGTTGCCAGGTAGGCCGTCGTAAGCGGGCACCTAATAAGGATCCTACTAGTAGCGGCC 879
          |||
Sbjct 190  CGTTGCCAGGTAGGCCGTCGTAAGCGGGCACCTAATAAGGATCCTACTAGTAGCGGCC 131

Query 880  GCTGCAG 886
          |||
Sbjct 130  GCTGCAG 124

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### Sequence 11: Complete eIF2 $\gamma$ in BioBrick (VF2)

```

Query 866  CTTTACCAGCCCACCGAATATGATTGTTATTCGAAGCTTCGATGTGAATAAACCGGGCAG 925
          |||
Sbjct 824  CTTTACCAGCCCACCGAATATGATTGTTATTCGAAGCTTCGATGTGAATAAACCGGGCAG 765

Query 926  TGAAGTGGATGAAATTCGTGGCGCGCTGGCTGGGGGTTCCATTTTACGTGGCGTGTGCG 985
          |||
Sbjct 764  TGAAGTGGATGAAATTCGTGGCGCGCTGGCTGGGGGTTCCATTTTACGTGGCGTGTGCG 705

Query 986  GGTGAACCAGAACATTGAAGTGCCTCCGGGCATTGTGATGAAAGACGAATCGGGGAACAT 1045
          |||
Sbjct 704  GGTGAACCAGAACATTGAAGTGCCTCCGGGCATTGTGATGAAAGACGAATCGGGGAACAT 645

Query 1046 AAAGTGCACGCCCATTTATAGCCGATTGTGAGCTTATATGCGGAACAAAATGAGCTGCA 1105
          |||
Sbjct 644  AAAGTGCACGCCCATTTATAGCCGATTGTGAGCTTATATGCGGAACAAAATGAGCTGCA 585

Query 1106 ATTCGCGGTGCTGGGGCCGATTGGGGTGGGCACGACTATGGACCCGACCCTGACTCG 1165
          |||
Sbjct 584  ATTCGCGGTGCTGGGGCCGATTGGGGTGGGCACGACTATGGACCCGACCCTGACTCG 525

Query 1166 TGCGGATCGTCTGGTGGGCCAAGTGTGGGAGAAATTTGGTAGCCTGCCGGATGCTTTGT 1225
          |||
Sbjct 524  TGCGGATCGTCTGGTGGGCCAAGTGTGGGAGAAATTTGGTAGCCTGCCGGATGCTTTGT 465

Query 1226 TGAATTAGAGATTAACCTTTTTCAGCTGCGTCTGCTGGGTGTTTCGGACCAAAGGGAC 1285
          |||
Sbjct 464  TGAATTAGAGATTAACCTTTTTCAGCTGCGTCTGCTGGGTGTTTCGGACCAAAGGGAC 405

Query 1286 CGAGAAAGCGGGGAAAGTGAAGCAAAATGACCAAGGGGAAATTCGATGTTAAACATCGG 1345
          |||
Sbjct 404  CGAGAAAGCGGGGAAAGTGAAGCAAAATGACCAAGGGGAAATTCGATGTTAAACATCGG 345

Query 1346 AAGCATGAGCACCGGAGCGCGTGTGGTTGCGGTCAAAAATGACCTGGCGAAGTTACAGTT 1405
          |||
Sbjct 344  AAGCATGAGCACCGGAGCGCGTGTGGTTGCGGTCAAAAATGACCTGGCGAAGTTACAGTT 285

Query 1406 AACAGCCCCAGTATGTACTTCGAAAGGCGAGAAGGTAGCGCTGTCCCGTGTGTGAAAA 1465
          |||
Sbjct 284  AACAGCCCCAGTATGTACTTCGAAAGGCGAGAAGGTAGCGCTGTCCCGTGTGTGAAAA 225

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Query 1466 GCATTGGCGTCTGATCGGTTGGGGCCAGATTCAAGCTGGAGCTACCCCTGGAAGTGCCGCC 1525  
 |||  
 Sbjct 224 GCATTGGCGTCTGATCGGTTGGGGCCAGATTCAAGCTGGAGCTACCCCTGGAAGTGCCGCC 165  
  
 Query 1526 GTGCCCCTGTAAATAAGGATCCTACTAGTAGCGGCCGCTGCAG 1568  
 |||  
 Sbjct 164 GTGCCCCTGTAAATAAGGATCCTACTAGTAGCGGCCGCTGCAG 122  
  
 Query 496 TGCGGCAAATGAATCTTGCCCTCAGCCGACAGCCAGCGAACATCTGGCTGCGGTGGAAA 554  
 |||  
 Sbjct 1088 TGCGGCAAANGAAT-NTG-CCTCAGCNNNCNGACCAGCGANCATCNGGCTGCGGTGG-AA 1032  
  
 Query 555 TTATGCGTCTTCAGCACCTGATAATTCTTCAGAATAAGATTGATCTGATTAGGAATCCG 614  
 |||  
 Sbjct 1031 NTATGCGT-NTCAGCNNNNGAT-NNNNNTCAGAATAAGA-TGATNTGATTAGGAATCCG 975  
  
 Query 615 CAGCGATGAATCAACATGAGGCGATTCAAAAGTTCATTCAAGGCACCATAGCGGAGGGCG 674  
 |||  
 Sbjct 974 CAGCGATGAATCAACATGAGGCGATTCAAAAGTTCATTCAAGGCACCATAGCGGAGGGCG 915  
  
 Query 675 CTCCAGTGGTCCGATTAGCGCGCAGTTGAAGTATAACATCGACGTAATTTGCGAATACA 734  
 |||  
 Sbjct 914 CTCCAG-NGNTCCGATTAGCGCGCAGTTGAAGTATAACATCGACGTAATTTGCGAATACA 856  
  
 Query 735 TCATAAAGAAGATTCCGATCCCTGAACGTAACCTTACCAGCCCACCGAATATGATTGTTA 794  
 |||  
 Sbjct 855 TCATAAAGAAGATTCCGATCCCTGAACGTAACCTTACCAGCCCACCGAATATGATTGTTA 796  
  
 Query 795 TTCGAAGCTTCGATGTGAATAAACCGGGCAGTGAAGTGGATGAAATTCGTGGCGGCGTGG 854  
 |||  
 Sbjct 795 TTCGAAGCTTCGATGTGAATAAACCGGGCAGTGAAGTGGATGAAATTCGTGGCGGCGTGG 736  
  
 Query 855 CTGGGGTTCCCTTT 869  
 |||  
 Sbjct 735 CTGGGGTTCCATT 721

## Sequence 12: Complete eIF2 $\alpha$ in BioBrick (VF2)

Query 1 GAATTCGCGGCCGCTTCTAGAGATTAAAGAGGAGAAATACCATATGGCGAACCTGGAATG 60  
 |||  
 Sbjct 86 GAATTCGCGGCCGCTTCTAGAGATTAAAGAGGAGAAATACCATATGGCGAACCTGGAATG 145  
  
 Query 61 CCGTATGTATGAACCGCGTTTCCCGAAGTGGATGCTGCGGTGATGATTCAGGTGAAACA 120  
 |||  
 Sbjct 146 CCGTATGTATGAACCGCGTTTCCCGAAGTGGATGCTGCGGTGATGATTCAGGTGAAACA 205  
  
 Query 121 TATTGCGGATATGGGCGCGTATGTGAGCCTGCTGGAATATAACAATGTGGAAGGCATGAT 180  
 |||  
 Sbjct 206 TATTGCGGATATGGGCGCGTATGTGAGCCTGCTGGAATATAACAATGTGGAAGGCATGAT 265  
  
 Query 181 TCTGTTTTCCGAGCTGAGCCGTCGCCGATTCCGAGCATAAGCAGCCTGATAAAAAGTGGG 240  
 |||  
 Sbjct 266 TCTGTTTTCCGAGCTGAGCCGTCGCCGATTCCGAGCATAAGCAGCCTGATAAAAAGTGGG 325  
  
 Query 241 TCGTCAAGAACCGCGATAGTCTTCTGCTGTGGATCGTGATAAAGGCTATATTGACTTGTC 300  
 |||  
 Sbjct 326 TCGTCAAGAACCGCGATAGTCTTCTGCTGTGGATCGTGATAAAGGCTATATTGACTTGTC 385  
  
 Query 301 CAAGCGCCGTGTTTCCGAAGAAGAAGCGCGTAGCTGCGAGGATAAATACAATAAGAGCAA 360  
 |||  
 Sbjct 386 CAAGCGCCGTGTTTCCGAAGAAGAAGCGCGTAGCTGCGAGGATAAATACAATAAGAGCAA 445  
  
 Query 361 GCTGGTCCATAGCATTATGCGTCACGTTGCTGAGACCCTGGAGATTGATCTGGAACCGAT 420  
 |||  
 Sbjct 446 GCTGGTCCATAGCATTATGCGTCACGTTGCTGAGACCCTGGAGATTGATCTGGAACCGAT 505  
  
 Query 421 CTATCAGCGGATTGGCTGGCCGCTGTATCGTAAATACGGCCACGCATTTGAAGCGTTCAA 480

|       |     |   |     |
|-------|-----|---|-----|
| Sbjct | 506 | <br>CTATCAGCGGATTGGCTGGCCGCTGTATCGTAAATACGGCCACGCATTTGAAGCGTTCAA  | 565 |
| Query | 481 | GCTGATTGTTGCGGACCCTGACGCCATATTAGACGTGCTGACCTATGAAGAGCGTGAAAC      | 540 |
| Sbjct | 566 | <br>GCTGATTGTTGCGGACCCTGACGCCATATTAGACGTGCTGACCTATGAAGAGCGTGAAAC  | 625 |
| Query | 541 | CGGGCCAGATGGCCAAGAAGTGACCAAAGTGGTACCAGCAGTCACCCCGGAAATAAAGGA      | 600 |
| Sbjct | 626 | <br>CGGGCCAGATGGCCAAGAAGTGACCAAAGTGGTACCAGCAGTCACCCCGGAAATAAAGGA  | 685 |
| Query | 601 | AACCCTGGTGCAGAACATTCGTCGTCGGATGACACCACAACCCTGAAAATTCGTGCGGA       | 660 |
| Sbjct | 686 | <br>AACCCTGGTGCAGAACATTCGTCGTCGGATGACACCACAACCCTGAAAATTCGTGCGGA   | 745 |
| Query | 661 | CGTGGAATGAAATGCTTTTCAGTTTGTATGGCGTGCTGCATATCAAACAGCGGATGCGTAA     | 720 |
| Sbjct | 746 | <br>CGTGGAATGAAATGCTTTTCAGTTTGTATGGCGTGCTGCATATCAAACAGCGGATGCGTAA | 805 |
| Query | 721 | GGCGGAGGCTGCGGGCAATACCAATTGTCGGTAAAGATTAAGTTAGTGGCGCCTCCACT       | 780 |
| Sbjct | 806 | <br>GGCGGAGNNTGCGGGCAATACCAATTGTCGGTAAAGATTAAGTTAGTGGCGCCTCCACT   | 865 |
| Query | 781 | GTATGTAAGTACCACCCAGACACTGGACAAAGATCAGGGCATTAGCGTTTTAACCAGATGC     | 840 |
| Sbjct | 866 | <br>GTATGTAAGTACCACCCAGACACTGGACAAAGATCAGGGCATTAGCGTTTTAACCAGATGC | 924 |
| Query | 841 | GGTCAAAGCGTGCACCTGCGGAAATCGAAAAACATAAGGGCAAACCTGGTGGTGAA          | 894 |
| Sbjct | 925 | <br>GGTCAAAGCGTGCACCTGCGGAAATCGAAAAACATAAGG-CAA-CTGGTGGTGAA       | 976 |

### Sequence 13: Complete eIF2 $\alpha$ in BioBrick (VR)

|       |      |  |      |
|-------|------|--|------|
| Query | 1883 | CTTTACCAGCCCACCGAATATGATTGTTATTCGAAGCTTCGATGTGAATAAACCGGGCAG     | 1942 |
| Sbjct | 821  | <br>CTTTACCAGCCCACCGAATATGATTGTTATTCGAAGCTTCGATGTGAATAAACCGGGCAG | 762  |
| Query | 1943 | TGAAGTGGATGAAATTCGTGGCGCGTGGCTGGGGTTCCATTTTACGTGGCGTGTGCG        | 2002 |
| Sbjct | 761  | <br>TGAAGTGGATGAAATTCGTGGCGCGTGGCTGGGGTTCCATTTTACGTGGCGTGTGCG    | 702  |
| Query | 2003 | GGTGAACCAGAACATTGAAGTGCCTCCGGGCATTGTGATGAAAGACGAATCGGGGAACAT     | 2062 |
| Sbjct | 701  | <br>GGTGAACCAGAACATTGAAGTGCCTCCGGGCATTGTGATGAAAGACGAATCGGGGAACAT | 642  |
| Query | 2063 | AAAGTGCACGCCCATTTATAGCCGTATTGAGCTTATATGCGGAACAAAATGAGCTGCA       | 2122 |
| Sbjct | 641  | <br>AAAGTGCACGCCCATTTATAGCCGTATTGAGCTTATATGCGGAACAAAATGAGCTGCA   | 582  |
| Query | 2123 | ATTCGCGGTGCTGGGGCCATGATTGGGGTGGGCACGACTATGGACCCGACCCTGACTCG      | 2182 |
| Sbjct | 581  | <br>ATTCGCGGTGCTGGGGCCATGATTGGGGTGGGCACGACTATGGACCCGACCCTGACTCG  | 522  |
| Query | 2183 | TGCGGATCGTCTGGTGGGCCAAGTGTGGGAGAAAATGGTAGCCTGCCGATGTCCTTGT       | 2242 |
| Sbjct | 521  | <br>TGCGGATCGTCTGGTGGGCCAAGTGTGGGAGAAAATGGTAGCCTGCCGATGTCCTTGT   | 462  |
| Query | 2243 | TGAATTAGAGATTAACCTTTTTCAGCTGCGTCTGCTGCTGGGTGTTTCGACCAAAGGGAC     | 2302 |
| Sbjct | 461  | <br>TGAATTAGAGATTAACCTTTTTCAGCTGCGTCTGCTGCTGGGTGTTTCGACCAAAGGGAC | 402  |
| Query | 2303 | CGAGAAAGCGGGGAAAGTGAAGCAAAATGACCAAGGGGAAATTCGATGTTAAACATCGG      | 2362 |
| Sbjct | 401  | <br>CGAGAAAGCGGGGAAAGTGAAGCAAAATGACCAAGGGGAAATTCGATGTTAAACATCGG  | 342  |
| Query | 2363 | AAGCATGAGCACCGGAGCGCGTGTGGTTGCGGTCAAAAATGACCTGGCGAAGTTACAGTT     | 2422 |
| Sbjct | 341  | <br>AAGCATGAGCACCGGAGCGCGTGTGGTTGCGGTCAAAAATGACCTGGCGAAGTTACAGTT | 282  |
| Query | 2423 | AACAGCCCCAGTATGTAAGTTCGAAAGGCGAGAAGGTAGCGCTGTCCCGTGTGGAAAA       | 2482 |

|       |      |  |      |
|-------|------|--|------|
| Sbjct | 281  | <br>AACAGCCCCAGTATGTACTTCGAAAGGCGAGAAAGGTAGCGCTGTCCCGTGTGGAAAA     | 222  |
| Query | 2483 | GCATTGGCGTCTGATCGGTTGGGGCCAGATTCAAGCTGGAGCTACCCTGGAAGTCCGCC<br>    | 2542 |
| Sbjct | 221  | GCATTGGCGTCTGATCGGTTGGGGCCAGATTCAAGCTGGAGCTACCCTGGAAGTCCGCC        | 162  |
| Query | 2543 | GTGCCCGCTGTAATAAGGATCCTACTAGTAGCGGCCGCTGCAG 2585<br>               |      |
| Sbjct | 161  | GTGCCCGCTGTAATAAGGATCCTACTAGTAGCGGCCGCTGCAG 119                    |      |
| Query | 1612 | TGATCTGATTCAAGAAATCCGAGCGATGAATCAACATGAGGCGATTCAAAAAGTTTCATTCA<br> | 1671 |
| Sbjct | 988  | TGATNTGA-TCAGGAATCNGCAGCGATGAAT-NACATGNNCGATTCAAAAAGTTTCATTCA      | 931  |
| Query | 1672 | AGGCACCATAGCGGAGGGCGCTCCAGTGGTTCCGATTAGCGCGCAGTTGAAGTATAACAT<br>   | 1731 |
| Sbjct | 930  | AGGCNCCATAGCGGAGGGCGNCCAGTGGTTNCGATTAGCGCGCAGTTGAAGTATAACAT        | 871  |
| Query | 1732 | CGACGTAATTTGCGAATACATCATAAAGAAGATTCCGATCCCTGAACGTAACCTTACCAG<br>   | 1791 |
| Sbjct | 870  | CGACGT-ANTTGCGAATACATCATAAAGAAGATTCCGA-NNCNGAACGTAACCTTACCAG       | 813  |
| Query | 1792 | CCCACCGAATATGATTGTTATTTCGAAGCTTCGATGTGAATAAACCGGGCAGTGAAGTGGA<br>  | 1851 |
| Sbjct | 812  | CCCACCGAATATGATTGTTATTTCGAAGCTTCGATGTGAATAAACCGGGCAGTGAAGTGGA      | 753  |
| Query | 1852 | TGAAATTCGTGGCGCGTGGCTGGGGTTCCCTTT 1886<br>                         |      |
| Sbjct | 752  | TGAAATTCGTGGCGCGTGGCTGGGGTTCCATTT 718                              |      |

#### Sequence 14: Complete eIF2 $\beta\gamma$ in BioBrick (VF2)

|       |      |  |      |
|-------|------|--|------|
| Query | 2592 | CCATATGGCGGACGAAGAACAGATGGAACTAAGGAAGAAGCGACCGAGATTGCGCCGTT<br>    | 2651 |
| Sbjct | 97   | CCATATGGCGGACGAAGAACAGATGGAACTAAGGAAGAAGCGACCGAGATTGCGCCGTT        | 156  |
| Query | 2652 | TGACCCGACTaagaagaagaagaagaagaagTGGTGATTCAAGACCCAGCGGATGAAGT<br>    | 2711 |
| Sbjct | 157  | TGACCCGACTAAGAAGAAGAAGAAGAAGAAGTGGTGATTNNNGACCCANNGGATGAAGT        | 216  |
| Query | 2712 | GGATAAATTTGGCCGAAAAGACCGAAGGCCTGAGCGTGACCGAAAAGCGGCGAGGCGAGCTT<br> | 2771 |
| Sbjct | 217  | GGATAAATTTGGCCGAAAAGANNGAAGGNCTGANNGTGACCGAAAAGNGGAGGNGAGNTT       | 276  |
| Query | 2772 | TGTGGGCTTGAAGAAAAAGAAAAAGAAGTTAGTAGAACTGGACCCAGCCTTGTGCGAAGC<br>   | 2831 |
| Sbjct | 277  | TGTGGGCTTGAAGAAAAAGAAAAAGAAGTTAGTAGAANTGGACCCCNCTTGTGCGAAGC        | 336  |
| Query | 2832 | CGGTGATGGCGAAGATACCCTGGACGATCAGGTGGGCGAGGATGAACAAGGAGAGGGCAT<br>   | 2891 |
| Sbjct | 337  | CGGTGATGGCGAAGATACNCTGGACGATCNGGTGGGCGAGGATGAACAAGGAGAGGGCAT       | 396  |
| Query | 2892 | TGTGCTGGGCGGTGCCACCCAGTATCCGTGGGAAGGAACCGATCGTGATTACAAAATATGA<br>  | 2951 |
| Sbjct | 397  | TGTGCTGGGCGGTGCCNCCAGTATCCGTGGGAAGGAACCGATCGTGATTACNAATATGA        | 456  |
| Query | 2952 | TGAGCTGTGGGCGGTGTGTCAATATTCTGCGGAGAATAATCCAGACCTGGGCGGAGA<br>      | 3011 |
| Sbjct | 457  | TGAGCTGTGGGCGGTGTGTCAATATTCTGCGGAGAATAATCCAGACCTGGGCGGAGA          | 516  |
| Query | 3012 | CCGTCGACGTACCGTGTGCGTCCACCGCAGGTCTGCGTGAAGGCACCaaaaaaCCGT<br>      | 3071 |
| Sbjct | 517  | CCGTCNACGTACCGTGTGCGTCCNCCGAGGTCTGCGTGAAGGCACCNAAAAAACCGT          | 576  |
| Query | 3072 | GTTTGTGAATTTTCATGGATCTGTGCAAAACCATGCATCGTCAGCCGGAACATGTGATGAT<br>  | 3131 |
| Sbjct | 577  | GTTTGTGAATTTTCATGGATCTGTGCAAAACCATGCNTCGTCNGCCGGAACATGTGATGAT      | 636  |
| Query | 3132 | GTTTCTGCTTGGCGAAATGGGCACACGCGCAGCCTGGATGGGCGAGCGCTGGTGCAT          | 3191 |

|       |      |  |      |
|-------|------|--|------|
| Sbjct | 637  | <br>GTTTCTGCTTGCAGAAATGGGCACCAGCGCNGCCTGGATGGGCAGCAGCNTCTGGTCNT  | 696  |
| Query | 3192 | TAAAGGCCGCTTCGCTCCCAAGAATTTGAAGCGATCCTGCGGCGTTACATCAATGAATA      | 3251 |
| Sbjct | 697  | <br>TAAAGGCCNCTTCNCTCCCNAGAATTTGAAGCGATCCTGCNGCGTTACNTCNATGAATA  | 756  |
| Query | 3252 | TGTGATTTGCAATGGCTGCAAGAGCCCGGATAC-CATTCTGAGCAAAGAGAATCGTCTGT     | 3310 |
| Sbjct | 757  | <br>TGTGATTTGCAATGGCTGCNAGAGCCCGGATACNCNTTCTGAGCNAAGAGAATCNTCTGT | 816  |
| Query | 3311 | TTTTCTGCGTTGCGAACAGTGC CGCAGTAGCCGTAGCGTGGCGCCGATTAAGGCCG-GG     | 3369 |
| Sbjct | 817  | TTTTCTGCGTTGCNAACAGTGC CGCAGTAGCCGTAGCGTGGNNNNNATTAAGNCNNGG      | 876  |
| Query | 3370 | TTCGTGCCCAGGTAGGCCGTCGTAAGGCGGGCACCTAATA 3410                    |      |
| Sbjct | 877  | <br>TTCGTGCCCNGGTAGGNCNTNNTAAGNGGGCNCCTAATA 917                  |      |

### Sequence 15 Complete eIF2 $\beta$ in BioBrick (VR)

|       |      |   |      |
|-------|------|---|------|
| Query | 1883 | CTTTACCAGCCACCAGAAATGATTGTTATTCGAAGCTTCGATGTGAATAAACCGGGCAG       | 1942 |
| Sbjct | 826  | <br>CTTTACCANCCACNGAATAATGATTGTTATNGAAGCTTNGANGAATAAACCGGGCAG     | 767  |
| Query | 1943 | TGAAGTGGATGAAATTCGTGGCGCGTGGCTGGGGTTCCATTTTACGTGGCGTGTGCG         | 2002 |
| Sbjct | 766  | <br>TGAAGNGGANGAATAATNGNGCGGNGGCTGGGGTTCCATTTTACGTGGCGTGTGNG      | 707  |
| Query | 2003 | GGTGAACCAGAACATTGAAGTGCCTCCGGGCATGTGATGAAAGACGAATCGGGGAACAT       | 2062 |
| Sbjct | 706  | <br>GGTGAACCAGAACATNGAAGNGCGTCCGGGCATGTGATGAAAGACGAATCGGGGAACAT   | 647  |
| Query | 2063 | AAAGTGACAGCCCATTTATAGCCGATTGTGAGCTTATATGCGGAACAAAATGAGCTGCA       | 2122 |
| Sbjct | 646  | <br>AAAGTGACAGCCCATTTATAGCCGATTGTGAGCTTATATGCGGAACAAAATGAGCTGCA   | 587  |
| Query | 2123 | ATTCGGGTGCTTGGGGCCGATTGGGGTGGGCACGACTATGGACCCGACCCTGACTCG         | 2182 |
| Sbjct | 586  | <br>ATTCGGGTGCTTGGGGCCGATTGGGGTGGGCACGACTATGGACCCGACCCTGACTCG     | 527  |
| Query | 2183 | TGCGGATCGTCTGGTGGGCCAAGTGTCTGGGAGAAATT-GGTAGCCTGCGGATGTCCTTTG     | 2241 |
| Sbjct | 526  | <br>TGCGGATCGTCTGGTGGGCCAAGTGTCTGGGAGAAATTNGGTAGCCTGCNNNANGTCTTTN | 467  |
| Query | 2242 | TTGAATTAGAGATTAACCTTTTTTCAGCTCGCTCGTCTGCTGGGTGTTCCGACCAAAGGGA     | 2301 |
| Sbjct | 466  | <br>NNNAATTANNGATTAACCTTTTTTCANNTNNNTCGTCTNNNNGNNNTTCGNACCAAAGGNA | 407  |
| Query | 2302 | CCGAGAAAGCGGGGAAAGTGAGCAAATTGACCAAGGGGAAATTTCTGATGTTAAACATCG      | 2361 |
| Sbjct | 406  | <br>CCNAGAAAGCGGNGAAANNANCAAATTNACCAAGGGGAAATTTNANNTTAAACATCG     | 347  |
| Query | 2362 | GAAGCATGAGCACCAGCGCGTGTGGTTGCGGTCAAAAAATGACCTGGCGAAGTTACAGT       | 2421 |
| Sbjct | 346  | <br>NAAGCANNAGCACCNNAGCGGNNTGGTTGCGGTCAAAAAATNACCTGNNGAAGTTACAGT  | 287  |
| Query | 2422 | TAACAGCCCCAGTATGTACTTCGAAAGGCGAGAAGGTAGCGCTGTCCCGTCGTGTGGAAA      | 2481 |
| Sbjct | 286  | <br>TAACAGCCCCAGTATGTACTTCGAAAGGCGAGAAGGTAGCGCTGTCCCGTCGTGTGGAAA  | 227  |
| Query | 2482 | AGCATTGGCGTCTGATCGGTTGGGGCCAGATTCAAGCTGGAGCTACCCTGGAAGTGCCGC      | 2541 |
| Sbjct | 226  | <br>AGCATTGGCGTCTNATCGNTTGGGGCCAGATTCAAGCTGGAGCTACCCTGGAAGTGCCGC  | 167  |
| Query | 2542 | CGTGCCCGCTGGAAT 2556  |      |
| Sbjct | 166  | <br>CGTNCCCGCTGTAAT 152   |      |

Query 3406 TAATAAGGATCCTACTAGTAGCGGCCGCTGCAG 3438  
 |||  
 Sbjct 155 TAATAAGGATCCTACTAGTAGCGGCCGCTGCAG 123

**Sequence 16: Complete eIF2 $\beta\gamma$  in pet22bb (T7pro)**

Query 2592 CCATATGGCGGACGAAGAACAGATGGAACTAAGGAAGAAGCGACCGAGATTGCGCCGTT 2651  
 |||  
 Sbjct 35 CCATATGGNGGACGAAGAACAGATGGAACTAAGGAAGAANNNNNNNNGATTGCGCCGTT 94

Query 2652 TGACCCGACTaagaagaagaagaagaagaagTGGTGATTCAGGACCCAGCGGATGAAGT 2711  
 |||  
 Sbjct 95 TGACCCGACTAAGAAGAAGAANAAGAAAGTGGTGATTCAGGACCCAGCGGATGAAGT 154

Query 2712 GGATAAATGGCCGAAAAGACCGAAGGCCTGAGCGTGACCGAAAGCGGCGAGGCGAGCTT 2771  
 |||  
 Sbjct 155 GGATAAATGGCCGATAAGACCGAAGGCCTGAGCGTGACCGAAAGCGGCGAGGCGAGCTT 214

Query 2772 TGTGGGCTTGAAGAAAAGAAAAGAAGTTAGTAGAACTGGACCCAGCCTTGTGCGAAGC 2831  
 |||  
 Sbjct 215 TGTGGGCTTGAAGAAAAGAAAAGAAGTTAGTAGAACTGGACCCAGCCTTGTGCGAAGC 274

Query 2832 CGGTGATGGCGAAGATACCCCTGGACGATCAGGTGGGCGAGGATGAACAAGGAGAGGGCAT 2891  
 |||  
 Sbjct 275 CGGTGATGGCGAAGATACCCCTGGACGATCAGGTGGGCGAGGATGAACAAGGAGAGGGCAT 334

Query 2892 TGTGCTGGGCGGTGCCACCCAGTATCCGTGGGAAGGAACCGATCGTGATTACAAATATGA 2951  
 |||  
 Sbjct 335 TGTGCTGGGCGGTGCCACCCAGTATCCGTGGGAAGGAACCGATCGTGATTACAAATATGA 394

Query 2952 TGAGCTGTGGGCCGTGTGTCAATATTCTGCGGAGAATAATCCAGACCTGGCGGGAGA 3011  
 |||  
 Sbjct 395 TGAGCTGTGGGCCGTGTGTCAATATTCTGCGGAGAATAATCCAAACCTGGCGGGAGA 454

Query 3012 CCGTGCACGTACCGTGATGCGTCCACCGCAGGTCTTGCCTGAAGGCACCaaaaaaCCGT 3071  
 |||  
 Sbjct 455 CCGTCCACGTACCGTGATGCGTCCACCGCAGGTCTTGCCTGAAGGCACCAAAAAAAAACCGT 514

Query 3072 GTTTGTGAATTTTCATGGATCTGTGCAAAACCATGCATCGTCAGCCGGAACATGTGATGAT 3131  
 |||  
 Sbjct 515 GTTTGTGAATTTTCATGGATCTGTGNANAANCNTGNNTNNNCNNCNGAACATGTGATGAN 574

Query 3132 GTTTCTGCTTGCGGAAATGGG 3152  
 |||  
 Sbjct 575 GTTTCTGNNTGCGGAAANGGG 595

**Sequence 17: Complete eIF2 $\beta\gamma$  in pet22bb (T7term)**

Query 1883 CTTTACCAGCCCACCGAATATGATTGTTATTCGAAGCTTCGATGTGAATAAACCGGGCAG 1942  
 |||  
 Sbjct 766 CTTTACCAGCCCACCGAATATGATTGTTATTCGAAGCTTCGATGTGAATAAACCGGGCAG 707

Query 1943 TGAAGTGGATGAAATTCGTGGCGCGGTGGCTGGGGTTCCATTTTACGTGGCGTGTGCG 2002  
 |||  
 Sbjct 706 TGAAGTGGATGAAATTCGTGGCGCGGTGGCTGGGGTTCCATTTTACGTGGCGTGTGCG 647

Query 2003 GGTGAACCAGAACATTGAAGTGCCTCCGGGCATTTGTGATGAAAGACGAATCGGGGAACAT 2062  
 |||  
 Sbjct 646 GGTGAACCAGAACATTGAAGTGCCTCCGGGCATTTGTGATGAAAGACGAATCGGGGAACAT 587

Query 2063 AAAGTGCACGCCCATTTATAGCCGATTGTGAGCTTATATGCGGAACAAAATGAGCTGCA 2122  
 |||  
 Sbjct 586 AAAGTGCACGCCCATTTATAGCCGATTGTGAGCTTATATGCGGAACAAAATGAGCTGCA 527

Query 2123 ATTCGCGGTGCTGGGGCCGTGATTGGGGTGGGCACGACTATGGACCCGACCCCTGACTCG 2182  
 |||  
 Sbjct 526 ATTCGCGGTGCTGGGGCCGTGATTGGGGTGGGCACGACTATGGACCCGACCCCTGACTCG 467

|       |      |  |      |
|-------|------|--|------|
| Query | 2183 | TGCGGATCGTCTGGTGGGCCAAGTCTGGGAGAAATTGGTAGCCTGCCGGATGTCTTTGT    | 2242 |
| Sbjct | 466  | TGCGGATCGTCTGGTGGGCCAAGTCTGGGAGAAATTGGTAGCCTGCCGGATGTCTTTGT    | 407  |
| Query | 2243 | TGAATTAGAGATTAACCTTTTTTTCAGCTGCGTCGCTGCTGGGTGTTTCGGACCAAAGGGAC | 2302 |
| Sbjct | 406  | TGAATTAGAGATTAACCTTTTTTTCAGCTGCGTCGCTGCTGGGTGTTTCGGACCAAAGGGAC | 347  |
| Query | 2303 | CGAGAAAGCGGGGAAAGTGAAGCAAAATGACCAAGGGGGAAATTCTGATGTTAAACATCGG  | 2362 |
| Sbjct | 346  | CGAGAAAGCGGGGAAAGTGAAGCAAAATGACCAAGGGGGAAATTCTGATGTTAAACATCGG  | 287  |
| Query | 2363 | AAGCATGAGCACCGGAGCGCGTGTGGTTGCGGTCAAAAATGACCTGGCGAAGTTACAGTT   | 2422 |
| Sbjct | 286  | AAGCATGAGCACCGGAGCGCGTGTGGTTGCGGTCAAAAATGACCTGGCGAAGTTACAGTT   | 227  |
| Query | 2423 | AACAGCCCCAGTATGTACTTCGAAAGGCGAGAAGGTAGCGCTGTCCCGTCGTGGAAAA     | 2482 |
| Sbjct | 226  | AACAGCCCCAGTATGTACTTCGAAAGGCGAGAAGGTAGCGCTGTCCCGTCGTGGAAAA     | 167  |
| Query | 2483 | GCATTGGCGTCTGATCGGTTGGGGCCAGATTCAAGCTGGAGCTACCCTGGAAGTGCCGCC   | 2542 |
| Sbjct | 166  | GCATTGGCGTCTGATCGGTTGGGGCCAGATTCAAGCTGGAGCTACCCTGGAAGTGCCGCC   | 107  |
| Query | 2543 | GTGCCCGCTGGAAT   | 2556 |
| Sbjct | 106  | GTGCCCGCTGTAAT   | 93   |
| Query | 1439 | TGCCCTGGTCATGATATTTTGATGGCGACCATGCTGAACGGAGCCGCTATTATGGATGGC   | 1498 |
| Sbjct | 1102 | TGCCCT-GTCNNGATNNTTTGATGGCGNCCATGCTGANC-GAGCCGNT-NTATGGATGNC   | 1046 |
| Query | 1499 | GCCCTGTTGCTGATTGCGGCAAAATGAATCTTGCCCTCAGCCGAGACCAGCGAACATCTG   | 1558 |
| Sbjct | 1045 | G-CCNGTTGCTG-NTGNNCAAANGAATC-T-NNNNCAGCCGAGACCAGCGAACATCTG     | 990  |
| Query | 1559 | GCTGCGGTGGAAATATGCGTCTTCAGCACCTGATAATTCTTCAGAATAAGATTGATCTG    | 1618 |
| Sbjct | 989  | GCTGCGGTGGAAATNATGCGTNTTCAGC-NCTGATAATTCTTCAGAATAAGATTGATCTG   | 931  |
| Query | 1619 | ATTCAGGAATCCGCAGCGATGAATCAACATGAGGCGATTCAAAAAGTTCATTCAAGGCACC  | 1678 |
| Sbjct | 930  | ATTCAGGAATCCGCAGCGATGAATCAACATGAGGCGATTCAAAAAGTTCATTCAAGGCACC  | 871  |
| Query | 1679 | ATAGCGGAGGGCGCTCCAGTGGTTCCGATTAGCGCGCAGTTGAAGTATAACATCGACGTA   | 1738 |
| Sbjct | 870  | ATAGCGGAGGGCGCTCCAGTGGTTCCGATTAGCGCGCAGTTGAAGTATAACATCGACGTA   | 811  |
| Query | 1739 | ATTTGCGAATACATCATAAAGAAGATTCGGATCCCTGAACGTAACCTTACCAGCCCACCG   | 1798 |
| Sbjct | 810  | ATTTGCGAATACATCATAAAGAAGATTCGGATCCCTGAACGTAACCTTACCAGCCCACCG   | 751  |
| Query | 1799 | AATATGATTGTTATTTCGAAGCTTCGATGTGAATAAACCGGGCAGTGAAGTGGATGAAATT  | 1858 |
| Sbjct | 750  | AATATGATTGTTATTTCGAAGCTTCGATGTGAATAAACCGGGCAGTGAAGTGGATGAAATT  | 691  |
| Query | 1859 | CGTGGCGGCGTGGCTGGGGGTTCCCTTT                                   | 1886 |
| Sbjct | 690  | CGTGGCGGCGTGGCTGGGGGTTCCATTT                                   | 663  |
| Query | 3406 | TAATAAGGATCCTACTAGTAGCGGCCGCTGCAG                              | 3438 |
| Sbjct | 96   | TAATAAGGATCCTACTAGTAGCGGCCGCTGCAG                              | 64   |